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<b>(21) International Application Number:</b> PCT/CA98/00544 <b>(22) International Filing Date:</b> 2 June 1998 (02.06.98) <b>(30) Priority Data:</b> 08/867,941 3 June 1997 (03.06.97) US 09/074,658 8 May 1998 (08.05.98) US <b>(71) Applicant (for all designated States except US):</b> CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Avenue West, North York, Ontario M2R 3T4 (CA). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> LOOSMORE, Sheena, M. [CA/CA]; 70 Crawford Rose Drive, Aurora, Ontario L4G 4R4 (CA). DU, Run-Pan [CA/CA]; (CA). WANG, Quijun [CA/CA]; 299 Chelwood Drive, Thornhill, Ontario L4J 7Y8 (CA). YANG, Yan-Ping [CA/CA]; Apartment 709, 120 Torresdale Avenue, Willowdale, Ontario M2R 3N7 (CA). KLEIN, Michel, H. [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA). <b>(74) Agent:</b> STEWART, Michael, I.; Sim & McBurney, 6th floor, 330 University Avenue, Toronto, Ontario M5G 1R7 (CA).		<b>(81) Designated States:</b> AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i>
<b>(54) Title:</b> LACTOFERRIN RECEPTOR GENES OF MORAXELLA  <b>(57) Abstract</b>  Purified and isolated nucleic acid molecules are provided which encode lactoferrin receptor proteins of <i>Moraxella</i> , such as <i>M. catarrhalis</i> , or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid sequence may be used to produce recombinant lactoferrin receptor proteins Lbp1, Lbp2 or ORF3 of the strain of <i>Moraxella</i> free of other proteins of the <i>Moraxella</i> strain for purposes of diagnostics and medical treatment. Furthermore, the nucleic acid molecule may be used in the diagnosis of infection.		

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TITLE OF INVENTIONLACTOFERRIN RECEPTOR GENES OF MORAXELLAFIELD OF INVENTION

5           The present invention relates to the molecular cloning of genes encoding lactoferrin receptor (LfR) proteins and, in particular, to the cloning of lactoferrin binding protein genes (lbp genes) from *Moraxella* (*Branhamella*) *catarrhalis*.

10                           REFERENCE TO RELATED APPLICATION

          This application is a continuation-in-part of copending United States patent application No. 08/867,941 filed June 3, 1997.

BACKGROUND OF THE INVENTION

15           *Moraxella* (*Branhamella*) *catarrhalis* bacteria are Gram-negative diplococcal pathogens which are carried asymptotically in the healthy human respiratory tract. However, in recent years, *M. catarrhalis* has been recognized as an important causative agent of otitis  
20   media. In addition, *M. catarrhalis* has been associated with sinusitis, conjunctivitis, and urogenital infections, as well as with a number of inflammatory diseases of the lower respiratory tract in children and adults, including pneumonia, chronic bronchitis,  
25   tracheitis, and emphysema (refs. 1 to 8). (Throughout this application, various references are cited in parentheses to describe more fully the state of the art to which this invention pertains. Full bibliographic information for each citation is found at the end of the  
30   specification, immediately preceding the claims. The disclosures of these references are hereby incorporated by reference into the present disclosure). Occasionally, *M. catarrhalis* invades to cause

septicaemia, arthritis, endocarditis, and meningitis (refs. 9 to 13).

5 *M. catarrhalis* colonizes the human upper respiratory tract and is an important cause of otitis media in infants and children as well as lower respiratory tract infections in adults with chronic obstructive pulmonary disease.

10 Otitis media is one of the most common illnesses of early childhood and approximately 80% of all children suffer at least one middle ear infection before the age of three (ref. 14). Chronic otitis media has been associated with auditory and speech impairment in children, and in some cases, has been associated with learning disabilities. Conventional treatments for  
15 otitis media include antibiotic administration and surgical procedures, including tonsillectomies, adenoidectomies, and tympanocentesis. In the United States, treatment costs for otitis media are estimated to be between one and two billion dollars per year.

20 In otitis media cases, *M. catarrhalis* is commonly co-isolated from middle ear fluid along with *Streptococcus pneumoniae* and non-typable *Haemophilus influenzae*, which are believed to be responsible for 50% and 30% of otitis media infections, respectively. *M.*  
25 *catarrhalis* is believed to be responsible for approximately 20% of otitis media infections (ref. 15). Epidemiological reports indicate that the number of cases of otitis media attributable to *M. catarrhalis* is increasing, along with the number of antibiotic-resistant isolates of *M. catarrhalis*. Thus, prior to  
30 1970, no  $\beta$ -lactamase-producing *M. catarrhalis* isolates had been reported, but since the mid-seventies, an increasing number of  $\beta$ -lactamase-expressing isolates have been detected. Recent surveys suggest that up to



80 to 85% of clinical isolates produce  $\beta$ -lactamase (ref. 16, 22, 23).

Iron-restriction is a general host defence mechanism against microbial pathogens. A number of bacterial species including *Neisseria meningitidis* (ref. 17, 24), *N. gonorrhoeae* (ref. 25) and *M. catarrhalis* (ref. 17), express outer membrane proteins which specifically bind human lactoferrin.

*M. catarrhalis* infection may lead to serious disease. It would be advantageous to provide a recombinant source of lactoferrin binding proteins as antigens in immunogenic preparations including vaccines, carriers for other antigens and immunogens and the generation of diagnostic reagents. The genes encoding lactoferrin binding proteins and fragments thereof are particularly desirable and useful in the specific identification and diagnosis of *Moraxella* and for immunization against disease caused by *M. catarrhalis* and for the generation of diagnostic reagents.

20

#### SUMMARY OF THE INVENTION

The present invention is directed towards the provision of purified and isolated nucleic acid molecules encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid molecules and isolated and purified lactoferrin binding proteins provided herein are useful for the specific detection of strains of *Moraxella* and for diagnosis of infection by *Moraxella*. The purified and isolated nucleic acid molecules provided herein, such as DNA, are also useful for expressing the *lbp* genes by recombinant DNA means for providing, in an economical manner, purified and isolated lactoferrin receptor

proteins free of other *Moraxella* proteins, as well as subunits, fragments or analogs thereof.

5 The lactoferrin receptor, subunits or fragments thereof or analogs thereof, as well as nucleic acid molecules encoding the same and vectors containing such nucleic acid molecules, are useful in immunogenic compositions for vaccinating against diseases caused by *Moraxella*, the diagnosis of infection by *Moraxella*, and as tools for the generation of immunological reagents.

10 Monoclonal antibodies or mono-specific antisera (antibodies) raised against the lactoferrin receptor protein produced in accordance with aspects of the present invention are useful for the diagnosis of infection by *Moraxella*, the specific detection of  
15 *Moraxella* (in, for example, *in vitro* and *in vivo* assays) and for the treatment of diseases caused by *Moraxella*.

In accordance with one aspect of the present invention, there is provided a purified and isolated  
20 nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella*, more particularly a strain of *M. catarrhalis*, specifically *M. catarrhalis* strain 4223, Q8 or VH19 or a fragment or an analog of the lactoferrin receptor protein. A fragment of the  
25 lactoferrin receptor protein is a portion of the protein which retains the immunological properties of the protein.

In one preferred embodiment of the invention, the nucleic acid molecule may encode only the Lbp1 protein  
30 of the *Moraxella* strain or only the Lbp2 protein of the *Moraxella* strain or only the ORF3 protein of the *Moraxella* strain. In another preferred embodiment of the invention, the nucleic acid may encode a fragment

of the lactoferrin receptor protein of a strain of *Moraxella* having a conserved amino acid sequence.

In a further aspect of the present invention, there is provided an isolated and purified nucleic acid molecule encoding at least one lactoferrin binding protein of *Moraxella* having a restriction map as shown in Figure 3 for *M. catarrhalis* 4223, Figure 5 for *M. catarrhalis* Q8 or Figure 17 for *M. catarrhalis* VH19 or the equivalent map from other strains of *Moraxella*.

In another aspect of the present invention, there is provided a purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of (a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto; (b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and (c) a DNA sequence encoding a functional lactoferrin receptor preprotein of *Moraxella*, which may be a DNA sequence which hybridizes under stringent conditions to any one of the DNA sequences defined in (a) or (b). The DNA sequence defined in (c) may have at least about 90% sequence identity with any one of the DNA sequences defined in (a) or (b). Stringent conditions of hybridization are described below. Sequence identity is determined in the manner described below.

In an additional aspect, the present invention includes a vector adapted for transformation of a host, comprising a nucleic acid molecule as provided herein and may have the characteristics of a nucleotide sequence contained within vectors pLD3, pLDW3, pLD1-8 and pLDW1.

The vector may be adapted for expression of the encoded lactoferrin receptor protein, fragments or

analogs thereof, in a heterologous or homologous host, in either a lipidated or non-lipidated form. Accordingly, a further aspect of the present invention provides an expression vector adapted for transformation of a host comprising a nucleic acid molecule as provided herein and expression means operatively coupled to the nucleic acid molecule for expression by the host of the lactoferrin receptor protein or the fragment or analog of the lactoferrin receptor protein.

In specific embodiments of this aspect of the invention, the nucleic acid molecule may encode substantially all the lactoferrin receptor protein, only the Lbp1 protein of the *Moraxella* strain, only the Lbp2 protein of the *Moraxella* strain, only the ORF3 protein of the *Moraxella* strain, or fragments of the Lbp1, Lbp2 or ORF3 proteins.

The expression means may include a nucleic acid portion encoding a leader sequence for secretion from the host of the lactoferrin receptor protein or the fragment or the analog of the lactoferrin receptor protein. The expression means also may include a nucleic acid portion encoding a lipidation signal for expression from the host of a lipidated form of the lactoferrin receptor protein or the fragment or the analog of the lactoferrin receptor protein. The host may be selected from, for example, *Escherichia coli*, *Bacillus*, *Bordetella*, *Haemophilus*, *Moraxella*, fungi, yeast or baculovirus and Semliki Forest virus expression system may be used. In a particular embodiment, the plasmid adapted for expression of Lbp2 is pRD2A, pRD2B, pQW2A or pQW2B; the plasmid adapted for expression of Lbp1 is pRD1A, pRD1B, PQ1A or pQ1B; and the plasmid adapted for expression of ORF3 is pLRD3 or pLQW3.

In an additional aspect of the invention, there is provided a transformed host containing an expression vector as provided herein. The invention further includes a recombinant lactoferrin receptor protein or  
5 fragment or analog thereof of a strain of *Moraxella* producible by the transformed host.

Such recombinant lactoferrin receptor protein may be provided in substantially pure form according to a further aspect of the invention, which provides a method  
10 of forming a substantially pure recombinant lactoferrin receptor protein, which comprises growing the transformed host provided herein and isolating and purifying the lactoferrin receptor protein, analog or fragment thereof. The lactoferrin receptor protein may  
15 be expressed in inclusion bodies, which may be purified free from cellular material and soluble proteins and lactoferrin receptor protein solubilized from the purified inclusion bodies, and the lactoferrin receptor protein purified free from other solubilized materials.  
20 The substantially pure recombinant lactoferrin receptor protein may comprise Lbp1 alone, Lbp2 alone, ORF3 or a mixture of two or more of such proteins. The recombinant protein is generally at least about 70% pure, preferably at least about 90% pure.

25 Further aspects of the present invention, therefore, provide recombinantly-produced Lbp1 protein (or a fragment or analog thereof) of a strain of *Moraxella* devoid of the Lbp2 and ORF3 proteins of the *Moraxella* strain and any other protein of the *Moraxella*  
30 strain, recombinantly-produced Lbp2 protein (or a fragment or analog thereof) of a strain of *Moraxella* devoid of the Lbp1 and ORF3 proteins of the *Moraxella* strain and any other protein of the *Moraxella* strain, and recombinantly-produced ORF3 protein (or a fragment  
35 or analog thereof) of a strain of *Moraxella* devoid of

the Lbp1 and Lbp2 proteins of the *Moraxella* strain and any other protein of the *Moraxella* strain. The *Moraxella* strain may be *M. catarrhalis* 4223, Q8 or VH19 strain.

5           The invention further includes, in an additional aspect, an open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the lactoferrin binding protein which is encoded by region downstream from the genes encoding Lbp2 and Lbp1  
10       proteins of the *Moraxella* strain. The ORF3 may be from a strain of *M. catarrhalis*, which may be strain 4223 or Q8. The Lbp3 may have a molecular mass of about 60 kDa.

          In accordance with another aspect of the  
15       invention, an immunogenic composition is provided which comprises at least one active component selected from at least one nucleic acid molecule as provided herein, at least one recombinant protein as provided herein or at least one novel protein as provided herein, and a  
20       pharmaceutically acceptable carrier therefor or vector therefor. The at least one active component produces an immune response when administered to a host.

          The immunogenic compositions provided herein may be formulated as a vaccine for *in vivo* administration  
25       to a host to provide protection against disease caused by *M. catarrhalis*. For such purpose, the compositions may be formulated as a microparticle, capsule, ISCOM or liposome preparation. The immunogenic composition may be provided in combination with a targeting molecule  
30       for delivery to specific cells of the immune system or to mucosal surfaces. The immunogenic compositions of the invention (including vaccines) may further comprise at least one other immunogenic or immunostimulating material and the immunostimulating material may be at  
35       least one adjuvant or at least one cytokine.



Suitable adjuvants for use in the present invention include (but are not limited to) aluminum phosphate, aluminum hydroxide, QS21, Quil A, derivatives and components thereof, ISCOM matrix, calcium phosphate, calcium hydroxide, zinc hydroxide, a glycolipid analog, an octadecyl ester of an amino acid, a muramyl dipeptide, polyphosphazene, ISCOPREP, DC-chol, DDBA and a lipoprotein and other adjuvants to induce a TH1 response. Advantageous combination of adjuvants are described in copending United States Patent Applications No. 08/261,194 filed June 16, 1994 and 08/483,856 filed June 7, 1995, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference (WO 95/34308, published November 21, 1995).

In accordance with another aspect of the invention, there is provided a method for generating an immune response in a host, comprising the step of administering to a susceptible host, such as a human, an effective amount of the immunogenic composition as recited above. The immune response may be humoral or a cell-mediated immune response and may provide protection against disease caused by *Moraxella*. Hosts in which protection against disease may be conferred include primates, including humans.

In a further aspect, there is provided a live vector for delivery of lactoferrin receptor to a host, comprising a vector containing the nucleic acid molecule as described above. The vector may be selected from *Salmonella*, *Mycobacterium bovis*, BCG, adenovirus, poxvirus, vaccinia and poliovirus.

The nucleic acid molecules provided herein are useful in diagnostic applications. Accordingly, in a further aspect of the invention, there is provided a method of determining the presence, in a sample, of



nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising the steps of:

5 a) contacting the sample with a nucleic acid molecule as provided herein to produce duplexes comprising the nucleic acid molecule and any nucleic acid molecule encoding the lactoferrin receptor protein of a strain of *Moraxella* present in the sample and specifically hybridizable therewith; and

b) determining the production of the duplexes.

10 In addition, the present invention provides a diagnostic kit for determining the presence, in a sample, of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising:

a) a nucleic acid molecule as provided herein;

15 b) means for contacting the nucleic acid molecule with the sample to produce duplexes comprising the nucleic acid molecule and any such nucleic acid present in the sample and hybridizable with the nucleic acid present in the sample and hybridizable with the nucleic acid molecule; and

20 c) means for determining production of the duplexes.

The invention further includes the use of the nucleic acid molecules and proteins provided herein as medicines. The invention additionally includes the use of the nucleic acid molecules and proteins provided herein in the manufacture of medicaments for protection against disease caused by strains of *Moraxella*.

Advantages of the present invention include:

- 30 - an isolated and purified nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein;
- recombinantly-produced lactoferrin receptor proteins, including Lbp1, Lbp2 and ORF3 and fragments
- 35

and analogs thereof free from each other and other *Moraxella* proteins;

- open reading frame protein 3; and
- diagnostic kits and immunological reagents for specific identification of *Moraxella*.

#### **BRIEF DESCRIPTION OF DRAWINGS**

The present invention will be further understood from the following description with reference to the drawings, in which:

10        Figure 1 shows partial sequence of the 2.2 kb PCR amplified fragments of the *lbpA* genes from *M. catarrhalis* 4223 or Q8, which were used to probe the phage libraries. In the figure, Tbp1 is the deduced 4223 Tbp1 sequence (as described in United States  
15        Patent Application No. 08/613,009 filed March 8, 1996, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference) (SEQ ID No: 19), Lbp1 is the deduced full-length 4223 Lbp1 sequence (SEQ ID No: 3) used here solely for aligning the PCR  
20        fragments, PCR4 is the 4223 PCR fragment (SEQ ID No: 20), and PCR5 is a partial sequence of the Q8 PCR fragment (SEQ ID No: 21). Only single strand sequence was obtained for the PCR fragments and "X" has been inserted where there was a doubtful sequence. Dashes  
25        have been used for maximum alignment. The underlined sequence in Lbp1 (MVQYTYRKGKKNKAH - SEQ ID No: 22) represents the position of a CNBr peptide used to generate the 5'-PCR primer.

30        Figure 2 shows the nucleotide (SEQ ID No: 1, full sequence; SEQ ID No: 2, Lbp2 coding sequence; SEQ ID No: 3, Lbp1 coding sequence, first methionine; SEQ ID No: 4, Lbp1 coding sequence, second methionine; SEQ ID No: 5, ORF3 coding sequence) and deduced amino acid sequences (SEQ ID No: 11, Lbp2; SEQ ID No: 12, Lbp1,  
35        first methionine; SEQ ID No: 13, Lbp1, second

methionine; SEQ ID No: 14, ORF3) of the putative *lfr* locus from *M. catarrhalis* 4223. There are three tandem genes in the putative *lfr* locus identified as *lbpB*, *lbpA* and *orf3*. Potential promoter elements found upstream of the *lbpB* and *lbpA* genes are indicated by underlining.

Figure 3 shows a restriction map of clone pLD1-8 containing the *lbpA*, *lbpB*, and *orf3* genes from *M. catarrhalis* isolate 4223.

Figure 4 shows the nucleotide (SEQ ID No: 6, full sequence; SEQ ID No: 7, *Lbp2* coding sequence; SEQ ID No: 8, *Lbp1* coding sequence, first methionine; SEQ ID No: 9, *Lbp2*, second methionine; SEQ ID No: 10, ORF3 coding sequence) and deduced amino acid sequences (SEQ ID No: 15, *Lbp2*; SEQ ID No: 16, *Lbp1*, first methionine; SEQ ID No: 17, *Lbp1*, second methionine; SEQ ID No: 18, *Lbp3*) of the putative *lfr* locus from *M. catarrhalis* Q8. There are three tandem genes in the putative *lfr* locus identified as *lbpB*, *lbpA* and *orf3*. Potential promoter elements found upstream of the *lbpB* and *lbpA* genes are indicated by underlining.

Figure 5 shows a restriction map of clone pLDW1 containing the *lbpA*, *lbpB* and *orf3* genes from *M. catarrhalis* isolate Q8.

Figure 6 shows a comparison of the amino acid sequences of *Lbp1* from *M. catarrhalis* strains 4223 (SEQ ID No: 12) and Q8 (SEQ ID No: 16), *N. meningitidis* strains BNCV (SEQ ID No: 23) and H44/76 (SEQ ID No: 75), and *N. gonorrhoeae* strain FA19 (SEQ ID No: 24). Also shown is the partial carboxy terminal sequence of *Lbp2* from *N. meningitidis* strains BNCV (SEQ ID No: 76) and H44/76 (SEQ ID No: 77) and *N. gonorrhoeae* strain FA19 (SEQ ID No: 88). Dots indicate identical residues

and dashes have been introduced to achieve maximum sequence alignment.

Figure 7 shows a comparison of the amino acid sequences of Lbp2 from *M. catarrhalis* strains 4223 (SEQ ID No: 11), Q8 (SEQ ID No: 15) and VH19 (SEQ ID No: 70). Dots indicate identical residues. The arrow indicates the lipidated cysteine of a potential mature Lbp2 lipoprotein. The residues conserved with Tbp2 proteins are underlined and the RGD sequence is italicized.

Figure 8 shows a comparison of the amino acid sequences of Tbp2 (USPA No: 08/613,009) (SEQ ID No: 25) and Lbp2 from *M. catarrhalis* strain 4223 (SEQ ID No: 11). Dots indicate identical residues and dashes have been inserted to achieve maximum sequence alignment. The asterisks indicate conserved residues and the putative site of lipidation for both proteins is indicated by the arrow.

Figure 9 shows a comparison of the amino acid sequences of ORF3 from *M. catarrhalis* strains 4223 (SEQ ID No: 14) and Q8 (SEQ ID No: 18). Dots indicate identical residues and dashes have been introduced for maximum alignment.

Figure 10 shows the construction of plasmids for expression of recombinant Lbp1 protein from *E. coli*. Plasmids pRD1A and pRD1B express 4223 Lbp1 from the first or second methionine residues, respectively. Plasmids pQW1A and pQW1B express Q8 Lbp1 from the first or second methionine residues, respectively.

Figure 11, comprising panels A and B, shows the expression of recombinant Lbp1 (rLbp1) proteins from *E. coli*. Panel A shows the expression of the Q8 Lbp1 proteins and panel B shows the expression of the 4223 Lbp1 proteins. Lane 1, molecular weight marker. Lanes 2 and 3 demonstrate the induced expression of the

longer Lbp1 starting from the first methionine residues and lanes 4 and 5 illustrate the expression of the shorter Lbp1 proteins starting from the second methionine residues. Lanes 6, 7, 8 and 9 are uninduced samples.

Figure 12 shows the construction of plasmids for expression of recombinant Lbp2 (rLbp2) protein from *E. coli*. Plasmids pRD2A and pRD2B express 4223 Lbp2 with or without the native leader sequence, respectively. Plasmids pQW2A and pQW2B express Q8 Lbp2 with or without the native leader sequence, respectively.

Figure 13 shows the construction of a plasmid for expression of recombinant ORF3 (rORF3) proteins from *E. coli*.

Figure 14 shows a purification scheme for rLbp1 expressed from *E. coli*.

Figure 15 shows an SDS PAGE gel of the purification of Q8 Lbp1 from *E. coli*. Lane 1, BL21(DE3) lysate; lane 2, soluble proteins after 50 mM Tris/5 mM AEBSF/0.5 M NaCl, pH 8.0 extraction; lane 3, soluble proteins after 50 mM Tris/0.5% Triton X-100/10 mM EDTA, pH 8.0 extraction; lane 4, soluble proteins after 50 mM Tris-HCl/1% octylglucoside, pH 8.0 extraction; lane 5, solubilized inclusion bodies; lane 6, purified Lbp1.

Figure 16 shows the nucleotide sequence (SEQ ID No: 69) of the *M. catarrhalis* strain VH19 *lbpB* gene and the deduced amino acid sequence (SEQ ID No: 70) of the corresponding Lbp2 protein.

Figure 17 shows a partial restriction map of the *M. catarrhalis* strain VH19 *lbpB* gene.

Figure 18, comprising panels A, B and C, shows SDS-PAGE gels of the purification of recombinant Lbp proteins. Panel A shows an SDS-PAGE gel of the purification of Q8 rLbp1. Panels B and C show the

purification of Q8 rLbp2 and 4223 rLbp2, respectively. Lane 1, molecular weight markers; lane 2, whole cell lysates; lane 3, inclusion bodies; lane 4, purified protein.

5           Figure 19, comprising panels A and B, shows binding of recombinant Lbp proteins to lactoferrin. Panel A shows an SDS PAGE gel of purified recombinant proteins. Panel B shows the binding of recombinant proteins to human lactoferrin. Lane 1, molecular  
10           weight markers; lane 2, Q8 rLbp1; lane 3, Q8 rLbp2; lane 4, 4223 rLbp2.

          Figure 20, comprising panels A, B and C, shows an immunoblot of *M. catarrhalis* strains reacted with anti-rLbp1 and anti-rLbp2 antibodies. Panel A: whole cell  
15           lysates probed with anti-Q8 rLbp1 + anti-Q8 rLbp2 antisera. All cells were grown in the presence of EDDA. Panel B: whole cell lysates probed with anti-Q8 rLbp1 antibody. Panel C: whole cell lysates probed with anti-Q8 rLbp2 antibody. Lane 1, strain Q8; lane  
20           2, strain 4223; lane 3, strain VH19; lane 4, strain LES-1; lane 5, strain H-04; lane 6, strain 3. + indicates growth in the presence of EDDA and - indicates growth in the absence of EDDA.

#### GENERAL DESCRIPTION OF THE INVENTION

25           Any *Moraxella* strain may be conveniently used to provide the purified and isolated nucleic acid, which may be in the form of DNA molecules, comprising at least a portion of the nucleic acid coding for a lactoferrin receptor as typified by embodiments of the  
30           present invention. Such strains are generally available from clinical sources and from bacterial culture collections, such as the American Type Culture Collection.



In this application, the terms "lactoferrin receptor" (LfR) and "lactoferrin binding proteins" (Lbp) are used to define a family of Lbp1, Lbp2 and/or ORF3 proteins which includes those having variations in their amino acid sequences including those naturally occurring in various strains of, for example, *Moraxella*. The purified and isolated DNA molecules comprising at least a portion coding for lactoferrin receptor of the present invention also includes those encoding functional analogs of lactoferrin receptor proteins Lbp1, Lbp2 and/or Lbp3 of *Moraxella*. In this application, a first protein is an "analog" of a second protein if the first protein is immunologically related to and/or has the same function as the second protein. The analog may be, for example, a substitution, addition or deletion mutant thereof.

Lactoferrin receptor proteins were purified from *M. catarrhalis* membrane preparations by affinity chromatography on biotinylated human lactoferrin. Cyanogen bromide fragments were generated and amino acid sequence analysis of a 13 kDa fragment provided an internal Lbp1 sequence of MVQYTYRKGKENKAH (SEQ ID No: 22) underlined in Figure 6. The C-terminus of *M. catarrhalis* Tbp1 (United States Patent Application No. 08/613,009), *N. meningitidis* Tbp1 (ref. 27) and *H. influenzae* Tbp1 (ref. 31) has a conserved LEMKF (SEQ ID No: 26) sequence. Oligonucleotide primers were generated based upon these two sequences and used to PCR amplify an approximately 2.2kb fragment of the *lbpA* gene from *M. catarrhalis* strains 4223, Q8 and VH19. Partial sequence analysis demonstrated that the amplified genes were *lbpA* and not *tbpA* (see Fig. 1). The 2.2 kb PCR fragments were used to screen genomic libraries.



Chromosomal DNA from 4223, Q8 and VH19 was partially digested with Sau3A I and 15 to 23 kb fragments were purified before cloning into BamH I arms of the lambda vector EMBL3. The libraries were  
5 screened with the PCR fragment and positive clones were subjected to three rounds of plaque purification. Phage clone 4223LfR.17 containing an approximately 16 kb insert from 4223 and phage clone Q8LfR.13 containing an approximately 16 kb insert from Q8 were selected for  
10 further analysis.

Restriction enzyme and Southern blot analyses revealed that an internal Hind III fragment of approximately 9 kb contained at least a portion of the *lbpA* gene for both phage clones. The approximately 9  
15 kb Hind III fragments were subcloned into pUC or pBluescript-based plasmids and sequenced. In each case, they contained the complete *lbpA* gene as well as an upstream gene identified as *lbpB*, and a downstream gene designated as *orf3*. The *lbpB-lbpA* gene  
20 arrangement is the same as present for *Neisseria* strains, but there has been no identification of a third gene for these organisms.

The gene arrangement is different than that observed for the *M. catarrhalis* *tfr* operon which was  
25 *tbpA-orf-tbpB* (United States Patent Application No. 08/613,009). There are promoter elements found upstream of both the *lbpB* and *lbpA* genes from strains 4223 and Q8. The third ORF is located immediately downstream of *lbpA*, separated by a single nucleotide.

30 By analogy with the *N. meningitidis* and *N. gonorrhoeae* transferrin receptor operons (ref. 26, 27, 28), the lactoferrin receptor operon was presumed to consist of two genes encoding lactoferrin binding proteins 1 and 2 (*Lbp1* and *Lbp2*) (ref. 29). However,

we report here that, for *M. catarrhalis*, there also appears to be a third gene located immediately downstream of *lbpA* encoding a potential lactoferrin binding protein 3 (ORF3).

5       The *M. catarrhalis* 4223 and Q8 *lbpA* genes encode proteins of molecular mass about 110 kDa and that are highly conserved with only seven residues difference between them. The N-terminal sequence of the native Lbp protein is unknown and there are two possible ATG  
10       start codons at positions 1 or 16. The first of these is adjacent to consensus sequences for promoter elements and the second is followed by a putative signal sequence. The exact peptide sequence used to design the PCR amplification primers was not found.  
15       When compared with other known Lbp1 sequences from *N. meningitidis* (refs. 31, 24) or *N. gonorrhoeae* (ref. 25) there is about 32% sequence identity and about 50% sequence homology between the *M. catarrhalis* and the *Neisseria* proteins. There is some homology between the  
20       *M. catarrhalis* Lbp1 and Tbp1 proteins as shown in Figure 1, but it is very scattered.

      The *M. catarrhalis* 4223, Q8 and VH19 *lbpB* genes encode 898, 894 and 906 amino acid proteins, respectively. The *M. catarrhalis* Lbp2 proteins from  
25       strains 4223 and Q8 are 92% identical and 95% homologous while that from VH19 is 77% identical and 84% similar to the 4223 and Q8 Lbp2 proteins (Figure 7). There is a consensus sequence for lipidation at the Cys<sup>32</sup> residue, suggesting that Lbp2 is a lipoprotein  
30       like Tbp2. There is little homology between the *M. catarrhalis* Lbp2 and Tbp2 proteins (Fig. 8) with the exception of a previously identified peptide sequence (LEGGFY (SEQ ID No: 27)) that is also found in *N. meningitidis* and *H. influenzae* Tbp2 (ref. 30).

The sequence of the proposed *M. catarrhalis* *lfr*-related downstream *orf3* is conserved between strains 4223 and Q8. The encoded 4223 and Q8 ORF3 proteins when compared to the PIR and Swiss Prot protein databases were found to be previously unknown. The ORF3 protein may bind lactoferrin itself or may be an associated or regulatory protein for Lbp1 and/or Lbp2.

Expression vectors have been assembled from the *lbpA* and *lbpB* genes and recombinant Lbp1 and Lbp2 proteins isolated and purified, as described in detail in the Examples below.

Results shown in Table 1 below illustrate the ability of anti-Lbp1 guinea pig antiserum, produced by immunization with affinity purified Lbp1, to lyse *M. catarrhalis*. The results show that the antisera produced by immunization with Lbp1 protein isolated from *M. catarrhalis* isolate 4223 was bactericidal against a homologous non-clumping *M. catarrhalis* strain RH408 (a strain previously deposited in connection with United States Patent Application No. 08/328,589, assigned to the assignee hereof (WO 96/12733 published May 2, 1996)) derived from isolate 4223. In addition, antisera produced by immunization with Lbp1 protein isolated from *M. catarrhalis* 4223 were bactericidal against the heterologous non-clumping strain Q8. The results in Table 3 show that similarly-produced anti-Lbp2 guinea pig antiserum was bactericidal for the homologous strain and for three of five heterologous strains. The ability of isolated and purified lactoferrin binding protein to generate bactericidal antibodies is *in vivo* evidence of utility of these proteins as vaccines to protect against disease caused by *Moraxella*.

Thus, in accordance with another aspect of the present invention, there is provided a vaccine against *Moraxella* comprising an immunogenically-effective amount of lactoferrin binding protein or fragment or analog thereof, or a nucleic acid molecule (DNA or RNA) encoding the lactoferrin binding protein or fragment or analog thereof, and a physiologically-acceptable carrier therefor. The lactoferrin binding protein or fragment or analog thereof provided herein may also be used as a carrier protein for haptens, polysaccharide or peptides to make conjugate vaccines against antigenic determinants unrelated to lactoferrin binding proteins.

In additional embodiments of the present invention, therefore, the lactoferrin binding protein as provided herein may be used as a carrier molecule to prepare chimeric molecules and conjugate vaccines (including glycoconjugates) against pathogenic bacteria, including encapsulated bacteria. Thus, for example, glycoconjugates of the present invention may be used to confer protection against disease and infection caused by any bacteria having polysaccharide antigens including lipooligosaccharides (LOS) and PRP. Such bacterial pathogens may include, for example, *Haemophilus influenzae*, *Streptococcus pneumoniae*, *Escherichia coli*, *Neisseria meningitidis*, *Salmonella typhi*, *Streptococcus mutans*, *Cryptococcus neoformans*, *Klebsiella*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. Particular antigens which can be conjugated to lactoferrin binding protein and methods to achieve such conjugations are described in U.S. Patent Application No. 08/433,522 filed November 23, 1993 (WO 94/12641), assigned to the assignee hereof and the disclosure of which is hereby incorporated by reference thereto.

In another embodiment, the carrier function of lactoferrin binding protein may be used, for example, to induce an immune response against abnormal polysaccharides of tumour cells, or to produce anti-tumour antibodies that can be conjugated to chemotherapeutic or bioactive agents.

The lactoferrin binding protein provided herein is useful as a diagnostic reagent, as an antigen or for the generation of anti-lactoferrin protein binding antibodies, antigen for vaccination against disease caused by species of *Moraxella* and for detecting infection by *Moraxella* and other such bacteria.

The invention extends to lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same from *Moraxella catarrhalis* for use as an active ingredient in a vaccine against disease caused by infection with *Moraxella*. The invention also extends to a pharmaceutical vaccinal composition containing lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same from *Moraxella catarrhalis* and optionally, a pharmaceutically acceptable carrier and/or diluent.

In a further aspect the invention provides the use of lactoferrin binding proteins or fragments or analogs thereof or nucleic acid molecules encoding the same for the preparation of a pharmaceutical vaccinal composition for immunization against disease caused by infection with *Moraxella*.

It is clearly apparent to one skilled in the art, that the various embodiments of the present invention have many applications in the fields of vaccination, diagnosis, treatment of, for example, *Moraxella* infections and the generation of immunological and other

diagnostic reagents. A further non-limiting discussion of such uses is further presented below.

### 1. Vaccine Preparation and Use

Immunogenic compositions, suitable to be used as vaccines, may be prepared from immunogenic lactoferrin receptor proteins, analogs and fragments thereof encoded by the nucleic acid molecules as well as the nucleic acid molecules disclosed herein. The vaccine elicits an immune response which produces antibodies, including anti-lactoferrin receptor antibodies and antibodies that are opsonizing or bactericidal. Should the vaccinated subject be challenged by *Moraxella*, the antibodies bind to the lactoferrin receptor and thereby prevent access of the bacteria to an iron source which is required for viability. Furthermore, opsonizing or bactericidal anti-lactoferrin receptor antibodies may also provide protection by alternative mechanisms.

Immunogenic compositions, including vaccines, may be prepared as injectables, as liquid solutions or emulsions. The lactoferrin receptor proteins, analogs and fragments thereof and encoding nucleic acid molecules as well as the nucleic acid molecules described herein may be mixed with pharmaceutically acceptable excipients which are compatible with the lactoferrin receptor proteins, fragments, analogs or nucleic acid molecules. Such excipients may include water, saline, dextrose, glycerol, ethanol, and combinations thereof. The immunogenic compositions and vaccines may further contain auxiliary substances, such as wetting or emulsifying agents, pH buffering agents, or adjuvants, to enhance the effectiveness of the vaccines. Immunogenic compositions and vaccines may be administered parenterally, by injection subcutaneously, intradermally or intramuscularly. Alternatively, the immunogenic compositions provided according to the



present invention, may be formulated and delivered in a manner to evoke an immune response at mucosal surfaces. Thus, the immunogenic composition may be administered to mucosal surfaces by, for example, the nasal or oral (intragastric) routes. The immunogenic composition may be provided in combination with a targeting molecule for delivery to specific cells of the immune system or to mucosal surfaces. Some such targeting molecules include vitamin B12 and fragments of bacterial toxins, as described in WO 92/17167 (Biotech Australia Pty. Ltd.), and monoclonal antibodies, as described in U.S. Patent No. 5,194,254 (Barber et al). Alternatively, other modes of administration, including suppositories and oral formulations, may be desirable. For suppositories, binders and carriers may include, for example, polyalkylene glycols or triglycerides. Oral formulations may include normally employed incipients such as, for example, pharmaceutical grades of saccharine, cellulose and magnesium carbonate. These compositions may take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 1 to 95% of the lactoferrin receptor proteins, fragments, analogs and/or nucleic acid molecules.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective, protective and immunogenic. The quantity to be administered depends on the subject to be treated, including, for example, the capacity of the individual's immune system to synthesize antibodies, and, if needed, to produce a cell-mediated immune response. Precise amounts of active ingredient required to be administered depend on the judgement of the practitioner. However, suitable dosage ranges are readily determinable by one skilled in



the art and may be of the order of micrograms of the lactoferrin receptor proteins, analogs and fragments thereof and/or nucleic acid molecules. Suitable regimes for initial administration and booster doses are also  
5 variable, but may include an initial administration followed by subsequent administrations. The dosage of the vaccine may also depend on the route of administration and will vary according to the size of the host.

10 The nucleic acid molecules encoding the lactoferrin receptor of *Moraxella* may be used directly for immunization by administration of the DNA directly, for example, by injection for genetic immunization or by constructing a live vector, such as *Salmonella*, BCG,  
15 adenovirus, poxvirus, vaccinia or poliovirus containing the nucleic acid molecules. A discussion of some live vectors that have been used to carry heterologous antigens to the immune system is contained in, for example, O'Hagan (ref. 18). Processes for the direct  
20 injection of DNA into test subjects for genetic immunization are described in, for example, Ulmer et al. (ref. 19).

Immunogenicity can be significantly improved if the antigens are co-administered with adjuvants, commonly  
25 used as an 0.05 to 1.0 percent solution in phosphate - buffered saline. Adjuvants enhance the immunogenicity of an antigen but are not necessarily immunogenic themselves. Adjuvants may act by retaining the antigen locally near the site of administration to produce a  
30 depot effect facilitating a slow, sustained release of antigen to cells of the immune system. Adjuvants can also attract cells of the immune system to an antigen depot and stimulate such cells to elicit immune responses.

Immunostimulatory agents or adjuvants have been used for many years to improve the host immune responses to, for example, vaccines. Intrinsic adjuvants, such as lipopolysaccharides, normally are the components of killed or attenuated bacteria used as vaccines. Extrinsic adjuvants are immunomodulators which are typically non-covalently linked to antigens and are formulated to enhance the host immune responses. Thus, adjuvants have been identified that enhance the immune response to antigens delivered parenterally. Some of these adjuvants are toxic, however, and can cause undesirable side-effects, making them unsuitable for use in humans and many animals. Indeed, only aluminum hydroxide and aluminum phosphate (collectively commonly referred to as alum) are routinely used as adjuvants in human and veterinary vaccines. The efficacy of alum in increasing antibody responses to diphtheria and tetanus toxoids is well established and an HBsAg vaccine has been adjuvanted with alum.

A wide range of extrinsic adjuvants can provoke potent immune responses to antigens. These include saponins complexed to membrane protein antigens (immune stimulating complexes), pluronic polymers with mineral oil, killed mycobacteria and mineral oil, Freund's complete adjuvant, bacterial products, such as muramyl dipeptide (MDP) and lipopolysaccharide (LPS), as well as lipid A, and liposomes.

To efficiently induce humoral immune responses (HIR) and cell-mediated immunity (CMI), immunogens are often emulsified in adjuvants. Many adjuvants are toxic, inducing granulomas, acute and chronic inflammations (Freund's complete adjuvant, FCA), cytotoxicity (saponins and pluronic polymers) and pyrogenicity, arthritis and anterior uveitis (LPS and MDP). Although FCA is an excellent adjuvant and widely

used in research, it is not licensed for use in human or veterinary vaccines because of its toxicity.

Desirable characteristics of ideal adjuvants include:

- 5 (1) lack of toxicity;
- (2) ability to stimulate a long-lasting immune response;
- (3) simplicity of manufacture and stability in long-term storage;
- 10 (4) ability to elicit both CMI and HIR to antigens administered by various routes, if required;
- (5) synergy with other adjuvants;
- (6) capability of selectively interacting with populations of antigen presenting cells (APC);
- 15 (7) ability to specifically elicit appropriate  $T_H1$  or  $T_H2$  cell-specific immune responses; and
- (8) ability to selectively increase appropriate antibody isotype levels (for example, IgA) against antigens.

20 U.S. Patent No. 4,855,283 granted to Lockhoff et al on August 8, 1989, which is incorporated herein by reference thereto, teaches glycolipid analogues including N-glycosylamides, N-glycosylureas and N-glycosylcarbamates, each of which is substituted in the  
25 sugar residue by an amino acid, as immuno-modulators or adjuvants. Thus, Lockhoff et al. 1991 (ref. 20) reported that N-glycolipid analogs displaying structural similarities to the naturally-occurring glycolipids, such as glycopospholipids and glyco glycerolipids, are  
30 capable of eliciting strong immune responses in both herpes simplex virus vaccine and pseudorabies virus vaccine. Some glycolipids have been synthesized from long chain-alkylamines and fatty acids that are linked directly with the sugars through the anomeric carbon

atom, to mimic the functions of the naturally occurring lipid residues.

U.S. Patent No. 4,258,029 granted to Moloney, assigned to the assignee hereof and incorporated herein by reference thereto, teaches that octadecyl tyrosine hydrochloride (OTH) functions as an adjuvant when complexed with tetanus toxoid and formalin inactivated type I, II and III poliomyelitis virus vaccine. Also, Nixon-George et al. 1990, (ref. 21) reported that octadecyl esters of aromatic amino acids complexed with a recombinant hepatitis B surface antigen, enhanced the host immune responses against hepatitis B virus.

## 2. Immunoassays

The lactoferrin receptor proteins, analogs and/or fragments thereof of the present invention are useful as immunogens, as antigens in immunoassays including enzyme-linked immunosorbent assays (ELISA), RIAs and other non-enzyme linked antibody binding assays or procedures known in the art for the detection of anti-*Moraxella*, lactoferrin receptor protein antibodies. In ELISA assays, the lactoferrin receptor protein, analogs and/or fragments corresponding to portions of Lfr protein, are immobilized onto a selected surface, for example, a surface capable of binding proteins or peptides such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed lactoferrin receptor, analogs and/or fragments, a non-specific protein such as a solution of bovine serum albumin (BSA) or casein that is known to be antigenically neutral with regard to the test sample may be bound to the selected surface. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by non-specific bindings of antisera onto the surface.

The immobilizing surface is then contacted with a sample, such as clinical or biological materials, to be tested in a manner conducive to immune complex (antigen/antibody) formation. This procedure may include diluting the sample with diluents, such as BSA, bovine gamma globulin (BGG) and/or phosphate buffered saline (PBS)/Tween. The sample is then allowed to incubate for from about 2 to 4 hours, at temperatures such as of the order of about 25° to 37°C. Following incubation, the sample-contacted surface is washed to remove non-immunocomplexed material. The washing procedure may include washing with a solution such as PBS/Tween or a borate buffer.

Following formation of specific immunocomplexes between the test sample and the bound lactoferrin receptor protein, analogs and/or fragments and subsequent washing, the occurrence, and even amount, of immunocomplex formation may be determined by subjecting the immunocomplex to a second antibody having specificity for the first antibody. If the test sample is of human origin, the second antibody is an antibody having specificity for human immunoglobulins and in general IgG. To provide detecting means, the second antibody may have an associated activity such as an enzymatic activity that will generate, for example, a color development upon incubating with an appropriate chromogenic substrate. Quantification may then be achieved by measuring the degree of color generation using, for example, a spectrophotometer.

### 30      3.      Use of Sequences as Hybridization Probes

The nucleotide sequences of the present invention, comprising the sequence of the lactoferrin receptor gene, now allow for the identification and cloning of the lactoferrin receptor genes from any species of *Moraxella*.

The nucleotide sequences comprising the sequence of the lactoferrin receptor genes of the present invention are useful for their ability to selectively form duplex molecules with complementary stretches of other *lfr* genes. Depending on the application, a variety of hybridization conditions may be employed to achieve varying degrees of selectivity of the probe toward the other *lfr* genes. For a high degree of selectivity, relatively stringent conditions are used to form the duplexes, such as low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of between about 50°C to 70°C. For some applications, less stringent hybridization conditions are required such as 0.15 M to 0.9 M salt, at temperatures ranging from between about 20°C to 55°C. Hybridization conditions can also be rendered more stringent by the addition of increasing amounts of formamide, to destabilize the hybrid duplex. Thus, particular hybridization conditions can be readily manipulated, and will generally be a method of choice depending on the desired results. In general, convenient hybridization temperatures in the presence of 50% formamide are: 42°C for a probe which is 95 to 100% homologous to the target fragment, 37°C for 90 to 95% homology and 32°C for 85 to 90% homology.

Such hybridization conditions may be employed to determine DNA sequences which encode a functional lactoferrin receptor of *Moraxella* and which hybridize under stringent conditions to any one of the DNA sequences (a) or (b), described above.

In a clinical diagnostic embodiment, the nucleic acid sequences of the *lfr* genes of the present invention may be used in combination with an appropriate means, such as a label, for determining hybridization. A wide



variety of appropriate indicator means are known in the art, including radioactive, enzymatic or other ligands, such as avidin/biotin and digoxigenin-labelling, which are capable of providing a detectable signal. In some  
5 diagnostic embodiments, an enzyme tag such as urease, alkaline phosphatase or peroxidase, instead of a radioactive tag may be used. In the case of enzyme tags, colorimetric indicator substrates are known which can be employed to provide a means visible to the human  
10 eye or spectrophotometrically, to identify specific hybridization with samples containing *lfr* gene sequences.

The nucleic acid sequences of *lfr* genes of the present invention are useful as hybridization probes in  
15 solution hybridizations and in embodiments employing solid-phase procedures. In embodiments involving solid-phase procedures, the test DNA (or RNA) from samples, such as clinical samples, including exudates, body fluids (e. g., serum, amniotic fluid, middle ear  
20 effusion, sputum, bronchoalveolar lavage fluid) or even tissues, is adsorbed or otherwise affixed to a selected matrix or surface. The fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes comprising the nucleic acid sequences of  
25 the *lfr* genes or fragments thereof of the present invention under desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required depending on, for example, the G+C contents, type of target nucleic  
30 acid, source of nucleic acid, size of hybridization probe etc. Following washing of the hybridization surface so as to remove non-specifically bound probe molecules, specific hybridization is detected, or even quantified, by means of the label. It is preferred to  
35 select nucleic acid sequence portions which are



conserved among species of *Moraxella*. The selected probe may be at least 18bp and may be in the range of about 30 to 90 bp.

#### 4. Expression of the Lactoferrin Receptor Genes

5 Plasmid vectors containing replicon and control sequences which are derived from species compatible with the host cell may be used for the expression of the lactoferrin receptor genes in expression systems. The vector ordinarily carries a replication site, as well as  
10 marking sequences which are capable of providing phenotypic selection in transformed cells. For example, *E. coli* may be transformed using pBR322 which contains genes for ampicillin and tetracycline resistance and thus provides easy means for identifying transformed  
15 cells. The pBR322 plasmid, or other microbial plasmid or phage, must also contain, or be modified to contain, promoters which can be used by the host cell for expression of its own proteins.

In addition, phage vectors containing replicon and  
20 control sequences that are compatible with the host can be used as a transforming vector in connection with these hosts. For example, the phage in lambda GEM<sup>TM</sup>-11 may be utilized in making recombinant phage vectors which can be used to transform host cells, such as *E.*  
25 *coli* LE392.

Promoters commonly used in recombinant DNA construction include the  $\beta$ -lactamase (penicillinase) and lactose promoter systems and other microbial promoters, such as the T7 promoter system as described in U.S.  
30 Patent No. 4,952,496. Details concerning the nucleotide sequences of promoters are known, enabling a skilled worker to ligate them functionally with genes. The particular promoter used will generally be a matter of choice depending upon the desired results. Hosts that

are appropriate for expression of the lactoferrin receptor genes, fragments or analogs thereof, may include *E. coli*, *Bacillus* species, *Haemophilus*, fungi, yeast, *Moraxella*, *Bordetella*, or the baculovirus expression system may be used.

In accordance with this invention, it is preferred to produce the lactoferrin receptor protein, fragment or analog thereof, by recombinant methods, particularly since the naturally occurring LfR protein as purified from a culture of a species of *Moraxella* may include trace amounts of toxic materials or other contaminants. This problem can be avoided by using recombinantly produced LfR protein in heterologous systems which can be isolated from the host in a manner to minimize contaminants, including other proteins of the *Moraxella* strain, in the purified material. Particularly desirable hosts for expression in this regard include Gram positive bacteria which do not have LPS and are, therefore, endotoxin free. Such hosts include species of *Bacillus* and may be particularly useful for the production of non-pyrogenic lactoferrin receptor proteins, fragments or analogs thereof. Furthermore, recombinant methods of production permit the manufacture of Lbp1 or Lbp2 or ORF3 or respective analogs or fragments thereof, separate from one another which is distinct from the normal combined proteins present in *Moraxella*.

#### Sequence Alignment and Analysis

Sequence alignments were performed using the ALIGN (Trademark) or GENALIGN (Trademark) computer programs (Intelligenetics Suite 5.4, Oxford Molecular). ALIGN® uses the Needleman-Wunsch algorithm (ref. 35) and its later modifications to locate regions of similarity between two sequences. Finding regions of maximum

similarity between two sequences can be solved in a rigorous manner using the iterative matrix calculation of the Needleman and Wunsch 1997 algorithm. The analysis is restricted to regions with no internal deletions or insertions, joined by a minimum number of loop-outs or deletions. Sellers (ref. 36) developed a true metric measure of the "distance" between sequences and Waterman (ref. 37) extended this algorithm to include insertions and deletions of arbitrary length. Smith (ref. 38) improved the early algorithms to find the subsequences of maximum similarity. The algorithm has been used to analyze sequences as long as 5000 bases by dividing these sequences into segments of 200 to 400 bases, and then reassembling them into a final best match. This method of dividing the sequence and then reassembling it has proven quite robust. The algorithm permits the size of the segment to be specified which the program searches for similarities. The program then assembles the segments after checking overlaps of adjacent subsequences. The weighting of deletions and the relative size of overlaps may be controlled. The program displays the results to show the differences in closely related sequences.

GENALIGN® is a multiple alignment program. Up to 99 sequences using the Martinez/Regions (ref. 39) or Needleman-Wunsch (ref. 35) method may be analyzed for alignment. GENALIGN places the sequences in an order that puts the most closely aligned sequence pairs adjacent to each other. A consensus sequence is displayed under the multiple sequence alignments. The sequences used in developing the consensus sequence file for use in other programs. GENALIGN allows the parameters of the search to be changed so that alternate alignments of the sequences can be formed.

These programs are used employing their default settings. The default settings are as follows:

## FastDB

	AMINO-Res-length	=	2
5	DEletion-weight	=	5.00
	Length-factor	=	0
	Matching-weight	=	1.00
	NUCLEIC-Res-length	=	4
	SPread-factor	=	50

## Findseq

Search Parameters:

	Similarity matrix	Unitary
	K-tuple	4
	Mismatch penalty	1
15	Joining Penalty	30
	Randomization group length	0
	Cutoff score	5
	<u>Alignment Parameters:</u>	
	Window size	32
20	Gap penalty	1.00
	Gap size penalty	0.33

Such procedures may be used to determine DNA sequences which encode a functional lactoferrin receptor of *Moraxella* and which may have at least about 90% sequence identity with any one of the DNA sequences (a) or (b), described above.

## Biological Deposits

Certain vectors that contain at least a portion coding for a lactoferrin receptor protein from strains of *Moraxella catarrhalis* strain 4223 and Q8 and a strain of *M. catarrhalis* RH408 that are described and referred to herein have been deposited with the American Type Culture Collection (ATCC) located at 12301 Parklawn Drive, Rockville, Maryland 20852, USA, pursuant to the

Budapest Treaty and prior to the filing of this application. Samples of the deposited vectors and bacterial strain will become available to the public and the restrictions imposed on access to the deposits will be removed upon grant of a patent based upon this United States patent application. In addition, the deposit will be replaced if viable samples cannot be dispensed by the Depository. The invention described and claimed herein is not to be limited in scope by the biological materials deposited, since the deposited embodiment is intended only as an illustration of the invention. Any equivalent or similar vectors or strains that encode similar or equivalent antigens as described in this application are within the scope of the invention.

#### Deposit Summary

Deposit	ATCC Designation	Date deposited
Plasmid pLD1-8	97,997	April 23, 1997
Plasmid pLDW1	97,998	April 23, 1997
Strain RH408	55,637	Dec. 9, 1994

#### EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific Examples. These Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations.

Methods of molecular genetics, protein biochemistry and immunology used but not explicitly described in this disclosure and these Examples are amply reported in the scientific literature and are well within the ability of those skilled in the art.

### Example 1

This Example illustrates the generation of oligonucleotide primers for PCR amplification of *M. catarrhalis lbpA*.

Native Lbp1 was purified by affinity chromatography using high stringency conditions as described in United States Patent Application No. 08/552,232, assigned to the assignee hereof and the disclosure of which is incorporated herein by reference, and in ref. 40.

The purified Lbp1 protein was digested overnight with cyanogen bromide, then fragments separated by SDS PAGE and submitted to sequence analysis on an ABI model 477A. A 13 kDa protein fragment was found to have the N-terminal sequence MVQYTYRKGKENKAH (SEQ ID No: 22). A degenerate oligonucleotide primer (4393.RD) was prepared based upon this sequence:

	Q	Y	T	R	K	G	E	N	K	A	(SEQ ID No: 28)
25	5'										3'
	CAA	TAT	ACI	CGT	AAA	GGT	GAA	AAT	AAA	GC	(SEQ ID No: 29)
	CAA	TAT	ACI	CGT	AAA	GGC	GAA	AAC	AAA	GC	(SEQ ID No: 30)
	CAA	TAT	ACI	CGT	AAA	GGT	GAA	AAC	AAA	GC	(SEQ ID No: 31)
	CAA	TAT	ACI	CGT	AAA	GGC	GAA	AAT	AAA	GC	(SEQ ID No: 32)
30	CAA	TAT	ACI	CGC	AAA	GGC	GAA	AAC	AAA	GC	(SEQ ID No: 33)
	CAA	TAT	ACI	CGC	AAA	GGC	GAA	AAT	AAA	GC	(SEQ ID No: 34)
	CAA	TAT	ACI	CGC	AAA	GGT	GAA	AAT	AAA	GC	(SEQ ID No: 35)
	CAA	TAT	ACI	CGC	AAA	GGT	GAA	AAC	AAA	GC	(SEQ ID No: 36)



The Y<sup>6</sup> and K<sup>10</sup> residues were omitted from the sequence analysis report for the N-terminal sequence and the oligonucleotides used to PCR amplify the 2.2 kb fragment were incorrect, but nevertheless were  
5 successful.

There is a conserved C-terminal pentapeptide found in all known Lbp1 and Tbp1 protein sequences: LEMKF (SEQ ID No. 26). An oligonucleotide primer (4572.RD) was prepared based upon the complementary DNA sequence  
10 encoding this pentapeptide:

L E M K F \*

5' CTT GAA ATG AAG TTT TAA 3' (SEQ ID No: 37)

3' GAA CTT TAC TTC AAA ATT 5' 4572.RD (SEQ ID No: 38)

## 15 Example 2

This Example illustrates the preparation of chromosomal DNA from *M. catarrhalis* strains 4223 and Q8.

*M. catarrhalis* isolate 4223 was inoculated into  
20 100 ml of BHI broth, and incubated for 18 hr at 37°C with shaking. The cells were harvested by centrifugation at 10,000 x g for 20 min. The pellet was used for extraction of *M. catarrhalis* 4223 chromosomal DNA.

25 The cell pellet was resuspended in 20 ml of 10 MM Tris-HCl (pH 7.5)-1.0 mM EDTA (TE). Pronase and SDS were added to final concentrations of 500 µg/ml and 1.0%, respectively, and the suspension was incubated at 37°C for 2 hr. After several sequential extractions  
30 with phenol, phenol:chloroform (1:1), and chloroform:isoamyl alcohol (24:1), the aqueous extract was dialysed, at 4°C, against 1.0 M NaCl for 4 hr, and against TE (pH 7.5) for a further 48 hr with three buffer changes. Two volumes of ethanol were added to  
35 the dialysate, and the DNA was spooled onto a glass

rod. The DNA was allowed to air-dry, and was dissolved in 3.0 ml of water. Concentration was estimated, by UV spectrophotometry, to be about 290 µg/ml.

*M. catarrhalis* strain Q8 was grown in BHI broth.

5 Cells were pelleted from 50 ml of culture by centrifugation at 5000 rpm for 20 minutes, at 4°C. The cell pellet was resuspended in 10 ml of TE (10 mM Tris-HCl, 1 mM EDTA, pH 7.5) and proteinase K and SDS were added to final concentrations of 500 µg/ml and 1%,  
10 respectively. The sample was incubated at 37°C for 4 hours until a clear lysate was obtained. The lysate was extracted twice with Tris-saturated phenol/chloroform (1:1), and twice with chloroform. The final aqueous phase was dialysed for 24 hours  
15 against 2 X 1000 ml of 1 M NaCl at 4°C, changing the buffer once, and for 24 hours against 2 x 1000 ml of TE at 4°, changing the buffer once. The final dialysate was precipitated with two volume of 100% ethanol. The DNA was spooled, dried and resuspended in 5 to 10 ml of  
20 TE buffer.

### Example 3

This Example illustrates the PCR amplification of a fragment of *M. catarrhalis lbpA* and the generation of probes for screening libraries.

25 PCR amplification was performed on chromosomal DNA isolated in Example 2 using primers 4393.RD and 4572.RD under the following cycling conditions: 25 cycles of 94°C for 1 min, 47°C for 30 sec and 72°C for 1 min. PCR4 is the amplification of the 4223 *lbpA* fragment and  
30 PCR5 is the amplification of the Q8 *lbpA* fragment. A specific band of about 2.2 kb was amplified and partial sequence analysis was performed to ensure that the gene product was related to *lbpA* and was not *tbpA*. The derived amino acid sequences are shown in Figure 1 and  
35 have been aligned with the complete 4223 *Lbp1* sequence

to show their placement and the 4223 Tbp1 sequence (USAN 08/613,009) to indicate their uniqueness.

The full-length 2.2 kb gene fragment was randomly labeled with  $^{32}\text{P}$  and used to probe genomic libraries.

5     **Example 4**

This Example illustrates the generation and screening of the EMBL 3 libraries.

Chromosomal DNA was prepared as described in Example 2. A series of Sau3AI restriction digests of  
10     chromosomal DNA, in final volumes of 10  $\mu\text{L}$  each, were carried out in order to optimize the conditions necessary to generate maximal amounts of restriction fragments within a 15 to 23 kb size range. Using the optimized digestion conditions, a large-scale digestion  
15     was set up in a 100  $\mu\text{L}$  volume, containing the following: 50  $\mu\text{L}$  of chromosomal DNA (290  $\mu\text{g}/\text{ml}$ ), 33  $\mu\text{L}$  water, 10  $\mu\text{L}$  10X Sau3A buffer (New England Biolabs), 1.0  $\mu\text{L}$  BSA (10  $\text{mg}/\text{ml}$ , New England Biolabs), and 6.3  $\mu\text{L}$  Sau3A (0.04 U/ $\mu\text{L}$ ). Following a 15 min. incubation at  
20     37°C, the digestion was terminated by the addition of 10  $\mu\text{L}$  of 100 mM Tris-HCl (pH 8.0)-10 mM EDTA-0.1% bromophenol blue-50% glycerol (loading buffer). Digested DNA was electrophoresed through a 0.5% agarose gel in 40 mM Tris acetate-2 mM Na<sub>2</sub>EDTA.2H<sub>2</sub>O (pH 8.5) (TAE  
25     buffer) at 50 V for 6 hr. The region containing restriction fragments within a 15 to 23 kb molecular size range was excised from the gel, and placed into dialysis tubing containing 3.0 ml of TAE buffer. DNA was electroeluted from the gel fragment by applying a  
30     field strength of 1.0 V/cm for 18 hr. Electroeluted DNA was extracted once each with phenol and phenol:chloroform (1:1), and precipitated with ethanol. The dried DNA was dissolved in 5.0  $\mu\text{L}$  water.

Size-fractionated chromosomal DNA was ligated with

*Bam*HI-digested EMBL3 arms (Promega), using T4 DNA ligase in a final volume of 9  $\mu$ L. The entire ligation mixture was packaged into lambda phage using a commercial packaging kit (Amersham), following manufacturer's instructions.

The packaged DNA library was amplified on solid media. 0.1 ml aliquots of *Escherichia coli* strain NM539 in 10 mM  $\text{MgSO}_4$  ( $\text{OD}_{260} = 0.5$ ) were incubated at 37°C for 15 min. with 15 to 25  $\mu$ L of the packaged DNA library. Samples were mixed with 3 ml of 0.6% agarose containing 1.0% BBL trypticase peptone-0.5% NaCl (BBL top agarose), and mixtures were plated onto 1.5% agar plates containing 1.0% BBL trypticase peptone-0.5% NaCl, and incubated at 37°C for 18 hr. 3 ml quantities of 50 mM Tris-HCl (pH 7.5)-8 mM magnesium sulfate heptahydrate-100 mM NaCl-0.01% (w/v) gelatin (SM buffer) were added to each plate, and plates were left at 4°C for 7 hr. SM buffer containing phage was collected from the plates, pooled together, and stored in a screwcap tube at 4°C, with chloroform.

Ten  $\mu$ L aliquots of phage stock were combined each with 100  $\mu$ L of *E. coli* strain LE392 in 10 mM  $\text{MgSO}_4$  ( $\text{OD}_{260} = 0.5$ ) (plating cells), and incubated at 37°C for 15 min. The samples were mixed with 3 ml each of BBL top agarose, and the mixtures were poured onto 1.5% agarose plates containing 1% bacto tryptone-0.5% bacto yeast extract-0.05% NaCl (LB agarose; Difco) and supplemented with 200  $\mu$ M EDDA. The plates were incubated at 37°C for 18 hr. Plaques were lifted onto nitrocellulose filters (Amersham Hybond-C Extra) which were hybridized with the  $^{32}\text{P}$ -labelled 2.2 kb PCR fragment. Several putative phage clones were obtained from each library and clones 4223LfR.17 and Q8LfR.13 were chosen for further analysis.

**Example 5**

This Example illustrates the subcloning of the phage clones containing *M. catarrhalis lfr* genes.

5        Restriction enzyme analysis and Southern blotting using the screening probes, indicated that at least a portion of *lbpA* was localized to an about 9 kb Hind III fragment from each phage clone. The about 9 kb Hind III fragment from 4223Lfr.17 was subcloned into pUC 18, 10        generating clone pLD1-8. The about 9 kb Hind III fragment from Q8Lfr.13 was subcloned into pBluescript, generating plasmid pLDW1. Internal about 5.5 kb EcoR V fragments were subcloned generating plasmids pLD3 and pLDW3 for the 4223 and Q8 genes, respectively.

15        **Example 6**

This Example illustrates the sequence analysis of clones containing the *M. catarrhalis lfr* genes from strains 4223 and Q8.

20        Sequence analysis of the 5.5 kb EcoR V fragments from pLD3 and pLDW3, revealed that they each contained the 3'-end of *lbpB*, the complete *lbpA* gene, and a third complete gene designated *orf3*. The remainder of the *lbpB* genes was found on the about 9 kb Hind III fragments from pLD1-8 and pLDW1. Partial restriction 25        enzyme analysis of the 4223 *lbpA*, *lbpB*, and *orf3* genes, based upon the nucleotide sequences is shown in Figure 3. Partial restriction enzyme analysis of the Q8 *lbpA*, *lbpB*, and *orf3* genes, based upon the nucleotide sequences is shown in Figure 5. The complete sequences 30        of the *lbpB*, *lbpA*, and *orf3* genes comprising the putative *lfr* locus from *M. catarrhalis* 4223 and Q8 is shown in Figures 2 and 4, respectively. The intergenic distance between the *lbpB* and *lbpA* genes is 184 nucleotides, while a single nucleotide separates the

*lbpA* and *orf3* genes. A putative promoter and ribosome binding site is indicated by underlining upstream of both *lbpB* and *lbpA*. A fourth potential gene was cloned on the approximately 9 kb Hind III fragments.

5       The N-terminal sequence of the native Lbp1 protein is unknown. Examination of the deduced amino acid sequence of the *lbpA* gene indicates that there are two possible ATG start codons at positions 1 and 16. The first position is downstream of strong promoter  
10       elements found in the *lbpB-lbpA* intergenic region and the second position is followed by a putative signal sequence. The *M. catarrhalis* 4223 and Q8 Lbp1 proteins (from the first ATG) have molecular mass values of about 110 kDa and are 99% identical. The deduced Lbp1  
15       protein sequences from *M. catarrhalis* strains 4223 and Q8 are compared in Figure 6. They are also compared with the *iroA/lbpA* gene from *N. meningitidis* strain BNCV (ref. 24) and the *lbpA* gene from *N. gonorrhoeae* strain FA19 (ref. 25). The *M. catarrhalis* proteins are  
20       found to be about 32% identical and about 50% similar to the *Neisseria* proteins. As shown in Figure 1, there is very limited sequence homology between the *M. catarrhalis* Tbp1 and Lbp1 sequences.

      The deduced Lbp2 protein sequences from *M.*  
25       *catarrhalis* strains 4223 and Q8 are compared in Figure 7. The 4223 and Q8 Lbp2 proteins both have molecular masses of about 99 kDa and are 92% identical and 95% similar to each other. A comparison to the *M. catarrhalis* Tbp2 proteins shows very little homology  
30       except the LEGGFY (SEQ ID No: 27) epitope previously identified in *H. influenzae* and *N. meningitidis* Tbp2 proteins (Fig. 8). A cysteine residue at position 32 is preceded by a consensus sequence for lipoproteins suggesting that Lbp2, like Tbp2, is a lipoprotein. An



unusual feature of the Lbp2 proteins is the high combined aspartic acid and asparagine content which is nearly 20%. In addition, the 4223 Lbp2 amino acid composition from residues 698 to 751 is about 52% aspartic acid.

The 4223 and Q8 *lfr orf3* genes would encode proteins of molecular mass about 60 kDa, respectively. A notable feature of the ORF3 protein is a potential signal sequence, a terminal phenylalanine which is often associated with membrane anchored proteins, an internal repeat sequence of DGLG (SEQ ID No: 39), and a high leucine content of 15%. The deduced Lbp3 protein sequences are compared in Figure 9. These proteins are 98% identical and 99% similar.

#### Example 7

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp1 from the first methionine in *E. coli*.

There are two possible start codons at the beginning of the *lbpA* gene and hence two expression constructs were made. The construction scheme for 4223 or Q8 *lbpA* expressed from the first methionine is shown in Figure 10. An approximately 200 bp fragment of the 5'-end of *lbpA* from the ATG to a BstE II site was PCR amplified using primers 5405.RD and 5407.RD. An Nde I site was engineered at the 5'-end to facilitate cloning into the pT7-7 vector.

NdeI

30 5' M S K S I T (SEQ ID No: 40)  
GGAATTCCAT ATG TCA AAA TCT ATC ACA AA 3' 5405.RD  
(SEQ ID No: 41)

BstE II

L D A I T V T A A (SEQ ID No: 42)  
5' T TTA GAT GCC ATC ACG GTA ACC GCC GCC CC 3' (SEQ ID No: 43)

3' A AAT CTA CGG TAG TGC CAT TGG CGG CGG GG 5' 5407.RD

(SEQ ID No: 44)

5 In order to subclone the *lbpA* gene into pT7-7, a approximately 515 bp fragment of the 3'-end of the gene from an Sph I site to the stop codon was PCR amplified using primers 5281.RD and 5282.RD and a BamH1 site was engineered at 3'-end.

Sph I

10 G K L D L H A M T S (SEQ ID No: 45)

5' GGC AAA CTG GAT TTG CAT GCC ATG ACA TCA 3' 5281.RD

(SEQ ID No: 46)

S L E M K F \* (SEQ ID No: 47)

15 5' AGT CTT GAA ATG AAG TTT TAA 3'

(SEQ ID No: 48)

3' TCA GAA CTT TAC TTC AAA ATT GCC CTA GGG C 5' 5282.RD

BamH I (SEQ ID No: 49)

20 For the Q8 subclone, plasmid pLDW3, prepared as described in Example 5, was digested with BstE II and Sph I generating a 2.3kb fragment of *lbpA* which was ligated with the Nde I-BstE II and SphI-BamH I PCR fragments and cloned into pT7-7 digested with NdeI and  
25 BamH I. The resulting plasmid pQW1A thus contains the full-length Q8 *lbpA* gene from the first methionine, under the control of the T7 promoter. DNA from pQW1A was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate strain  
30 QW1A which was grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 protein was visualized by Coomassie blue staining (Fig. 11).

35 For the 4223 subclone, plasmid pLD3, prepared as described in Example 5 was digested with BstEII and SphI, generating a 2.3 kb fragment of *lbpA*, which was

ligated with the Nde I-BstE II and SphI-BamH I PCR fragments and cloned into pT7-7 digested with NdeI and BamH I. The resulting plasmid pRD1A thus contains the full-length 4223 *lbpA* gene from the first possible methionine under the control of the T7 promoter. DNA from pRD1A was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate strain RD1A which was grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 protein was visualized by Coomassie blue staining (Fig. 11).

The Q8 Lbp1 protein was expressed at very high levels but the 4223 Lbp1 protein was expressed at substantially lower levels.

#### Example 8

This Example illustrates the extraction and purification of rLbp1 from *E. coli*. The procedure is illustrated generally in Figure 14.

*E. coli* cells from a 500 ml culture, prepared as described in Example 7, were resuspended in 40 ml of 50 mM Tris-HCl, pH 8.0 containing 5 mM AEBSF (protease inhibitor) and 0.1 M NaCl, and disrupted by sonication (3 x 10 min, 70% duty cycle). The extract was centrifuged at 20,000 x *g* for 30 min and the resultant supernatant, which contained greater than 95% of the soluble proteins from *E. coli*, was discarded. The remaining pellet (Figure 14, PPT1) was further extracted in 40 ml of 50 mM Tris, pH 8.0 containing 0.5% Triton X-100 and 10 mM EDTA. The mixture was stirred at 4°C for at least 1 hour and then centrifuged at 20,000 x *g* for 30 min and the supernatant containing residual soluble proteins and the majority of the membrane proteins was discarded. The resultant pellet (Figure 14, PPT2) was further extracted in 40 ml of 50

5 mM Tris, pH 8.0 containing 1% octylglucoside. The mixture was stirred at 4°C for at least 1 hour and then centrifuged at 20,000 x g for 30 min. The supernatant containing residual contaminating proteins was discarded. The resultant pellet (Figure 14, PPT3) obtained after the above extractions contained the Lbp1 protein as inclusion bodies.

The rLbp1 protein was solubilized from the inclusion bodies in 50 mM Tris, pH 8.0, containing 6 M guanidine and 5 mM DTT. After centrifugation, the resultant supernatant was further purified on a Superdex 200 gel filtration column equilibrated in 50 mM Tris-HCl, pH 8.0, containing 2 M guanidine and 5 mM DTT. The fractions were analysed by SDS-PAGE and those containing purified rLbp1 were pooled. Triton X-100 was added to the pooled rLbp1 fraction to a final concentration of 0.1%. The fraction was dialysed overnight at 4°C against PBS, and then centrifuged at 20,000 x g for 30 min. The purified rLbp1 was stored at -20°C. Samples from the purification were analyzed by SDS-PAGE (Fig. 15).

#### Example 9

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp1 from the second methionine in *E. coli*.

The construction scheme for 4223 or Q8 *lbpA* expressed from the second methionine is shown in Figure 10. An approximately 200 bp fragment of the 5'-end of *lbpA* from the ATG to a BstE II site was PCR amplified using primers 5406.RD and 5407.RD. An Nde I site was engineered at the 5'-end to facilitate cloning into the pT7-7 vector.

NdeI

M T T H R L

(SEQ ID No: 50)

5' GGAATTCCAT ATG ACC ACG CAC CGC TTA AA 3' 5406.RD

(SEQ ID No: 51)

#### BstE II

5 L D A I T V T A A

5' T TTA GAT GCC ATC ACG GTA ACC GCC GCC CC 3'

3' A AAT CTA CGG TAG TGC CAT TGG CGG CGG GG 5' 5407.RD

The 3'-end of the *lbpA* gene was PCR amplified from the SphI restriction site to the stop codon using primers 5281.RD and 5282.RD as described in Example 8. The 2.3 kb BstE II-Sph I fragments described in Example 8 were ligated to the Nde I-BstE II and Sph I-BamH I PCR fragments and cloned into pT7-7 that had been digested with NdeI and BamH I. Plasmid pQW1B thus contains a full-length Q8 *lbpA* gene from the second methionine and plasmid pRD1B contains a full-length 4223 *lbpA* gene from the second methionine under the direction of the T7 promoter. DNA was purified and transformed by electroporation into electrocompetent BL21(DE3) cells to generate recombinant strains which were grown and induced using IPTG. Expressed proteins were resolved by SDS-PAGE and the induced Lbp1 proteins were visible by Coomassie blue staining (Fig. 11).

As seen for the longer protein in Example 8, the shorter Lbp1 from Q8 was expressed to much higher levels than the corresponding 4223 protein.

#### Example 10

This Example illustrates the construction of vectors to express *M. catarrhalis* Lbp2 with a leader sequence from *E. coli*.

The construction scheme is illustrated in Figure 12. There are two BspH I sites within the *lbpB* genes of strains 4223 and Q8. The 5'-end of the *lbpB* gene was PCR amplified from the ATG start codon through the

first BspH I site generating an approximately 201 bp fragment. An NdeI site was engineered at the ATG to facilitate cloning into the pT7-7 expression vector. The oligonucleotides used for amplification are illustrated below:

## NdeI

M S T V K T P H (SEQ ID No: 52)

5' GGAATTCCAT ATG AGT ACT GTC AAA ACC CCC CAC A 3' 5533.RD  
(SEQ ID No: 53)

10

## BspH I

I P N T G H D N T N (SEQ ID No: 54)

5' A ATA CCG AAC ACA GGT CAT GAC AAC ACC AAT 3'  
(SEQ ID No: 55)

T TAT GGC TTG TGT CCA GTA CTG TTG TGG TTA 5' 5534.RD

15

(SEQ ID No: 56)

The 3'-end of the *lbpB* gene was PCR amplified from the second BspH I site to the TAA stop codon generating a 381 bp fragment. A BamH I site was introduced after the stop codon for cloning purposes. The oligonucleotides used for amplification are illustrated below:

N E P T H E K T F A (SEQ ID No: 57)

5' AAT GAG CCT ACT CAT GAA AAA ACC TTT GCC 3' 5535.RD

25

(SEQ ID No: 58)

G A V F G A V K D K \* (SEQ ID No: 59)

5' GG GCT GTC TTT GGG GCT GTT AAA GAT AAA TAA 3'

30

(SEQ ID No: 60)

CC CGA CAG AAA CCC CGA CAA TTT CTA TTT ATT CCTAGGGC 5' 5536.RD

BamH I (SEQ ID No: 61)

Plasmids pLD1-8 or pLDW1, prepared as described in Example 4, were digested with BspH I to release a 2.1 kb internal fragment of the *lbpB* gene which was ligated with the 5'- and 3'-PCR fragments and cloned into pT7-7



that had been digested with NdeI and BamH I. The resulting plasmids, pLD2A and pLDW2A, contain the full-length 4223 and Q8 *lbpB* genes under the control of the T7 promoter, respectively.

#### 5 **Example 11**

This Example illustrates the construction of vectors to express the mature *M. catarrhalis* Lbp2 proteins from *E. coli*.

The construction scheme is illustrated in Figure 12. The putative mature Lbp2 lipoproteins start at the Cys<sup>32</sup> residue. A scheme similar to that described in Example 10 can be used to generate expression clones. To amplify the 5'-end of the *lbpB* gene, a sense PCR primer is designed that includes an NdeI site for subsequent cloning and an ATG start codon for initiation of translation followed immediately by the Cys<sup>32</sup> residue. The antisense primer is the same as that described in Example 9 (5534.RD) and includes the BspH I cloning site. The amplified fragment is ~112 bp long. The oligonucleotides are illustrated below:

NdeI

M C R S D D I S V N

(SEQ ID No: 62)

5' GGAATTCCAT ATG TGC CGC TCT GAT GAC ATC AGC GTC AAT 3' .RD

(SEQ ID No: 63)

BspH I

I P N T G H D N T N (SEQ ID No: 54)

5' A ATA CCG AAC ACA GGT CAT GAC AAC ACC AAT 3'

(SEQ ID No: 55)

3' T TAT GGC TTG TGT CCA GTA CTG TTG TGG TTA 5' 5534.RD

(SEQ ID No: 56)

The BspH I-BamH I 3'-end of the *lbpB* gene is PCR amplified as in Example 9 and the plasmid expressing

mature Lbp2 is constructed by ligating the 5'- and 3'- PCR fragments with the 2.1 kb BspH I fragment and vector pT7-7 digested with NdeI and BamH I. The resulting plasmids, pLD2B and pLDW2B, contain the *lbpB* gene encoding the mature Lbp2 proteins from strains 4223 and Q8 under the direction of the T7 promoter, respectively.

### Example 12

This Example illustrates the construction of a vector to express the *M. catarrhalis* *lfr* Lbp3 from *E. coli*.

The construction scheme is illustrated in Figure 13. Oligonucleotides were used to generate the 5'-end of the *orf3* gene from the ATG start codon to an AlwN I site. An NdeI site was engineered at the 5'-end for subsequent cloning into pT7-7. The oligonucleotides are shown below:

```

NdeI
20      M   T   C   L   P   K   T   N   P   A   L   K   V   K   H   R
5'  T ATG ACC TGT TTA CCA AAG ACC AAC CCT GCT TTA AAA GTC AAG CAC AGA
3'      AC TGG ACA AAT GGT TTC TGG TTG GGA CGA AAT TTT CAG TTC GTG TCT

      AlwN I
      F   L   K   Q   V
25      TTT TTA AAG CAG GTG      3'      5532.RD      (SEQ ID No: 65)
      AAA AAT TTC GTC      5'      5457.RD      (SEQ ID No: 66)

```

The pLD1-8 or pLDW1 plasmid, prepared as described in Example 5, was digested with BstE II generating a 4.6 kb fragment which was filled in with Klenow polymerase before being digested with AlwNI. The resultant 1.8 kb fragment was ligated with the annealed NdeI-AlwN I oligonucleotides and cloned into pT7-7 that had been digested with NdeI and SmaI. The resulting plasmids, pLRD3 and pLQW3, contain the full-length *orf3*

genes from strains 4223 and Q8 under the direction of the T7 promoter, respectively.

### Example 13

5 This Example describes the cloning and sequencing of the *lbpB* gene from *M. catarrhalis* strain VH19.

Chromosomal DNA was prepared from *M. catarrhalis* strain VH19, as described previously in Example 2. Oligonucleotide primers were designed based upon the flanking sequence of the 4223 *lbpB* gene. The sense  
10 primer was 5' AAGCTTAGCATGATGGCATCGGCT 3' (SEQ ID No: 67) and the antisense primer was 5' TTAGCCCAAGGCAAATCTGGTGCA 3' (SEQ ID No: 68). PCR was performed in buffer containing 10mM Tris-HCl (pH 8.3), 50 mM potassium chloride and 1.5 mM magnesium chloride.  
15 Each 100 µl reaction mixture contained 1 µg chromosomal DNA, 0.1 µg each primer, 2.5 units amplitaq DNA polymerase (Perkin Elmer Cetus, Foster City, California) and 10 mM of each dNTP (Perkin Elmer Cetus). The cycling conditions were 24 cycles of 94°C  
20 for 1 min, 47°C for 30 sec and 72°C for 1 min. Specific 2.9 kb fragments were amplified from two independent reactions and subcloned into pCR II (Invitrogen, Carlsbad, California), generating plasmids pVH19pcr1 and pVH19pcr2 for sequence analysis. A third  
25 PCR amplification was performed without subcloning the resultant DNA. Plasmid DNA from pVH19pcr1 and pVH19pcr2 was prepared from 50 ml overnight cultures using the Qiagen Plasmid Midi kit (Qiagen Inc, Chatsworth, California). PCR amplified DNA was  
30 purified for direct sequencing using a Qiagen PCR purification kit. DNA samples were sequenced on an ABI model 373A DNA sequencer using dye terminator chemistry. Oligonucleotide primers 17 to 25 bases in length were used to sequence both strands of the DNA.

The nucleotide sequence (SEQ ID No: 69) of the VH19 *lbpB* gene and the deduced amino acid sequence of the corresponding Lbp2 protein (SEQ ID No: 70) are shown in Figure 16. The encoded VH19 Lbp2 protein is 906 amino acids and is 77% identical and 84% similar to the 4223 and Q8 Lbp2 proteins. There is a putative lipoprotein signal sequence which is very similar to the 4223 and Q8 signal sequences. The high Asp and Asn content found in the 4223 and Q8 Lbp2 proteins is also present in the VH19 LbpB protein, as is the RGD sequence. A partial restriction map of the VH19 *lbpB* gene is shown in Figure 17.

An alignment of the Lbp2 proteins from *M. catarrhalis* strains 4223, Q8 and VH19 is shown in Figure 7. The *M. catarrhalis* Lbp2 proteins are also compared with partial Lbp2 sequences from *N. meningitis* strains BNCV (ref. 31) and H44/76 (ref. 24) and *N. gonorrhoeae* strain FA19 (ref. 25). There are small scattered regions of sequence homology to the known bacterial Tbp2 proteins (ref. 32). Residues that are conserved among the Tbp2 proteins and the *M. catarrhalis* Lbp2 proteins are underlined in Figure 7 and include the LEGGFYG (SEQ ID No: 75) motif.

#### Example 14

This Example describes the construction of vectors for expression of the *M. catarrhalis* Lbp2 protein.

By analogy with Tbp2 proteins, Lbp2 was assumed to be a lipoprotein and constructs were designed for expression of Lbp2 with or without a lipopeptide signal sequence. There is a unique Bgl I site in *lbpB*. To express the full-length Lbp2 protein with leader sequence (construct A), an approximately 429 bp 5'-fragment from the Met<sup>1</sup> start codon to the Bgl I site was PCR amplified and to express the mature protein

(construct B), an approximately 329 bp 5'-fragment from the putative Cys<sup>32</sup> start to the Bgl I site was PCR amplified. The following sense primers were used:

Nde I

5 M S T V K T P H (SEQ ID No: 52)

5' GGAATTCCAT ATG AGT ACT GTC AAA ACC CCC CAC A 3' (SEQ ID No: 53)

for construct A or

Nde I

10 M C R S D D I S V N (SEQ ID No: 62)

5' GGAATTCCAT ATG TGC CGC TCT GAT GAC ATC AGC GTC AAT 3' (SEQ ID No: 63)

for construct B and the anti-sense primer was:

15 G K N L R G P I (SEQ ID No: 72)

5' GGT AAA AAC TTG CGT CAG CCC ATC 3' (SEQ ID No: 73)

3' CCA TTT TTG AAC GCA GTC GGG TAG 5' (SEQ ID No: 74)

Bgl I

20 The Q8 *lfr*-containing plasmid, pLDW1 (Example 5), was digested with Bgl I and EcoR I to release a 2.3 kb *lbpB* fragment which was ligated with the Nde I - Bgl I PCR fragment and cloned into pT7-7 that had been digested with Nde I and EcoR I. The resulting plasmids, pQW2A and pQW2B, thus contain the Q8 *lbpB* gene encoding the full-length or mature Lbp2 proteins under the direction of the T7 promoter. The plasmids expressing the 4223 full-length or mature Lbp2 proteins were constructed in a similar manner and designated pRD2A and pRD2B. There was no measurable expression of rLbp2 from constructs containing the signal sequence, however the mature rLbp2 proteins were expressed at 5 to 10% of total proteins as inclusion bodies and were purified by the same process as that described for rLbp1 in Example 8. Samples from the purification were analyzed by SDS-PAGE (Figure 18).

**Example 15**

This Example describes the functional characterization of the recombinant lactoferrin binding proteins.

5 Human lactoferrin (Sigma) was conjugated to horseradish peroxidase using an EZ-Link maleimide activated horseradish peroxidase (HRP) kit (Pierce, Rockford, Illinois) according to the manufacturer's instructions. The lactoferrin binding activity of  
10 rLbp1 or rLbp2 was assessed by modifying the procedure described for transferrin binding proteins (ref. 17). Briefly, purified rLbp1 or rLbp2 was subjected to discontinuous electrophoresis through a 12.5% SDS PAGE gel. The proteins were electrophoretically transferred  
15 to a polyvinylidene difluoride (PVDF) membrane (Millipore, Bedford, Massachusetts) and incubated with horseradish peroxidase-conjugated human lactoferrin (1:20 dilution) at 4°C overnight. LumiGLO substrate (Kirkegaard and Perry Laboratories, Inc., Gaithersburg,  
20 Maryland) was used for chemiluminescent detection of HRP activity according to the manufacturer's instructions. The Q8 rLbp1 protein did not bind human lactoferrin under these conditions, but the 4223 rLbp2 and Q8 rLbp2 proteins did (Fig. 19).

**25 Example 16**

This Example describes the immunization of animals and immunoassays.

Groups of two guinea pigs (Hartley outbred, Charles River, Quebec) were immunized intramuscularly (i.m.)  
30 with 5 µg doses of purified rLbp1 or rLbp2 protein emulsified in CFA or IFA. Anti-Lbp antibody titers in guinea pig immune sera were determined by antigen-specific ELISA. Microtiter wells (Nunc-MAXISORB, Nunc, Denmark) were coated with 50 µl of protein (0.5 µg ml<sup>-1</sup>



1). The reactive titer of an antiserum was defined as the reciprocal of the highest dilution consistently showing a two-fold increase in absorbance at 450 nm over that obtained with the pre-immune serum samples. The recombinant proteins elicited high titre antibodies as shown in Tables 1 and 2.

#### Example 17

This Example describes the antigenic conservation of Lbp1 and Lbp2 in *M. catarrhalis* strains.

To demonstrate the iron-dependent expression of the *lbpA* and *lbpB* genes, representative *M. catarrhalis* strains were grown in BHI  $\pm$  25 mM EDDA. Whole cell lysates were separated by SDS PAGE and electrophoretically transferred to nitrocellulose membrane. Guinea pig anti-Q8 rLbp1, anti-Q8 rLbp2 and anti-4223 rLbp2 antisera were used as first antibodies and horseradish peroxidase-conjugated protein G (ZYMED) was used as secondary antibody. To assess antigenic conservation, approximately 90 *M. catarrhalis* strains, obtained from North America or Finland were grown in BHI + 25 mM EDDA, and immunoblots were probed with guinea pig anti-4223 rLbp2 antibody, as above. All strains showed a protein band reactive with anti-rLbp2 antibody. There was very little size heterogeneity for the Lbp2 proteins from the 90 *M. catarrhalis* strains, ranging from approximately 100 kDa to 105 kDa. Representative immunoblots are illustrated in Fig. 19.

#### Example 18

This Example describes the assay used to determine the bactericidal antibody activity of anti-Lbp antibodies.

The assay was performed as described by ref. 33. Briefly, the *M. catarrhalis* strains were grown to an OD<sub>578</sub> of 0.5 in BHI medium containing 25 mM EDDA. The

bacteria were diluted so that the pre-bleed control plates contained 100 to 300 cfu. Guinea pig anti-rLbp1 or anti-rLbp2 antisera and pre-bleed controls, were heated to 56°C for 30 min to inactivate endogenous complement and were diluted 1:64 with veronal buffer containing 0.1% BSA (VBS). Guinea pig complement (Biowhittaker, Walkersville, Maryland) was diluted 1:10 in VBS. Twenty-five µl each of diluted antiserum, bacteria and complement were added to duplicate wells of a 96 well microtiter plate (Nunc). The plates were incubated at 37°C for 60 min, gently shaking at 70 rpm on a rotary platform. Fifty µl of each reaction mixture were plated onto Mueller Hinton agar plates (Becton-Dickinson, Cockeysville, Maryland) which were incubated at 37°C for 24 h, then room temperature for 24 h, before the bacteria were counted. Antisera were determined to be bactericidal if  $\geq 50\%$  of bacteria were killed compared with negative controls.

Six strains of different geographical and anatomical origins were tested. The data in Table 3 illustrates that anti-4223 rLbp2 antibody was bactericidal for the homologous strain and three of five heterologous strains.

#### SUMMARY OF THE DISCLOSURE

In summary of this disclosure, the present invention provides purified and isolated DNA molecules containing lactoferrin receptor genes from *Moraxella catarrhalis*, the sequences of these lactoferrin receptor genes, and the derived amino acid sequences thereof. The genes and DNA sequences are useful for diagnosis, immunization, and the generation of diagnostic and immunological reagents. Immunogenic compositions, including vaccines, based upon expressed recombinant Lbp1 and/or Lbp2 and/or ORF3, portions

thereof, or analogs thereof, can be prepared for prevention of diseases caused by *Moraxella*. Modifications are possible within the scope of this invention.

Table 1

Bactericidal antibody titres for anti-native Lbp1

Antibody	Bactericidal titre - RH408		Bactericidal titre - Q8	
	Pre-immune	Immune	Pre-immune	Immune
Anti-4223 Lbp1	<8	114-330	<8	128-512

Bactericidal titres are expressed as the reciprocal dilution of antiserum capable of killing 50% of *M. catarrhalis* cells

Table 2 ELISA titers for guinea pig anti-Lbp antibodies raised against recombinant lactoferrin binding proteins

Coated antigen	Anti-Q8 rLbp1	Anti-Q8 rLbp2	Anti-4223 rLbp2
Q8 rLbp1	3,200 25,600	-	-
Q8 rLbp2	-	1,638,400 1,638,400	409,600 409,600
4223 rLbp2	-	409,600 409,600	819,200 819,200

Table 3. Bactericidal antibody activity of guinea pig anti-rLbp2 antibodies

Strain	locale <sup>1</sup>	source <sup>2</sup>	Lbp2 size	Bactericidal antibody activity <sup>3</sup>	
				Anti-4223 rLbp2	Anti-Q8 rLbp2
4223	New York	MEF	105 kDa	++	-
Q8	Quebec	sputum	105 kDa	±	-
VH19	Texas	MEF	105 kDa	+	NT <sup>4</sup>
LES-1	Finland	MEF	102 kDa	-	NT
H-04	Nova Scotia	MEF	100 kDa	+	NT
3	New York	sputum	100 kDa	++	NT

<sup>1</sup> geographic locale where strain was isolated

<sup>2</sup> anatomical source of clinical isolate. MEF is middle ear fluid from otitis media patients

<sup>3</sup> killing by antiserum diluted 1:64, compared to negative controls: - indicates 0-25% killing; ± indicates 26-49% killing; + indicates 50-75% killing; ++ indicates 76-100% killing.

<sup>4</sup> NT = not tested



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CLAIMS

What we claim is:

1. A purified and isolated nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of the lactoferrin receptor protein.
2. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the lactoferrin receptor binding protein 1 (Lbp1) of the *Moraxella* strain.
3. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the lactoferrin receptor binding protein 2 (Lbp2) of the *Moraxella* strain.
4. The nucleic acid molecule of claim 1 wherein the lactoferrin receptor protein is the open reading frame protein 3 (ORF3) of the *Moraxella* strain.
5. The nucleic acid molecule of claim 1 wherein the strain of *Moraxella* is a strain of *Moraxella catarrhalis*.
6. The nucleic acid molecule of claim 5 wherein the strain of *Moraxella catarrhalis* is *Moraxella catarrhalis* 4223, Q8 or VH16.
7. A purified and isolated nucleic acid molecule encoding at least one lactoferrin binding protein of *Moraxella* and having a restriction map as shown in Figure 3 for *M. catarrhalis* 4223, Figure 5 for *M. catarrhalis* Q8 or Figure 17 for *M. Catarrhalis* VH19 or the equivalent map for another strain of *M. catarrhalis*.
8. A purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of:

- (a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto;
  - (b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and
  - (c) a DNA sequence encoding a functional lactoferrin receptor protein of *Moraxella*.
9. The nucleic acid molecule of claim 8, wherein the DNA sequence defined in (c) hybridizes under stringent conditions to any one of the sequences defined in (a) or (b).
10. The nucleic acid molecule of claim 8, wherein the DNA sequence defined in (c) is that encoding the equivalent lactoferrin receptor protein from another strain of *Moraxella*.
11. A vector adapted for transformation of a host comprising the nucleic acid molecule of claim 1 or 8.
12. The vector of claim 11 encoding at least a fragment of a lactoferrin receptor protein and having the characteristics of a plasmid selected from the group consisting of pLD3, pLDW3, pLD1-8 (ATCC 97,997), pLDW1 (ATCC 97,998), pVH19pcr1 and pVH19pcr2.
13. The vector of claim 11 further comprising expression means operatively coupled to the nucleic acid molecule for expression of said lactoferrin receptor protein of a strain of *Moraxella* or the fragment or the analog of the lactoferrin receptor protein by the host containing the vector.
14. The vector of claim 13 having the characteristics of plasmid pRD1A, pRD1B, pQW1A, pQW1B, pRD2B, pQW2B, pLRD3 and pLQW3.



15. A transformed host containing an expression vector as claimed in claim 13.

16. A method of forming a substantially pure recombinant lactoferrin receptor protein, which comprises:

growing the transformed host of claim 15 to express a lactoferrin receptor protein as inclusion bodies,

purifying the inclusion bodies free from cellular material and soluble proteins,

solubilizing lactoferrin receptor protein from the purified inclusion bodies, and

purifying the lactoferrin receptor protein free from other solubilized materials.

17. The method of claim 16 wherein said lactoferrin receptor protein comprises Lbp1 alone, Lbp2 alone, ORF3 alone or a mixture of two or more of Lbp1, Lbp2 and ORF3.

18. The method of claim 17 wherein said lactoferrin receptor protein is at least about 70% pure.

19. The method of claim 18 wherein said lactoferrin receptor protein is at least about 90% pure.

20. A recombinant lactoferrin receptor protein or fragment or analog thereof producible by the transformed host of claim 14.

21. The protein of claim 20 which is lactoferrin receptor binding protein 1 (Lbp1) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

22. The protein of claim 20 which is lactoferrin receptor binding protein 2 (Lbp2) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

23. The protein of claim 20 which is open reading frame protein 3 (ORF3) of the *Moraxella* strain devoid of other proteins of the *Moraxella* strain.

24. The protein of claim 20 wherein the strain of *Moraxella* is a strain of *Moraxella catarrhalis*.

25. An open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the lactoferrin binding protein.

26. The protein of claim 25 wherein said *Moraxella* strain is a strain of *M. catarrhalis*.

27. The protein of claim 26 wherein said *M. catarrhalis* strain is *M. catarrhalis* 4223 or Q8.

28. The protein of claim 24 having a deduced amino acid as set forth in Figure 2 or 4 (SEQ ID No: 14, 18).

29. An immunogenic composition, comprising at least one active component selected from the group consisting of:

(A) a purified and isolated nucleic acid molecule encoding a lactoferrin receptor protein of a strain of *Moraxella* or a fragment or an analog of lactoferrin receptor protein;

(B) a purified and isolated nucleic acid molecule having a DNA sequence selected from the group consisting of:

(a) a DNA sequence as set out in Figure 2 or 4 (SEQ ID Nos. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 69) or the complementary DNA sequence thereto;

(b) a DNA sequence encoding an amino acid sequence as set out in Figure 2 or 4 (SEQ ID Nos. 11, 12, 13, 14, 15, 16, 17, 18, 70) or the complementary DNA sequence thereto; and

(c) a DNA sequence encoding a functional lactoferrin receptor protein of *Moraxella*;

(C) a recombinant lactoferrin receptor protein or fragment or analog thereof producible by a transformed host containing an expression vector comprising a nucleic acid molecule as defined in (A) or (B) and expression means operatively coupled to the nucleic acid molecule for expression by the host of the recombinant lactoferrin receptor protein or fragment or analog thereof; or

(D) an open reading frame protein 3 (ORF3) of a *Moraxella* strain or a fragment or analog of the open reading frame protein;

and a pharmaceutically acceptable carrier therefor, said at least one active component producing an immune response when administered to a host.

30. A method for generating an immune response in a host, comprising administering to the host an immunoeffective amount of the immunogenic composition of claim 29.

31. A method of determining the presence, in a sample, of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising the steps of:

(a) contacting the sample with the nucleic acid of claim 1 or 7 to produce duplexes comprising the nucleic acid molecule and any nucleic acid molecule encoding the lactoferrin receptor protein of a strain of *Moraxella* present in the sample and specifically hybridizable therewith; and

(b) determining production of the duplexes.

32. A diagnostic kit for determining the presence, in a sample of nucleic acid encoding a lactoferrin receptor protein of a strain of *Moraxella*, comprising:

(a) the nucleic acid molecule of claim 1 or 7;

(b) means for contacting the nucleic acid molecule with the sample to produce duplexes comprising the

nucleic acid molecule and any said nucleic acid present in the sample and hybridizable with the nucleic acid molecule; and

(c) means for determining production of the duplexes.

FIG.1A

Alignment of translated 2.2kb 1bpA PCR fragments

MNQSKQNNKSKKQVLKLSALSLGLLNITQVALANTTADK Tbp1  
MSKSI~~TK~~TQTPSVHTMTTHRLNLAIKAALFGVAVLPLSVWAQENTQTDAN Lbp1

AEATDKTNLVVVLDETVVTAKKNAPVSRKANEV~~TGL~~GKVVKTAETINKEQ Tbp1  
SDAKDTKTPVVYLDAITVTAAPSA--RFDTDVTGLGKTVK~~TAD~~TLAKEQ Lbp1

VLNIRDLTRYDPGIAVVEQGRGASSGYSIRGMDKNRVAVLV~~DGIN~~QAQHY Tbp1  
VQGI~~RDL~~VRYETGVSVEQGRGGSSGFAIHGVDKNRVGITVDGIAQIQSY Lbp1

--QGPVAGKNYAAGGAIN~~EIE~~EYENVRSVEISKGANSSEYGS~~GAL~~SGSVAFVT Tbp1  
ALKDESTKRAGAGSGAMNEIEIENIAAVA~~INK~~CGNALEAGSGALGGSVAFHT Lbp1

KTADDIIKDGKDWGVQTKTAYASKNNAWNSVAAAGKAGSF~~SGL~~IIYTDR Tbp1  
KDVSDVLKSGKNLGAQSKTTYN~~SKNDHF~~SQTLAAAGKTERVEAMVOYTYR Lbp1  
QYT-R PCR4  
QYT-R PCR5

RGQ~~EY~~KAHDDAYQGSQSFDRAVATTPN~~NR~~FLIANECANGN~~YE~~ACAAGG Tbp1  
KGKENKAHSDLNGINQSLYRLGAWQQKYDLRKP~~NEL~~FAGTSYITESCLAS Lbp1  
KG-ENKAHSDLNGINQSLYRLGAWQQKYDLRKP~~NEL~~FAGTSYITESCLAS PCR4  
KG-ENKAHSDLNGINQSLYRLGAWQQKYDLRKP~~NEL~~FAGTSYITESCLAS PCR5

QTKLQAKPTNVRDKVNVKDYTGPNRLIPNPLTQDSKSL~~LLR~~PGYQLNDKH Tbp1  
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQY~~LAST~~HPHEVVS~~AK~~ Lbp1  
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQY~~LAST~~HPHEVVS~~AK~~ PCR4  
DDPKSCVQYPYVYTKARPDGIGNRNFSELSDAEKAQY~~LAST~~HPHEVVS~~AK~~ PCR5

FIG.1B

YVGGVYEITKQNYAMQDKTVPAYLAVHDIKSRLSNHAQANGYYQGNNLGERIRD	Tbp1
DYTGIRLLPDPMDYRSDSYLARLNKIKITPNLVSKLLLEDTKQTYNIRDM	Lbp1
DYTGIRLLPDPMDYRSDSYLARLNKIKITPNLVXKLLLEDTKQTYNIRDM	PCR4
DYTGTYRLLPDPMDYRSDSYLARLNKIKITPNLVSKLLLEDTKQTYNIRDM	PCR5
IGPDSGYGINYAHGVFYDEKHQKDRGLGLEVVYDSKGENKWFDDVRVSYDKQDIT	Tbp1
RHCSYHGARLNDGKPPANGGSIVLCDDYQEYLNANDASQALFRPGANDAP	Lbp1
RHCSYHGARLNDGKPPANGGSIVLCDDYQEYLNANDASQALFRPGANDAP	PCR4
RHCSYHGARLNDGKPPANGGSIVLCDDYQEYLNANDASQASFRPGANDAP	PCR5
LRSQLTNTHCSTYPHIDKNCTPDVKNKPFVKEVDNNA YKEQHNL IKAVFN	Tbp1
IPKLAYARSSVFNQEHGKTRYGLSFEFKPDTPWFKQAKLNLHQQNIQI IN	Lbp1
IPKLAYARSSVFNQEHGKTRYGLSFEFKPDTPWFKQAKLNLHQQNIQI IN	PCR4
IPKLAYARSSVFNQEHGKTRYGLGFEFKPDTPWFKQAKLNLHQQNIQI IN	PCR5
KKMALGSTHHHINLQVGYDKFNSSLRVEYRLATHQSYQKLDYTPPSNPL	Tbp1
HDIKKSCSQYPKVDLNCGI SEIGHYEQNNRYKEGRASLTGKLDNFNFDL	Lbp1
HDIKKSCSQYPKVDNSNCGI SEIGHYEQXNYRYKEGRASLTGKLDNFNFDL	PCR4
PDKFKPILGSNNKPICLDAYGYGHDHPQACNAKNSTYQNFAIKKGIEQYN	Tbp1
LGQHDLTVLAGADKVKSQFRANNPRRTIIDTTQGD AIIDESTLT AQEQAK	Lbp1
LGQHDLTVLAGTDKVKSQFRANNPRRTIIDTTQGD AIIDESTLT AQEQAK	PCR4



FIG.1C

QKTNTDKIDYQAIIDQYDKQNPNSTLKPFEKIKQSLGQEKYNKIDELGFK Tbp1  
FKQSGAAWIVKNRLGRLEEKDACGNANECERAPIHGSNQYVGINNLYTPN Lbp1  
FKQSGAAWIVKNRLGRLEEKDACGNANECERAPIHGSNQYVGINNLYTPN PCR4

AYKDLRNEWAGWTNDNSQQNANKGTDNIYQPNQATVVKDDCKYSETNSY Tbp1  
DYVDLSFGGRLDKQRIHSTDSDNII SKTYTNKSYNFGAAVHLTPDFSLLYK Lbp1  
DYVDXSFGGRLDKQRIHSTDSDNII SKTYTNKSYNFGAAVHLTPDFSLLYK PCR4  
TDSNII SKTYTNKSYNFGAAVHXTPDFSLLYK PCR5

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ADCSTTRHISGDNYFIALKDNMTINKYVDLGLGARYDRIKHKSDVPLVDNSASNQLSWNFGVV Tbp1  
TAKGFRTPSFYELYNYNSTAAQHKNPDVSVFPKRAVDVKPETSNTNEYGF Lbp1  
TAKGFRTPSFYELYNYNSTAAQHKNPDVSVFPKRAVDVKPETSNTNEYGF PCR4  
TAKGFRTPSFYELYNYNSTAAQHKNPDVSVFPKRAVDVKPETSNTNEYGF PCR5

VKP TNWLDIAYRSSQGFRMPSEMYGERFGVTIGKGTQHGCKGLYYICQQTV Tbp1  
RYQHPWGDVEMSMFKSRYKDMLDKAI PNLTKAQQEQYCKAHLDSNECVGNP Lbp1  
RYQHPWGDVEMSMFKSRYKDMLDKAI PNLTKAQQEQYCKAHLDSNECVGNP PCR4  
RYQHPWGDIEMSMFKSRYKDMLDKAI PNLTKAQQEQYCKAHLDSNECVGNP PCR5

HQTKLKPEKSFNQEIGATLHNHLGSLEVSYFKNRYTDLIVGKSEEIRTLT Tbp1  
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK Lbp1  
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK PCR4  
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTLGYGHTK PCR5

QGDNAGKQQRKGDLGFHNGQDADLTGINILGRLLDLNAANSRLPYGLYSTL Tbp1

FIG.1D

AYNKVDVKGKTLNPTLAGTNILFDAIQPSRYVVGLGYDAPSQKWGANAI	Tbp1
LGKFDYIAPKDADGWYQARPAFWDAITPARYVVGLNYDHP	Lbp1
QVWGIGTTL	PCR4
LGKFXYYIAPKDADGWYQARPAFWDAITPARYVVGLNYDHP	PCR5
QVWGIGATL	
LGKFXYYIAPKDADGWYQARPAFWDAITPARYVVGLNYDHP	
QVWGIGTTL	
HSDAKNPSELLADKNLGNNGNIQTKQATKAKSTPWQTL-DLSGYVNIKD	Tbp1
NFT	Lbp1
THSKQKDENELSALRIR-NGKRETQTLTHTI	PCR4
PKAYTL	PCR5
LDMTGYYSPTESIT	
THSKQKDENELSALRIR-NGKRETQTLTHTI	
PKAYTL	
LDMTGYYSPTESIT	
THSKQKDENELSALRIR-NGKREIQTLTHTI	
PKAYTL	
LDMTGYYSPTESIT	
LRAGVYNVFNTYYTTWEALRQTAKGAVNQHTGLSQDKHYGRYAAPGRNYQL	Tbp1
ALEMKF*	Lbp1
ARLGINNVLNTRYTTWEAARQ-----LPSEAA	PCR4
STQSTRYIAPGRSYFASLEMKF*	PCR5
ARLGINNVLNTRYTTWEAARQ-----LPSEAA	
STQSTRYIAPGRSYFASLEMKF*	
ARLGINNVLNTRYTTWEAARQ-----LPSEAA	
STQSTRYIAPGRSYFASLEMKF*	

FIG. 2A

*M. catarrhalis* 4223 1fr sequence

AAGCTTAGCATGATGGCATCGGCTGATTGT  
 10 20 30  
 CTTTGTGCCCTTGTGTGTGTGGGAGT  
 40 50 60

-35

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TGATTGTACTTACCTTAGTGGTGGATGCTT  
70 80 90

-10

GGGCTGATTTAATAAAGCGGTCCTCACAC  
100 110 120

-10-

Libo2

RBS

MET SER THR

ACACCAAACGAGATATCACCATGAGTACTG  
130 140 150

VAL LYS THR PRO HIS ILE PHE TYR GLN LYS A  
T C A A A C C C C C A C A T T T C T A C C A A A A C  
160 170 180

```

RG   THR  LEU  SER  LEU  ALA  ILE  ALA  SER  ILE
GCC ACC CTT AGC CTT GCC ATT CGC CAG TAT TT
190                                     200      210
      PHE  ALA  ALA  LEU  VAL  MET  THR  GLY  CYS  ARG  S
      TTG CCG CCT TGG TGA TGA CAG GCTG CCG CTT
                                     220      230      240

```

FIG.2B

ER ASP ASP ILE SER VAL ASN ALA PRO ASN  
 CTGATGACATCAGCGTCAATGCCCAATG  
 250 260 270  
 VAL THR GLN LEU PRO GLN GLY THR VAL SER P  
 TTACCCAACTGCCCCCAAGGCACGGTTTCAAC  
 280 290 300

RO ILE PRO ASN THR GLY HIS ASP ASN THR  
 CAATACCGAACACAGGTCTATGACACACCA  
 310 320 330  
 ASN ASN THR ASN ASN GLN GLY ASN ASN THR A  
 ATAACCAACAATCAGGGCAACAACACGG  
 340 350 360

SP ASN SER THR SER THR THR ASP PRO ASN  
 ATAACAGCACCAAGCACACCTGACCCAAATG  
 370 380 390  
 GLY ASP ASN ASN GLN LEU THR GLN ALA GLN L  
 GCGATACAAACCAACTGACACACACAA  
 400 410 420

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# FIG.2C

YS THR ALA ALA ALA ALA GLY PHE PHE VAL  
 AGACCGCGCTGCCGCAAGGGTTT TTTGTA  
 430 440 450  
 MET GLY LYS ILE ARG ASP THR SER PRO LYS A  
 TGGGTAAATTCGTGATACCAAGCCCAA A A  
 460 470 480

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SN ASP PRO ASP TYR SER ASN ASP LEU VAL  
 ATGACCGAGATTATAGCAATGATTAGTAC  
 490 500 510  
 GLN GLN TRP GLN GLY LYS LEU TYR VAL GLY I  
 AGCAGTGGCAAGGCCAATTA TATGTTGGTA  
 520 530 540

LE ASP ALA HIS ARG PRO ASP GLY ILE GLY  
 TTGATGCCCATCGCCCA GATGGCATCGGCA  
 550 560 570  
 THR GLY LYS ASN LEU ARG GLN PRO ILE THR A  
 CAGGTAAACCTTGCGTCAAGCCCATCACC  
 580 590 600

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**FIG. 2D**

LA ASN ASP ILE LYS PRO LEU TYR PHE ASN  
C C A A T G A C A T C A A C C C T T G T A T T T A A C A  
610 620 630  
LYS PHE PRO ALA LEU SER ASP LEU HIS LEU A  
A T T C C C T G C A T T G T C T G A T T T G C A T T A G  
640 650 660

SP SER GLU ARG HIS ARG PHE ASP PRO LYS  
 A C A G T G A A C G C C A C C G T T T G A C C C A A A  
 670 680 690  
 LYS LEU ASN THR ILE LYS VAL TYR GLY TYR G  
 A G C T A A A C A C C A T T A A A G T G T A T G G T A T G  
 700 710 720

[illegible]



# FIG.2E

YS LYS ASN ASN LYS PRO VAL ASP PRO TYR  
 A G A A A T A A C A G C C T G T T G A C C C T T A T 800 810  
 790  
 GLU ASN ILE ARG PHE GLY TYR LEU GLU LEU G  
 G A A A T A T C C G T T T T G G G T A T C T T G A A C T A C 820 830 840

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 LN GLY SER SER LEU THR GLN LYS ASN ALA  
 A A G G A A G C A G T C T G A C C C A A A A A T G C C G 850 860 870  
 ASP THR PRO PRO ASN ASP LYS ASP ARG ILE PRO L  
 A T A C T C C A A A T G A C A A A G A C C G C A T T C C C A 880 890 900

YS PRO MET PRO PRO ILE LEU PHE TYR HIS GLY  
 A A C C C A T G C C C A T T T G T T T A T C A C G G A G 910 920 930  
 GLU ASN ALA SER SER GLN LEU PRO SER ALA G  
 A A A C G C C A G C A G C C A G C T G C C C A G T G C T G 940 950 960

# FIG.2F

LY LYS PHE ASN TYR THR GLY ASN TRP LEU  
 GTAAATTAACTACACAGGCAAC TGGCTGT  
 970 980 990  
 TYR LEU SER ASP VAL LYS LYS ARG PRO ALA L  
 ACC TAG TGA TGT CAAA AAGCCCTGCCAC  
 1000 1010 1020  
 EU SER ALA SER ASP ARG VAL GLY VAL  
 TTTCAGCATCAGATGATCGAGTGGGGTCT  
 1030 1040 1050  
 TYR LEU ASN ALA SER GLY LYS SER ASN GLU G  
 ATCTCAATGCCAGTGGCAAAATCCAAATGAGG  
 1060 1070 1080  
 LY ASP VAL VAL SER ALA ALA HIS ILE TYR  
 GCGATGTCGTCA GTGCCGCCACATTATC  
 1090 1100 1110  
 LEU ASN GLY PHE GLN TYR LYS HIS THR PRO A  
 TAAACGGCTTTCAATA TAAAGCACACCGCTG  
 1120 1130 1140

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# FIG.2G

LA THR TYR GLN VAL ASP PHE ASP THR ASN  
 C C A C T T A T C A G G T G G A T T T T G A C A C A A C T  
 1150 1160 1170  
 SER LEU THR GLY LYS LEU SER TYR TYR ASP A  
 C A T T A C A G G C A A G C T G T C T T A T T A T G A C A  
 1180 1190 1200

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 SN PRO ASN GLN GLN THR ALA GLN GLY LYS  
 A T C C C A A C C A G C A A A C T G C C C A A G G C A A A T  
 1210 1220 1230  
 TYR ILE LYS SER GLN PHE ASP THR THR LYS L  
 A C A T C A A A G C C A A T T T G A C A C T A C C A A A A  
 1240 1250 1260

YS VAL ASN GLU THR ASP VAL TYR GLN ILE  
 A A G T C A A T G A A A C C G A T G T G T A T C A A A T T G  
 1270 1280 1290  
 ASP ALA LYS ILE ASN GLY ASN ARG PHE VAL G  
 A T G C C A A A A T C A A C G G C A A C C G C T T C G T C G  
 1300 1310 1320

## FIG.2H

LY THR ALA LYS SER LEU VAL ASN GLU ASN  
 GTACGGCCAAATCTTTGGTTAATGAGAACAA  
 1330 1340 1350  
 THR GLU THR ALA PRO PHE ILE LYS GLU LEU P  
 CAGAAACCGCACCTTTTATCAAGAGCTGT  
 1360 1370 1380  
 HE SER LYS LYS ALA ASN PRO ASN ASN PRO  
 TCTCCAAAGCCCAATCCCAATAACCCAA  
 1390 1400 1410  
 ASN PRO ASN SER ASP THR LEU GLU GLY P  
 ACCCTAATTCAGACACGCTAGAGGCGGGT  
 1420 1430 1440  
 HE TYR GLY GLU SER GLY ASP GLU LEU ALA  
 TTATGGTGAGTCGGGCGATGAGCTGGCGG  
 1450 1460 1470  
 GLY LYS PHE LEU SER ASN ASP ASN ALA SER T  
 GTAAATTTTATCCCAATGACACACGCACTTT  
 1480 1490 1500

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FIG.2I.

YR VAL VAL PHE GLY GLY LYS ARG ASP LYS  
A T G T G G T C T T T G G T G G T A A A C G A G A C A A A  
1510 1520 1530  
THR ASP LYS PRO VAL ALA THR LYS THR VAL T  
C A G A C A A A C C T G T C G C C A C A A A C G G T G T  
1540 1550 1560

YR PHE SER ALA GLY PHE GLU LYS PRO SER  
A T T T A G T G C A G G C T T T G A A A A C C T A G C A  
1570 1580 1590  
THR SER PHE VAL ASP ASN GLU THR ILE GLY A  
C C A G T T T G T G G A T A A T G A A A C G A T T G G C A  
1600 1610 1620

RG ILE ILE ASN SER LYS LYS LEU ASN ASP  
G A A T T A T A C A G C A A A A G T T A A A T G A T G  
1630 1640 1650  
ALA VAL ASN GLU LYS ILE ASP ASN GLY ASP I  
C G G T G A A T G A G A A A T T G A T A A T G G T G A T A  
1660 1670 1680

FIG.2J

LE PRO THR SER ASP GLU ARG TYR ASP GLU  
T T C C T A C C A G T G A T G A A C G C T A T G A T G A A T  
1690 1700 1710  
PHE PRO TRP GLY GLU LYS LYS ALA GLU PHE T  
T T C C T T G G G C G A A A A A A G C A G A A T T C A  
1720 1730 1740

HR LYS LYS VAL SER SER THR GLN ALA  
C C A A A A A G T C A G C A G C A C C C A A G C C G  
1750 1760 1770  
VAL PRO ALA TYR PHE GLY GLN HIS ASP LYS P  
T G C C A G C T T A T T T T G G G C A C A T G A T A A A T  
1780 1790 1800

HE TYR PHE ASN GLY ASN TYR TYR ASP LEU  
T T T A T T T A A T G G C A A C T A T T A T G A C C T A T  
1810 1820 1830  
SER ALA SER SER VAL ASP LYS LYS LEU ALA PRO A  
C A G C C A G C A G T G T T G A T A A A T T G G C C C C T G  
1840 1850 1860

FIG.2K

LA ASP ALA VAL LYS ALA ASN GLN SER ILE  
CCGATGCTGTC A A A G C C A A C C A A T C C A T T A  
1870 1880 1890  
LYS GLU LYS TYR PRO ASN ALA THR LEU ASN L  
A A G A A A A T A C C C T A A T G C C A C T A A A T A  
1900 1910 1920

YS ASP ASN GLN VAL THR ALA ILE VAL LEU  
A G G A C A C C A A G T T A C C G C C A T C G T G C T A C  
1930 1940 1950  
GLN GLU ALA LYS ASP ASN LYS PRO TYR THR A  
A A G A A G C C A A A G A T A A T A A G C C T T A T A C C G  
1960 1970 1980

LA ILE ARG ALA LYS SER TYR GLN HIS ILE  
CCATTCTGCTGCC A A A A G C T A T C A G C A C A T C A  
1990 2000 2010  
SER PHE GLY GLU THR LEU TYR ASN ASP ALA A  
G T T T G C C G A G A C G C T G T A T A C G A T G C C A A  
2020 2030 2040



# FIG.2L

SN GLN THR PRO THR ARG SER TYR PHE VAL  
 ACCAACCACACACGCAAGTTATTGTGC  
 2050 2060 2070  
 GLN GLY GLY ARG ALA ASP THR SER THR L  
 AAGCGGTAGGCGCAGATACCAAGCACCGC  
 2080 2090 2100  
 16/130  
 EU PRO LYS ALA GLY LYS PHE THR TYR ASN  
 TGCCCAAGGCAAGGTAAATTCTTACACACG  
 2110 2120 2130  
 GLY LEU TRP ALA GLY TYR LEU ILE GLN LYS L  
 GTCCTTGCGCAGGCTATCTTATCCAAATAA  
 2140 2150 2160  
 YS ASP LYS GLY TYR SER ASN ASN GLU GLU  
 AGGACAAAGGTTATAGCAATAATGAAGAAA  
 2170 2180 2190  
 THR ILE LYS LYS LYS GLY HIS GLN ASP TYR L  
 CCATCAGAAAAAGGCCCATCAGATTATC  
 2200 2210 2220

FIG.2M

EU LEU THR GLU ASP PHE THR PRO GLU ASP  
T G T T A C C G A A G A C T T C A C C C A G A G A T G  
2230  
2240  
ASP ASP ASP ASP LEU THR ALA SER ASP ASP S  
A T G A C G A T G A T T T G A C C G C A T C T G A T G A T T  
2260 2270 2280

ER GLN ASP ASP ALA HIS GLY ASP ASP  
C A C A G A T G A T G A T G C A C A T G G C G A T G A T G  
2290 2300 2310  
ASP LEU ILE ALA SER ASP ASP SER GLN ASP A  
A T T T G A T T G C A T C T G A T G A T T C A C A G A T G  
2320 2330 2340

SP ASP ALA ASP GLY ASP ASP SER ASP  
A T G A C G C A G A T G G C G A T G A C G A T T C A G A T G  
2350 2360 2370  
ASP LEU GLY ASP GLY ALA ASP ASP ALA ALA A  
A T T T G G G T G A T G G T G C A G A T G A C C C G C C G  
2380 2390 2400

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## FIG.2N

LA GLY LYS VAL TYR HIS ALA GLY ASN ILE  
 CAGGCAAGTGTAATCATGCAAGGTAAATATTC  
 2410 2420 2430  
 ARG PRO GLU PHE GLU ASN LYS TYR LEU PRO I  
 GCCCTGAATTGAAACAAATACTTGCCCA  
 2440 2450 2460

LE ASN GLU PRO THR HIS GLU LYS THR PHE  
 TTAATGAGCCTACTCATGAATAACCTTTTG  
 2470 2480 2490  
 ALA LEU ASP GLY LYS ASN LYS ALA LYS PHE A  
 CCTAGATGGTAATAATAAGCTAAGTTTG  
 2500 2510 2520

SP VAL ASP PHE ASP THR ASN SER LEU THR  
 ATGTGGATTTTGACACCAACAGCCTAACTG  
 2530 2540 2550  
 GLY LYS LEU ASN ASP GLU ARG GLY ASP ILE V  
 GTAAATTAAACGATGAGAGAGGTGATATCG  
 2560 2570 2580

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FIG.2O

AL PHE ASP ILE LYS ASN GLY LYS ILE ASP  
T C T T G A T A T C A A A A T G G C A A A T T G A T G  
2590 2600 2610  
GLY THR GLY PHE THR ALA LYS ALA ASP VAL P  
G C A C A G G C T T T A C C G C C A A A G C C G A T G T G C  
2620 2630 2640

RO ASN TYR ARG GLU GLU VAL GLY ASN ASN  
C A A C T A T C G T G A A G A A G T G G G T A A C A C C  
2650 2660 2670  
GLN GLY GLY GLY PHE LEU TYR ASN ILE LYS A  
A A G G T G G C G G T T C T T A T A C A C A T C A A A G  
2680 2690 2700

SP ILE ASP VAL LYS GLY GLN PHE PHE GLY  
A T A T G A T G T C A A G G G C A A T T T T T G G C A  
2710 2720 2730  
THR ASN GLY GLU LEU ALA GLY GLN LEU G  
C A A A T G G C G A A G A G T T G G C A G G C A G T T A C  
2740 2750 2760

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# FIG.2P

LN TYR ASP LYS GLY ASP GLY ILE ASN ASP  
 AGTACGACAAAGGCGATGGCATCAATGACCA  
 2770 2780 2790  
 THR ALA GLU LYS ALA GLY ALA VAL PHE GLY A  
 CCGCCGAAAGGCGCTGCTTTGGGG  
 2800 2810 2820  
 LA VAL LYS ASP LYS \*\*\*  
 CTGTTAAAGATAATAAGCCCCCTTCATC  
 2830 2840 2850  
 ATCGTTTAGTCGCTTGACCGACAGTTGATG  
 2860 2870 2880  
 ACGCCCTTGGCAATGTCTTAAACAGCACT  
 2890 2900 2910  
 TTGAACAGTGCCCTTGGGCGAATTCTTGA  
 2920 2930 2940  
 TAAATGCCACGATTTGCCCTTGGGCTAATA  
 2950 2960 2970  
 -35  
 TCTTGATATAAACATCGCCATAAATAGAAA  
 2980 2990 3000  
 -10

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FIG. 20

2nd possible start

[illegible]

AL LEU PRO LEU SER VAL TRP ALA GLN GLU  
 T T T A C C C T A T C C G T C T G G C G C A A G A G A  
 3130 3140 3150  
 ASN THR GLN THR ASP ALA ASN SER ASP ALA L  
 A C A C T C A G A C A G A T G C C A A C T C T G A T G C C A  
 3160 3170 3180

# FIG.2R

YS ASP THR LYS THR PRO VAL VAL TYR LEU  
 A G A C A C A A A A C C C T G T C T A T T A G 3190  
 3200  
 ASP ALA ILE THR VAL THR ALA ALA PRO SER A  
 A T G C C A T C A C G G T A A C C G C C C C A T C T G 3220  
 3230 3240  
  
 LA PRO VAL SER ARG PHE ASP THR ASP VAL  
 C C C T G T T C T C G G T T T G A C A C C G A T G T A A 3250  
 3260 3270  
 THR GLY LEU GLY LYS THR VAL LYS THR ALA A  
 C A G G C C T T G G C A A A C G G T C A A A C C G C T G 3280  
 3290 3300  
  
 SP THR LEU ALA LYS GLU GLN VAL GLN GLY  
 A C A C G C T G G C A A A G A A C A A G T G C A G G C C 3310  
 3320 3330  
 ILE ARG ASP LEU VAL ARG THR GLU THR GLY V  
 A T T C G T G A T T T G G T G C G T T A T G A A C T G G G G 3340  
 3350 3360

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## FIG.2S

AL SER VAL VAL GLU GLN GLY ARG GLY GLY  
T G A G T G G T T G A G C A G G G C G T G G T G G C A  
3370 3380 3390  
SER SER GLY PHE ALA ILE HIS GLY VAL ASP L  
G C A G C G G A T T T G C C A T T C A T G G C G T G G A T A  
3400 3410 3420

YS ASN ARG VAL GLY ILE THR VAL ASP GLY  
A A A C C G A G T G G G C A T T A C C G T A G A T G G C A  
3430 3440 3450  
ILE ALA GLN ILE GLN SER TYR LYS ASP GLU S  
T T G C C C A A A T T C A A T C C T A C A A G A T G A A T  
3460 3470 3480

ER THR LYS ARG ALA GLY ALA GLY SER GLY  
C C A C C A A A C G A G C T G G T G C A G G C T C T G G G G  
3490 3500 3510  
ALA MET ASN GLU ILE GLU ILE GLU ASN ILE A  
C G A T G A A T G A G A T A G A G A T T G A A A C A T T G  
3520 3530 3540

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## FIG.2T

LA ALA VAL ALA ILE ASN LYS GLY GLY ASN  
 CCGCCGTTGCCCATCAATAAGGTGGTAATG  
 3550 3560 3570  
 ALA LEU GLU ALA GLY SER GLY ALA LEU GLY G  
 CCTAGAGCAGGCTCTGGTGCGCG  
 3580 3590 3600  
 LY SER VAL ALA PHE HIS THR LYS ASP VAL  
 GTCCGTTGGCGTTTCAATACCAAGATGTA  
 3610 3620 3630  
 SER ASP VAL LEU LYS SER GLY LYS ASN LEU G  
 CGGATGTCCTTAAATCTGGTAATACTTG  
 3640 3650 3660  
 LY ALA GLN SER LYS THR THR TYR ASN SER  
 GCGCTCAAGCAAAACCACTTATAACAGCA  
 3670 3680 3690  
 LYS ASN ASP HIS PHE SER GLN THR LEU ALA A  
 AATGACCATTTTAGTCAGACCTGGCAG  
 3700 3710 3720

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# FIG.2U

LA ALA GLY LYS THR GLU ARG VAL GLU ALA  
 CGG CAGG TAA AACC GAG CGTG TGG AAG CGA  
 3730 3740 3750  
 MET VAL GLN TYR THR TYR ARG LYS GLY LYS G  
 TGG TGC AATA TACCT ACCGT AAGG CAAAG  
 3760 3770 3780

LU ASN LYS ALA HIS SER ASP LEU ASN GLY  
 AAA CAAAG CACAC GAG CCAATA TGGA  
 3790 3800 3810  
 ILE ASN GLN SER LEU TYR ARG LEU GLY ALA T  
 TCA ACCA AAGCC TATA TCGCT TGGGT GCA T  
 3820 3830 3840

RP GLN GLN LYS TYR ASP LEU ARG LYS PRO  
 GGCA CAAATA TGA TTAA GAAAG CCA  
 3850 3860 3870  
 ASN GLU LEU PHE ALA GLY THR SER TYR ILE T  
 ATGA ACTGT TTG CAGGC ACAAGCT ATCA  
 3880 3890 3900

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# FIG.2V

HR GLU SER CYS LEU ALA SER ASP ASP PRO  
 CCGAAGCTGTTTGGAAGTGATGACCCAA  
 3910 3920 3930  
 LYS SER CYS VAL GLN TYR PRO TYR VAL TYR T  
 AAGCTGCGTACAAATACCTTATGCTACA  
 3940 3950 3960

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 HR LYS ALA ARG PRO ASP GLY ILE GLY ASN  
 CCAAGCCGACCAAGATGGCAATCGGCAATC  
 3970 3980 3990  
 ARG ASN PHE SER GLU LEU SER ASP ALA GLU L  
 GCAATTTTCTGAGTTAAGCGATGCTGAAA  
 4000 4010 4020

YS ALA GLN TYR LEU ALA SER THR HIS PRO  
 AAGCAATAATTGGAATCCACGCCCC  
 4030 4040 4050  
 HIS GLU VAL VAL SER ALA LYS ASP TYR THR G  
 ATGAGGTGTCTCTGCCAAGATATACAG  
 4060 4070 4080

# FIG.2W

LY ILE TYR ARG ILE ILE LEU PRO ASP PRO MET  
 G C A T T A T C G G T T G T T A C C T G A C C C A T G G  
 4090 4100 4110  
 ASP TYR ARG SER ASP SER TYR LEU ALA ARG L  
 A C T A T C G T T C A G A C T C G T A T T G G C A C G C C  
 4120 4130 4140

EU ASN ILE LYS ILE THR PRO ASN LEU VAL  
 T T A A C A T C A A A T C A C C C A A A T C T G G T C A  
 4150 4160 4170  
 SER LYS LEU LEU LEU GLU ASP THR LYS GLN T  
 G T A A A C T G T T A T T A G A A G A C C A A G C A A A  
 4180 4190 4200

HR TYR ASN ILE ARG ASP MET ARG HIS CYS  
 C A T A C A C A T T C G T G A T A T G C G T C A T T G T A  
 4210 4220 4230  
 SER TYR HIS GLY ALA ARG LEU GLY ASN ASP G  
 G T T A C C A T G G G G C A A G A T T G G C A A T G A T G  
 4240 4250 4260

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# FIG.2X

LY LYS PRO ALA ASN GLY GLY SER ILE VAL  
 GTAGCCCTGCCCAATGGTGCGCTCCATTGTTCT  
 4270 4280 4290  
 LEU CYS ASP ASP TYR GLN GLU TYR LEU ASN A  
 TTGGCGATGATTATCAAGAGTATCTAACG  
 4300 4310 4320  
 LA ASN ASP ALA SER GLN ALA LEU PHE ARG  
 CCATGACGCATCAACAGCATTTATTAGAC  
 4330 4340 4350  
 PRO GLY ALA ASN ASP ALA PRO ILE PRO LYS L  
 CAGGTGCTAATGATGCCCATTCCTCAAC  
 4360 4370 4380  
 EU ALA TYR ALA ARG SER SER VAL PHE ASN  
 TGGCTTATGCCCAGAGCAGTGTTTAAACC  
 4390 4400 4410  
 GLN GLU HIS GLY LYS THR ARG TYR GLY LEU S  
 AAGAGCATGGCAAACTCGCTATGGGTTAA  
 4420 4430 4440

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## 29/130

```

IN  ASN  ILE  GLN  ILE  ILE  ASN  HIS  ASP  ILE
A A A T A T C C A A A T C A T T A A C C A T G A C A T T A
4510
LYS  LYS  SER  CYS  SER  GLN  TYR  PRO  LYS  VAL  A
A A A A T C G T G C A G C C A A T A T C C T A A G G T G G
4540
4550
4560

```

SP LEU ASN CYS GLY ILE SER GLU ILE GLY  
 A T T A A A T G T G G C A T C A G T G A A A T T G G G C  
 4570 4580 4590  
 HIS TYR GLU TYR GLN ASN ASN TYR ARG TYR L  
 A T T A T G A A T A T C A A A T A C C G T T A T A  
 4600 4610 4620



## FIG.2Z

YS GLU GLY ARG ALA SER LEU THR GLY LYS  
 A A G A A G G C C G T G C C A G C T T G A C A G G C A A A C  
 4630 4640 4650  
 LEU ASP PHE ASN PHE ASP LEU LEU GLY GLN H  
 T T G A T T T A A T T T G A C C T G C T G G G T C A G C  
 4660 4670 4680

IS ASP LEU THR VAL LEU ALA GLY ALA ASP  
 A C G A T T G A C G G T G T T G G C T G G T G C A G A T A  
 4690 4700 4710  
 LYS VAL LYS SER GLN PHE ARG ALA ASN ASN P  
 A A G T T A A A G C C A A T T C G T G C C A A C A A C C  
 4720 4730 4740

RO ARG ARG THR ILE ILE ASP THR THR GLN  
 C C A G A C G C A C A A T C A T T G A C A C C A C C A A G  
 4750 4760 4770  
 GLY ASP ALA ILE ILE ASP GLU SER THR LEU T  
 G C G A T G C C A T C A T T G A T G A A A G C A C G C T G A  
 4780 4790 4800

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LE HIS GLY SER ASN GLN TYR VAL GLY ILE  
T T C A T G G C A G T A A C C A A T A T G T G G C A T T A  
4930 4940 4950  
ASN ASN LEU TYR THR PRO ASN ASP TYR VAL A  
A C A A C C T T A T A C A C C A A A T G A T A T G T G G  
4960 4970 4980

FIG.2B'

SP LEU SER PHE GLY GLY ARG LEU ASP LYS  
A T T A A G T T T G G T G G A C G C T T G G A T A A A C  
4990 5000 5010  
G L N A R G I L E H I S S E R T H R A S P S E R A S N I L E I  
A A C G C A T T C A C A G C A C C G A T T C A A A C A T C A  
5020 5030 5040  
LE SER LYS THR TYR TYR THR ASN LYS SER TYR  
T C A G C A A A C T T A C A C C A A A A G C T A T A  
5050 5060 5070  
A S N P H E G L Y A L A A L A V A L H I S L E U T H R P R O A  
A T T T G G A G C G C G G T T C A T C T G A C A C C T G  
5080 5090 5100  
SP PHE SER LEU LEU TYR LYS THR ALA LYS  
A T T T A G C C T G T T G T A T A A A C T G C C A A A G  
5110 5120 5130  
G L Y P H E A R G T H R P R O S E R P H E T Y R G L U L E U T  
G C T T T C G T A C G C C A A G T T T T A T G A A C T G T  
5140 5150 5160

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FIG.2C'

YR ASN TYR ASN SER THR ALA ALA GLN HIS  
 A C A C T A T A C A G C A C C G C C C A G C A T A  
 5170 5180 5190  
 LYS ASN ASP PRO ASP VAL SER PHE PRO LYS A  
 A A A T G A C C C T G A T G T G T C T T T C C C A A A C  
 5200 5210 5220

RG ALA VAL ASP VAL LYS PRO GLU THR SER  
 G A G C G T T G A T G T C A A C C T G A A C T T C C A  
 5230 5240 5250  
 ASN THR ASN GLU TYR GLY PHE ARG TYR GLN H  
 A T A C C A A T G A A T A C G G C T T T C G C T A T C A G C  
 5260 5270 5280

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IS PRO TRP GLY ASP VAL GLU MET SER MET  
 A C C C T T G G G G A T G T T G A G A T G A G C A T G T  
 5290 5300 5310  
 PHE LYS SER ARG TYR LYS ASP MET LEU ASP L  
 T C A A A G C C G T T A C A G A G A C A T G T T A G A T A  
 5320 5330 5340

FIG.2D'

YS ALA ILE PRO ASN LEU THR LYS ALA GLN  
 AAGCCATACCGAACCTAACCAAGCCCAAC  
 5350 5360 5370  
 GLN GLU TYR CYS LYS ALA HIS LEU ASP SER A  
 AAGATATTGTAGGCTCATTTGGATTCCA  
 5380 5390 5400  
 SN GLU CYS VAL GLY ASN PRO PRO THR PRO  
 ATGATTGTTGGCAATCCGCCCAAGCCCA  
 5410 5420 5430  
 LYS THR SER ASP GLU VAL PHE ALA ASN LEU T  
 AACCAGTGATGAGGTATTTGCCAACAATTAT  
 5440 5450 5460  
 YR ASN ALA THR ILE LYS GLY VAL SER VAL  
 AATGCCACCATCAAGGGGTGAGTGCA  
 5470 5480 5490  
 LYS GLY LYS LEU ASP LEU HIS ALA MET THR S  
 AAGCAAACTGGATTGCA TGCCATGACAT  
 5500 5510 5520

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## FIG.2E'

ER LYS LEU PRO ASP GLY LEU GLU MET THR  
 C A A A C T G C C A G A T G G T C T T G A A A T G A C C T  
 5530 5540 5550  
 LEU GLY TYR GLY HIS THR LYS LEU GLY LYS P  
 T G G G T T A T G G T C A T A C C A A A T T G G G A A A T  
 5560 5570 5580  
 35/130  
 HE ASP TYR ILE ALA PRO LYS ASP ALA ASP  
 T T G A T T A C A T T G C A C C C A A A G A T G C C G A T G  
 5590 5600 5610  
 GLY TRP TYR GLN ALA ARG PRO ALA PHE TRP A  
 G T T G G T A T C A G G C T C G C C C T G C T T T T G G G  
 5620 5630 5640  
 SP ALA ILE THR PRO ALA ARG TYR VAL VAL  
 A T G C C A T C A C C C A G C G C G C T A T G T G G T C G  
 5650 5660 5670  
 GLY LEU ASN TYR ASP HIS PRO SER GLN VAL T  
 G T C T A A A C T A T G A C C A C C C A G T C A A G T A T  
 5680 5690 5700

## FIG.2F

RP GLY ILE GLY THR THR LEU THR HIS SER  
 GGGCATTGGCCACACTTTAACGCACAGCA  
 5710 5720 5730  
 LYS GLN LYS ASP GLU ASN GLU LEU SER ALA L  
 AACAAAGATGAATAATGAGCTAAGTGCCC  
 5740 5750 5760  
 EU ARG ILE ARG ASN GLY LYS ARG GLU THR  
 TTAGAAATCCGAAATGGCCAAAGAGAAACAC  
 5770 5780 5790  
 GLN THR LEU THR HIS THR ILE PRO LYS ALA T  
 AACCTTAACGCACACAAATACCCAAAGCCT  
 5800 5810 5820  
 YR THR LEU LEU ASP MET THR GLY TYR TYR  
 ATACCTTACTGGACATGACAGGCTATTATA  
 5830 5840 5850  
 SER PRO THR GLU SER ILE THR ALA ARG LEU G  
 GCCCAACTGAGAGCATCACCGCTCGTCTTG  
 5860 5870 5880

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# FIG.2G'

LY ILE ASN ASN VAL LEU ASN THR ARG TYR  
 GTATCAACAATGTATTAAACACCGCTACA  
 5890 5900 5910  
 THR THR TRP GLU ALA ALA ARG GLN LEU PRO S  
 CCACATGGGAAGCGGCAACGCGCACTGCCCA  
 5920 5930 5940  
 ER GLU ALA ALA SER SER THR GLN SER THR  
 GGAGCGCTGCAAGCAGTACCCAAATCAACCC  
 5950 5960 5970  
 ARG TYR ILE ALA PRO GLY ARG SER TYR PHE A  
 GTACATTTGCCACCAAGGTCCGCACTTCTTG  
 5980 5990 6000  
 ORF3  
 LA SER LEU GLU MET LYS PHE \*\*\* MET THR  
 CCAGTCTTGAAATGAGTTTTAATATGACC  
 6010 6020 6030  
 CYS LEU PRO LYS THR ASN PRO ALA LEU . LYS  
 TGTTTACCAAGACCAACCTGCTTTAA  
 6040 6050 6060

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## FIG.2H'

VAL LYS HIS ARG PHE LEU LYS GLN VAL LEU  
 G T C A A G C A C A G A T T T T A A A G C A G G T G C T G  
 6070 6080 6090  
 LEU LEU LEU CYS VAL ASP THR LEU THR ALA  
 T T A T T G C C T T T G T G T T G A T A C A T T A C A G C A  
 6100 6110 6120  
 GLN ALA TYR ALA HIS SER HIS HIS THR PRO  
 C A G G C G T A C G C C C A C A G C C A T C A T A C G C C C  
 6130 6140 6150  
 ILE HIS THR PRO THR HIS GLU LEU PRO SER  
 A T T C A T A C A C C C A C G C A T G A G C T G C C A T C T  
 6160 6170 6180  
 ALA ASP ALA LEU SER ASP GLU GLY LEU GLY  
 G C T G A T G C T T T A T C A G A T G A A G G C T T G G G T  
 6190 6200 6210  
 LYS ASP LEU GLY SER LEU ASP SER LEU ASP  
 A A G G A T T T G G G C A G T T T G G A C A G T T T G G A T  
 6220 6230 6240

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## FIG.2I'

SER PRO ASP GLY LEU GLY ASP GLY LEU GLY  
 A G C C A G A T G G T T T G G T G A T G G T T A G G C  
 6250 6260 6270  
 ASP GLY LEU GLY ASP GLY LEU LYS SER ASP  
 G A T G G T T T G G G T G A T G G C T T A A A A G T G A T  
 6280 6290 6300

LYS ALA PRO LEU PRO ILE ASN ALA LEU THR  
 A A G C C C T T T A C C C A T C A A C G C C T T G A C C  
 6310 6320 6330  
 ALA HIS GLN THR ASN GLU SER GLN PRO ALA  
 G C C C A T C A G A C C A A T G A G A G C C A G C C T G C C  
 6340 6350 6360

PRO PRO SER VAL ASP VAL ASN PHE LEU LEU  
 C C A C C G A G C G T A G A T G T C A A T T T T A C T T  
 6370 6380 6390  
 ALA GLN PRO GLU ALA PHE TYR HIS VAL PHE  
 G C C C A G C C A G A G G C A T T T A T C A T G T C T T T  
 6400 6410 6420

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FIG.2J'

HIS GLN ALA ILE VAL GLN ASP ASP VAL ALA  
 C A T C A A G C G A T T G T G C A A G A T G A T G T G C A  
 6430 6440 6450  
 THR LEU ARG LEU LEU LEU LEU PRO PHE TYR ASP  
 A C A T T A C G C T T G T T A T T G C C A T T T A T G A C  
 6460 6470 6480

ARG LEU PRO ASP ASP TYR GLN ASP ASP VAL  
 C G C C T G C C T G A T T A T C A A G A T G A T G T T  
 6490 6500 6510  
 LEU LEU LEU PHE ALA GLN SER LYS LEU ALA  
 T T G T T G T T A T T T G C C C A A A G T A A C T T G C C  
 6520 6530 6540

LEU SER ASP GLY ASN THR LYS LEU ALA LEU  
 C T A A G T G A T G G C A A T A C C A A A T T G G C A T T G  
 6550 6560 6570  
 ASN LEU LEU THR ASP LEU SER SER ASN LYS GLU  
 A A T C T G C T G A C C G A T T T G A G T A C A A G A G  
 6580 6590 6600

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FIG.2K'

PRO THR LEU THR ALA VAL LYS LEU GLN LEU  
 C C A C A C T T A C G C G G T A A A T T A C A C T T  
 6610 6620 6630  
 ALA SER LEU LEU LEU THR ASN LYS HIS ASP  
 G C T T C C T T G T T G C T G A C C A A C A G C A C G A T  
 6640 6650 6660

LYS HIS ALA GLN MET VAL LEU ASP GLU LEU  
 A A C A C G C C C A A A T G G T G C T A G A T G A C T C  
 6670 6680 6690  
 LYS ASP ASP ALA HIS PHE LEU LYS LEU SER  
 A A A G A T G A T G C C C A C T T T T A A A T T A A G C  
 6700 6710 6720

LYS LYS GLU GLN ARG TRP VAL LEU SER GLN  
 A A A A G A G C A A A G A T G G G T G C T A T C G C A A  
 6730 6740 6750  
 SER ARG TYR LEU HIS LYS LYS TYR LYS MET  
 A G T C G C T A T T T A C A T A A A A A T A A A T G  
 6760 6770 6780

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## FIG.2L'

GLY LEU ASP LEU GLY ILE ASN TYR LEU HIS  
 GGC TTG GAT TTG GGC ATC AAC TAT CTGCAT  
 6790 6800 6810  
 LEU ASP ASN ILE ASN ALA ALA SER THR ILE  
 TTG GAT AAT ATCAACGCCCTCCACCATC  
 6820 6830 6840

THR GLN PRO ASN ILE LYS LYS ASP ALA PRO  
 ACCAGCCCAATATTAATAAGATGCCCA  
 6850 6860 6870  
 LYS PRO ALA HIS GLY LEU ALA LEU SER LEU  
 AACCTGCTCATGGGCTTGCCCTTATCGCTT  
 6880 6890 6900

GLY VAL ASN LYS TYR THR PRO LEU SER HIS  
 GGTGTGAATAATAACACGCCCTTAGTCAT  
 6910 6920 6930  
 GLY MET SER ILE TYR THR ALA LEU ASP VAL  
 GGCA TGAGTATTATACAGCCCTAGATGTT  
 6940 6950 6960

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[illegible]

ALA THR ASN GLN GLN HIS GLY ARG LEU SER  
 GCC ACC AATCAGCAGCATGGCAGATTATCC  
 7090 7100 7110  
 PRO ARG LYS ASP SER GLN GLY VAL ALA PHE  
 CCCAGAAAGACAGTCAGGGCGTGCGCTTT  
 7120 7130 7140



# FIG.2N'

GLY SER HIS HIS ARG ILE ASN ASP LYS TRP  
 GGCAGCCATCATCGGATCAATGATAATGG  
 7150 7160 7170  
 GLN ASN ALA PHE PHE ALA ARG MET GLU LYS  
 CAAATGCCGTTT TTTGCCACGCATGGA A A A  
 7180 7190 7200  
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 GLY ASN TYR THR GLU ARG TYR GLN GLY TYR  
 GGCAATATACCGAGCGTTATCAAGGTTAT  
 7210 7220 7230  
 ASP GLY LYS ARG TYR HIS VAL ASN ASP THR  
 GATGGCAAGCGTTATCATGTGAATGACACC  
 7240 7250 7260  
 ILE LEU LEU GLN ASP GLY PRO ASN ARG ARG  
 ATTTGTTGCAAGATGGCCCAATCGTCGT  
 7270 7280 7290  
 TYR SER LEU GLY VAL GLY TYR GLN LEU SER  
 TACTCTTTGGGCGTGGGTTATCAGCTTAGC  
 7300 7310 7320

FIG.20'

HIS LEU GLN ASP ALA THR LYS SER SER HIS  
 C A T C T G C C A A G A T G C A A C A A A G C A G T C A T  
 7330 7340 7350  
 ALA THR LYS ILE HIS PHE GLY VAL LEU GLN  
 G C C A C A A G A T A C A T T T T G G G G T G T T G C A A  
 7360 7370 7380  
 ARG LEU PRO ASN GLY LEU THR VAL GLN GLY  
 A G A T T G C C A A A T G G T C T G A C C G T G C A A G G T  
 7390 7400 7410  
 ARG VAL SER ALA GLU ARG GLU ARG TYR HIS  
 A G A G T G A G T G C T G A G C G T G A G C G T T A T C A T  
 7420 7430 7440  
 GLY LYS LEU LEU ARG LEU VAL ASN PRO ASP  
 G G T A A A T T A T T G C G T C T G G T T A A T C C T G A T  
 7450 7460 7470  
 ASP VAL TYR ARG THR ASP LYS THR LEU THR  
 G A T G T G T A T C G C A C A G A T A A A C C C T A A C C  
 7480 7490 7500

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## FIG.2P'

LEU GLN THR SER ILE TRP HIS LYS ASP ILE  
 C T A C A A C C T C C A T T T G G C A C A A G A C A T T 7520 7530  
 7510  
 HIS TRP LEU GLY LEU THR PRO LYS LEU THR  
 C A C T G G C T T G G A T T A C G C C A A A G C T G A C T 7540 7550 7560

TYR ARG TYR SER LYS ASN SER ASN LEU  
 T A T C G T T A C A G T A A A A T A A C A G T A A C T T A 7570 7580 7590  
 46/130  
 PRO ALA LEU TYR SER HIS ASN LYS GLN ASN  
 C C A G C A C T T T A T A G C C A T A A C A A A A A T 7600 7610 7620

PHE TYR LEU GLU LEU GLY ARG SER PHE \*\*\*  
 T T T A T T T G G A G C T T G G T C G G T C G T T T A A 7630 7640 7650

Restriction map of clone pLD1-8, *M. catarrhalis* strain 4223 *lfr*

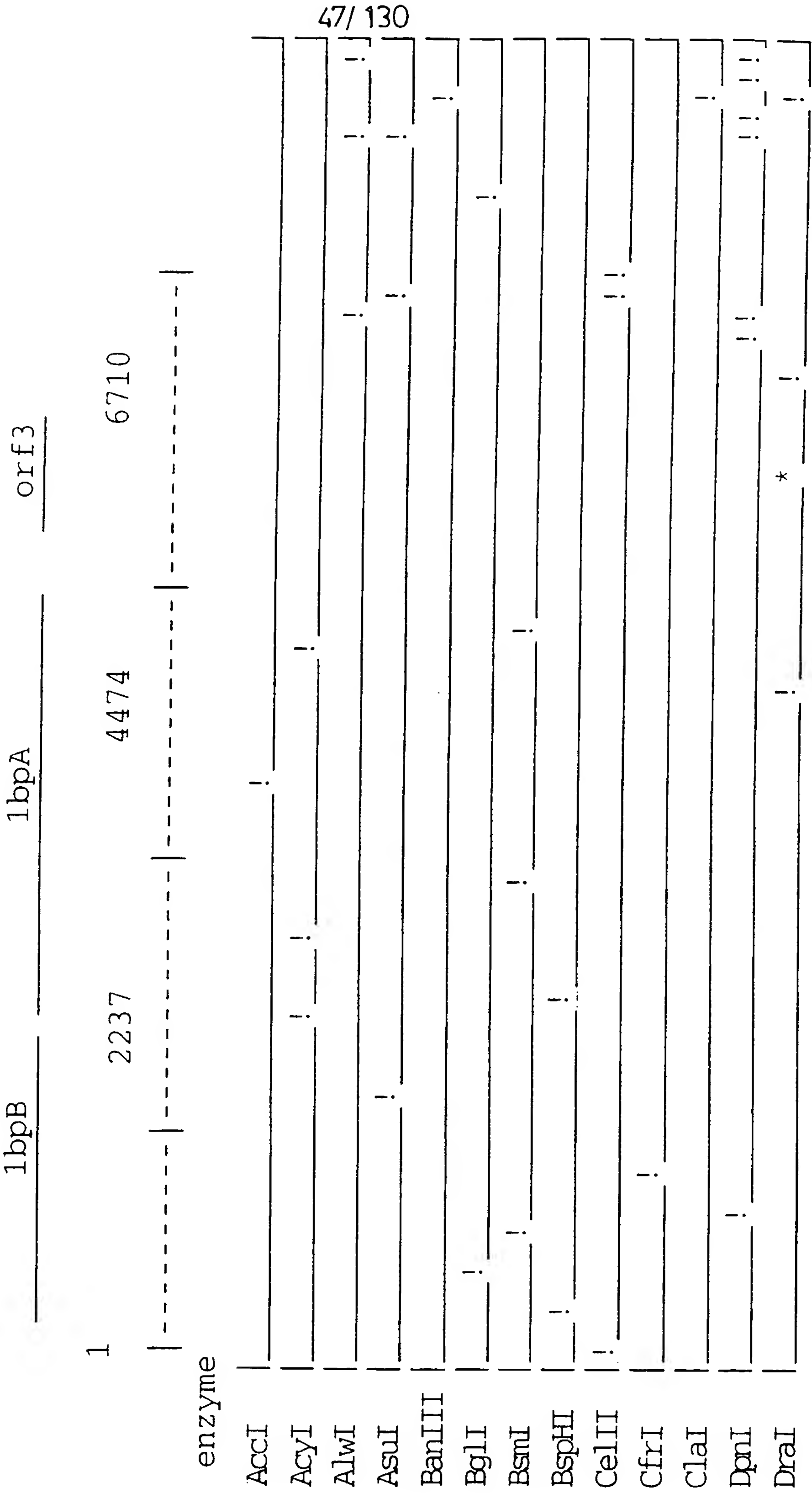


FIG.3A



FIG.4A *M. catarrhalis* Q8 lfr sequence

AAGCTTAGCATGATGGCATCGGCTGATTGT 20 30  
CTTTTGCCCTTGTGTGTGTGTGGGAGT 40 50 60

TGATTGTACTTACCCTTAGTGGTGATGCTT 70 80 90  
GGGCTGATTATAATAATTAATCAAGCG 100 110 120

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GTC TTC A C A C A C A C C C A A A C G A G A T C A C 130 140 150  
RBS  
lbp2  
MET SER THR VAL LYS THR PRO HIS ILE PHE  
C A T G A G T A C T G T C A A A C C C C C A T A T T T 160 170 180

TYR GLN LYS ARG THR LEU SER LEU ALA ILE  
C T A C C A A A A C G C A C C C T T A G C C T T G C C A T 190 200 210  
ALA SER ILE PHE ALA ALA LEU VAL MET THR  
C G C C A G T A T T T T G C C T G C C T T G G T G A T G A C 220 230 240

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[illegible][illegible][illegible]



## FIG.4C

GLN ALA GLN LYS THR ALA ALA ALA GLY  
 A C A G C G C A A A A A C T G C C G C C G C A G G 430  
 440  
 PHE PHE VAL MET GLY LYS ILE ARG ASP THR  
 G T T T T T G T G A T G G G T A A A A T T C G T G A T A C 460  
 470 480

SER GLU LYS ASN ASP PRO ASP TYR SER ASP  
 C A G C G A A A A A A T G A C C C A G A T T A T A G T G A T 490  
 500 510  
 ASP LEU LYS LYS GLN GLN TRP LEU GLY LYS LEU  
 T G A T T T A A A A C A G C A G T G G C T G G C C A A A T T 520  
 530 540

TYR VAL GLY ILE ASP ALA HIS ARG PRO ASP  
 A T A T G T T G G T A T T G A T G C C C A T C G C C C A G A 550  
 560 570  
 GLY ILE GLY LYS LYS GLY LYS ASN LEU ARG GLN  
 T G G C A T C G G A A A A G G T A A A A C T T G C G T C A 580  
 590 600

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FIG.4D

PRO ILE THR ALA ASN ASP ILE LYS PRO LEU  
GCCCATCACCGCCAAATGACATCAACCCCTT 610  
620  
TYR PHE ASN LYS PHE PRO ALA LEU SER ASP  
GTATTTTAAACAATAATCCCTGCA TTGCTGA 640  
650  
LEU HIS LEU ASP SER GLU ARG HIS ARG PHE  
TTTGCACTTAGACAGTGACGCCATCGTTT 670  
680  
ASP PRO GLN LYS ILE ASN THR ILE LYS VAL  
TGACCCCAAGATATAACCACTTAAGT 700  
710  
TYR GLY TYR GLY ASN LEU THR THR PRO SER  
GTATGGTTATGGTAACTTAACAACCAATC 730  
740  
ASN ASN ASN THR HIS ILE ASN HIS GLN GLN  
CAACAACAACCTCACATCAATCAGCA 760  
770  
780

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## FIG.4E

ALA ASP ASN LYS LYS LYS ASN ASN LYS PRO VAL  
 AGCTGATAATAAGAAATAACAGCCTGT  
 790 800 810  
 ASP PRO TYR GLU ASN ILE ARG PHE GLY TYR  
 TGACCCCTTATGAATAATATCCGTTTGGGTA  
 820 830 840  
 LEU GLU LEU GLN GLY SER SER LEU THR GLN  
 TCTTGAACTACAGGAAGCAGCCTGACCA  
 850 860 870  
 LYS ASN ALA ASP ASN GLN GLN GLU GLN ASP  
 AAAATGCCGATATAATAAGCAGCAAGA  
 880 890 900  
 ARG ILE PRO LYS PRO MET PRO ILE LEU PHE  
 CCGCATTCCTCAACCCATGCCCATTTGTT  
 910 920 930  
 TYR HIS GLY GLU ASN ALA SER SER GLN LEU  
 TTATCATGGAGAAACGCCAGCAGCCT  
 940 950 960

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## FIG. 4F

[illegible][illegible][illegible]

FIG.4G

HIS THR PRO ALA THR TYR GLN VAL ASP PHE  
 GCACAGCCCTGCCACTTATCAGGTGATT  
 1150 1160 1170  
 ASP THR ASN SER LEU THR GLY LYS LEU SER  
 TGACACAACCTCATTAACAGGCCAAGCTGTC  
 1180 1190 1200

TYR TYR ASP ASN PRO ASN GLN GLN ASN ASN  
 CTATTATGACAAATCCCAATCAGCAATAA  
 1210 1220 1230  
 LYS GLY GLU TYR LEU LYS SER GLN PHE ASP  
 TAAAGCGGAATATCTCAAAAGCCCAATTGA  
 1240 1250 1260

THR THR LYS LYS VAL ASN GLU THR ASP VAL  
 CACTACCAAAAGTCAATGAACCGATGT  
 1270 1280 1290  
 TYR GLN ILE ASP ALA LYS ILE ASN GLY ASN  
 GTATCAAAATTGATGCCAATAATCAACGGTAA  
 1300 1310 1320

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FIG.4H

ARG PHE VAL VAL GLY THR ALA LYS SER LEU VAL  
CCGCTTGTCTCGGTACGGCCAAATCTTTGGT  
1330 1340 1350  
ASN GLU LYS THR GLN THR ALA PRO PHE ILE  
TAATGAGAAACAACAACCGCACTTTAT  
1360 1370 1380  
LYS GLU LEU PHE SER LYS LYS ALA ASN PRO  
CAAGAGCTGTCTCTCCAATAAGCCAAACC  
1390 1400 1410  
ASN ASN PRO ASN PRO ASN SER ASP THR LEU  
CAATAACCCAAACCTAATTCAGACACGCT  
1420 1430 1440  
GLU GLY GLY PHE TYR GLY GLU SER GLY ASP  
AGAGCGGATTTTATGGTGAGTCGGCGGA  
1450 1460 1470  
GLU LEU ALA GLY LYS PHE LEU SER ASN ASP  
TGAGCTGGCGGTAATAATTTTATCCATA  
1480 1490 1500

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FIG. 4I

ASN ALA SER TYR VAL VAL PHE GLY GLY LYS  
 C A C G C A T C T T A T G T G T C T T T G G T G G C A A  
 1510 1520 1530  
 ARG ASP LYS LYS THR THR LYS PRO VAL ALA THR  
 A C G A G A C A A A C G A C T A A A C C T G T C G C C A C  
 1540 1550 1560

LYS THR VAL TYR PHE SER ALA GLY PHE GLU  
 A A A A C G G T G T A T T T A G T G C A G G C T T T G A  
 1570 1580 1590  
 LYS PRO SER THR SER PHE VAL ASP ASN GLU  
 A A A C C C A G C A C C A G T T T T G T G G A T A A T G A  
 1600 1610 1620

THR ILE GLY GLY ILE ILE ASP ARG LYS GLY  
 A A C G A T T G G T G G A A T T A T T G A C C G T A A A G G  
 1630 1640 1650  
 LEU ASN ASN HIS ILE ASN GLU ASP GLU ILE  
 G T T A A A T A A T C A C A T T A A T G A A G A T G A A T  
 1660 1670 1680

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## FIG. 4J

ILE PRO SER ASP ASP SER TYR TYR GLY TYR  
T A T T C C C A G T G A T G A T A T T A T G G A T A  
1690 1700 1710  
THR TRP GLY LYS PRO GLU LYS GLN PHE THR  
T A C T T G G G C C A A G C C A G A G A G C A G T T C A C  
1720 1730 1740

LYS LYS VAL SER SER SER THR GLN VAL VAL  
C A A A A A G T C A G C A G C A C C C A A G T C G T  
1750 1760 1770  
PRO ALA TYR PHE GLY GLN HIS ASP LYS PHE  
G C C A G C T T A T T T T G G G C C A A C A T G A T A A T T  
1780 1790 1800

TYR PHE ASN GLY ASN TYR TYR ASP LEU SER  
T T A T T T A A T G G C A A C T A T T A T G A C C T A T C  
1810 1820 1830  
ALA SER ARG ARG VAL ASP LYS LEU ALA PRO ALA  
A G C C A G T C G T G T T G A T A A A T T A G C C C C T G C  
1840 1850 1860

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FIG.4K

ASP ALA VAL LYS ALA ASN GLN SER ILE LYS  
CGATGCTGTCAAAGCCCAACCAATCCATTAA  
1870 1880 1890  
GLU LYS TYR PRO ASN ALA THR LEU ASN LYS  
AGAAATAATACCCCTAATGCCACCTAATAA  
1900 1910 1920  
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ASP ASN GLN VAL THR ALA ILE VAL LEU GLN  
GGACACCAAGTTACCGCCATCTGCTACA  
1930 1940 1950  
GLU ALA LYS ASP ASN LYS PRO TYR THR ALA  
AGAGCCAAAGATAATAAGCCTTATACCGC  
1960 1970 1980  
ILE ARG ALA LYS SER TYR GLN HIS ILE SER  
CATTCGTGCCAAAGCTATCAGCACATCAG  
1990 2000 2010  
PHE GLY GLU THR LEU TYR ASN ASP ALA ASN  
TTTTGGCGAGACCGCTGTATAACGATGCCAA  
2020 2030 2040

## FIG.4L

GLN THR PRO THR ARG SER TYR PHE VAL GLN  
 C C A A C C C A C A C G C A G T T A T T T G T G C A  
 2050 2060 2070  
 GLY GLY ARG ALA ASP THR SER THR LEU  
 A G G C G G T A G G G C A G A T A C C A G C A C A C T T T  
 2080 2090 2100  
 PRO GLN ALA GLY LYS PHE THR TYR ASN GLY  
 G C C C A G G C A G G T A A A T T C A C T T A C A A C G G  
 2110 2120 2130  
 LEU TRP ALA GLY TYR LEU THR GLN LYS LYS  
 T C T T T G G C A G G C T A C C T G A C C C A A A A A A  
 2140 2150 2160  
 ASP LYS GLY TYR SER ASP ASN ALA GLU THR  
 G G A C A A A G G T T A T A G C G A T A T G C A G A A A C  
 2170 2180 2190  
 ILE LYS GLU LYS GLY HIS PRO GLY TYR LEU  
 C A T C A A G G A A A A G G T C A T C C A G G T A T C T  
 2200 2210 2220

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FIG.4M

LEU THR GLU ASN PHE THR PRO GLU ASP ASP  
G T T A C C G A A A C T T C A C C C A G A T G A  
2230 2240 2250  
ASP ASP ASP LEU THR ALA SER ASP ASP SER  
T G A C G A T G A T T T G A C C G C A T C T G A T G A T T C  
2260 2270 2280

GLN ASP ASP ASN THR HIS GLY ASP ASP ASP  
A C A G A T G A T A A T A C A C A T G G C G A T G A  
2290 2300 2310  
LEU ILE ALA SER ASP ASP SER GLN ASP ASP  
T T T G A T T G C A T C T G A T G A T T C A C A G A T G A  
2320 2330 2340

ASP ALA ASP ASP GLY ASP ASP SER ASP ASP  
T G A C G C A G A T G G A G A T G A C G A T T C A G A T G A  
2350 2360 2370  
LEU GLY ASP ASP GLY ALA ASP ASP ASP ALA ALA  
T T T G G G T G A T G G T G C A G A T G A T G A C G C C G C  
2380 2390 2400

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FIG.4N

GLY LYS VAL TYR HIS ALA GLY ASN ILE ARG  
AGGCAAGTGTTATCATGCAAGGTAAATATTCG  
2410 2420 2430  
PRO GLU PHE GLU ASN LYS TYR LEU PRO ILE  
CCCTGAATTGTGAACAATACTTGCCCAT  
2440 2450 2460

ASN GLU PRO THR HIS GLU LYS THR PHE ALA  
TAATGAGCCTACTCATGAATAAACCTTTGGC  
2470 2480 2490  
LEU ASP GLY LYS ASN LYS ALA LYS PHE GLU  
CCTAGATGGTAATAATAAGCTAAGTTTGA  
2500 2510 2520

VAL ASP PHE ASN THR ASN SER LEU THR GLY  
AGTGGATTTTAACACCAACAGCCTAAC TTGG  
2530 2540 2550  
LYS LEU ASN ASP GLU ARG GLY ASP ILE VAL  
TAAATTAAACGATGAGAGGATATCGT  
2560 2570 2580

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## FIG. 4O

PHE ASP ILE LYS ASN GLY LYS ILE ASP GLY  
C T T G A T A T C A A A A T G G C A A A T T G A T G G  
2590 2600 2610  
THR GLY GLY PHE THR ALA LYS ALA ASP VAL PRO  
C A C A G G A T T T A C C G C C A A A G C C G A T G T G C C  
2620 2630 2640

ASN TYR ARG GLU GLU VAL GLY ASN ASN GLN  
A A C T A T C G T G A A G A A G T G G G T A A C A C C A  
2650 2660 2670  
GLY GLY GLY PHE LEU TYR ASN ILE LYS ASP  
A G G T G G C G G T T C T T A T A C A C A T C A A G A  
2680 2690 2700

ILE ASP VAL LYS GLY GLN PHE PHE GLY THR  
T A T G A T G T T A A G G G C A A T T T T T G G C A C  
2710 2720 2730  
ASN GLY GLU GLU LEU ALA GLY GLN LEU HIS  
A A A T G G C G A A G A G T T G G C A G G A C A G T T A C A  
2740 2750 2760

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FIG. 4P

HIS ASP LYS GLY ASP GLY ILE ASN ASP THR  
 TCA TGACA AAGCGATGGCAATCAATGACAC  
 2770 2780 2790  
 ALA GLU LYS ALA GLY ALA VAL PHE GLY ALA  
 CGCCGA AAGCAAGGGCTGTCTTTGGGGC  
 2800 2810 2820  
 VAL LYS ASP LYS \*\*\*  
 TGTTAAGATAATAAGCCCCCTTCATC  
 2830 2840 2850  
 ATCGTTTAGTCGCTTGACCGACAGTTGATG  
 2860 2870 2880  
 ACGCCCTTGGCAATGTCTTAACAAGCACT  
 2890 2900 2910  
 TTGAACAGTGCCCTTGGGCGAATTCTTGA  
 2920 2930 2940  
 TAAATGCCACGATTTGCCCTTGGGCTAATA  
 2950 2960 2970  
 -10  
 TCTTGATATAACATCGCCATATAATAGAA  
 2980 2990 3000  
 -35

FIG. 4Q.

Lbp1  
 RBS  
 A T A A G T T A G G A T T T T T A T G T C A A A A  
 3010 3020 3030  
 SER ILE THR LYS THR GLN THR PRO SER VAL H  
 T C T A T C A C A A A A C A C A A C A C C A T C A G T C C  
 3040 3050 3060  
 IS THR MET THR THR HIS ARG LEU ASN LEU  
 A T A C C A T G A C C A C G C A C C G C T T A A C C T T G  
 3070 3080 3090  
 ALA ILE LYS ALA ALA LEU PHE GLY VAL ALA V  
 C C A T C A A A G C G G C G T T A T T G G T G T G C C A G  
 3100 3110 3120  
 AL LEU PRO LEU SER VAL TRP ALA GLN GLU  
 T T T A C C C C T A T C C G T C T G G G C C A A G A G A  
 3130 3140 3150  
 ASN THR GLN THR ASP ALA ASN SER ASP ALA L  
 A C A C T C A G A C A G A T G C C A A C T C T G A T G C C A  
 3160 3170 3180

# FIG.4R

YS ASP THR LYS THR PRO VAL VAL TYR LEU  
 A G A C A C A A A A C C C T G T C T A T T A G  
 3190 3200 3210  
 ASP ALA ILE THR VAL THR ALA ALA PRO SER A  
 A T G C C A T C A C G G T A A C C G C C C A T C T G  
 3220 3230 3240

LA PRO VAL SER ARG PHE ASP THR ASP VAL  
 C C C T G T T C T C G G T T T G A C A C C G A T G T A A  
 3250 3260 3270  
 THR GLY LEU GLY LYS THR VAL LYS THR ALA A  
 C A G G G C T T G G C A A A C C G T C A A A C C G C T G  
 3280 3290 3300

SP THR LEU ALA LYS GLU GLN VAL GLN GLY  
 A C A C G C T G G C A A A G A A C A G T A C A G G C A  
 3310 3320 3330  
 ILE ARG ASP LEU VAL ARG TYR GLU THR GLY V  
 T T C G T G A T T G G T G C G T A T G A A C T G G G  
 3340 3350 3360

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FIG.4S

AL SER VAL VAL GLU GLN GLY GLY ARG GLY GLY  
T G A G T G T G G T T G A G C A G G G C G T G G T G G C A  
3370 3380 3390  
SER SER GLY PHE ALA ILE HIS GLY VAL ASP L  
G C A G C G G A T T T G C C A T T C A T G G C G T G G A T A  
3400 3410 3420

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YS ASN ARG VAL GLY ILE THR VAL ASP GLY  
A A A C C G A G T G G G C A T T A C C G T A G A T G G C A  
3430 3440 3450  
ILE ALA GLN ILE GLN SER TYR LYS ASP GLU S  
T T G C C C A A A T T C A A T C C T A C A A G A C G A A T  
3460 3470 3480

ER THR LYS ARG ALA GLY ALA GLY SER GLY  
C C A C T A A G C G A G C T G G G C A G G C T C T G G G G  
3490 3500 3510  
ALA MET ASN GLU ILE GLU ILE GLU ASN ILE A  
C G A T G A A C G A G A T A G A G A T T G A A A C A T T G  
3520 3530 3540

FIG.4T

LA ALA VAL ALA ILE ASN LYS GLY GLY ASN  
 CCGCCGTTGCCCATCATTAAGGCGGTAAATG  
 3550 3560 3570  
 ALA LEU GLU ALA GLY SER GLY ALA LEU GLY G  
 CCTTAGAAGCAGGCTCTGGTGCGTGGTG  
 3580 3590 3600

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 LY SER VAL ALA PHE HIS THR LYS ASP VAL  
 GTTCGGTGCGGTTTCATACCAAGATGTA  
 3610 3620 3630  
 SER ASP VAL LEU LYS SER GLY ASN LEU G  
 GCGATGTCCTTAATAATCTGGTAACATCTTG  
 3640 3650 3660

LY ALA GLN SER LYS THR THR TYR ASN SER  
 GTGCTCAAGCAAAACCACTTATACAGCA  
 3670 3680 3690  
 LYS ASN ASP HIS PHE SER SER GLN THR LEU ALA A  
 AAAATGACCATTTTAGTCAGACGCTGGCAG  
 3700 3710 3720

## FIG. 4U

LA ALA GLY LYS THR GLU ARG VAL GLU ALA  
 C G G C A G G T A A A C C G A G C G T G T G G A A G C G A  
 3730 3740 3750  
 MET VAL GLN TYR THR TYR ARG LYS GLY LYS G  
 T G G T G C A A T A T A C C T A C C G T A A G G C A A A G  
 3760 3770 3780

LU ASN LYS LYS ALA HIS SER ASP LEU ASN GLY  
 A A A C A A A G C A C A C A G C G A C C T A A A T G G C A  
 3790 3800 3810  
 ILE ASN GLN SER LEU TYR ARG LEU GLY ALA T  
 T C A A C C A A A G C C T A T A T C G C T T G G T G C A T  
 3820 3830 3840

RP GLN GLN LYS TYR ASP LEU ARG LYS PRO  
 G G C A A C A A A A T A T G A T T T A A G A A A G C C T A  
 3850 3860 3870  
 ASN GLU LEU PHE ALA GLY THR SER TYR ILE T  
 A C G A A C T G T T T G C A G G C A C A G C T A T A T C A  
 3880 3890 3900

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## FIG.4V

HR GLU SER CYS LEU ALA SER ASP ASP PRO  
 C C G A A A G C T G T T T G G C A A G T G A T G A C C C A A  
 3910 3920 3930  
 LYS SER CYS VAL GLN TYR PRO TYR VAL TYR T  
 A A A G C T G C G T A C A A T A C C C T T A T G T C T A C A  
 3940 3950 3960

HR LYS ALA ARG PRO ASP GLY ILE GLY ASN  
 C C A A A G C C C G A C C A G A T G G T A T C G G C A A T C  
 3970 3980 3990  
 ARG ASN PHE SER SER GLU LEU SER ASP ALA GLU L  
 G C A A T T T T C T G A G T T A A G C G A T G C T G A A A  
 4000 4010 4020

YS ALA GLN TYR LEU ALA SER THR HIS PRO  
 A A G C A C A A T A T T T G G C G T C C A C G C A C C C C C  
 4030 4040 4050  
 HIS GLU VAL VAL SER ALA LYS ASP TYR THR G  
 A T G A G G T T G T C T C T G C C A A A G A T T A T A C A G  
 4060 4070 4080

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## FIG. 4W

LY THR TYR ARG LEU LEU PRO ASP PRO MET  
 G C A C T T A T C G G T T G T T A C C T G A C C C A T G G  
 4090 4100 4110  
 ASP TYR ARG SER ASP SER TYR LEU ALA ARG L  
 A C T A T C G T T C A G A C T C G T A T T G G C A C G C C  
 4120 4130 4140

EU ASN ILE LYS ILE THR PRO ASN LEU VAL  
 T T A A C A T C A A A T C A C C C A A A T T G G T C A  
 4150 4160 4170  
 SER LYS LEU LEU LEU GLU ASP THR LYS GLN T  
 G T A A A C T G T T A T T A G A A G A C C A A G C A A A  
 4180 4190 4200

HR TYR ASN ILE ARG ARG ASP MET ARG HIS CYS S  
 C A T A C A C A T T C G T G A T A T G C G T C A T G T A  
 4210 4220 4230  
 ER TYR HIS GLY ALA ARG LEU GLY ASN ASP G  
 G T T A T C A T G G G G C A A G A T T G G C A A T G A C G  
 4240 4250 4260

FIG.4X

LY LYS PRO ALA ASN GLY GLY SER ILE VAL  
G T A G C C T G C C A A T G G C G G C T C C A T T G T C C  
4270 4280 4290  
LEU CYS ASP ASP TYR GLN GLU TYR LEU ASN A  
T T T G C G A T G A T T A T C A A G A G T A T C T A A A T G  
4300 4310 4320

LA ASN ASP ALA SER GLN ALA SER PHE ARG  
C C A A T G A C G C A T C A C A G C A T C A T T T A G A C  
4330 4340 4350  
PRO GLY ALA ASN ASP ALA PRO ILE PRO LYS L  
C A G G G C C T A A T G A C G C C C C A T T C C A A A C  
4360 4370 4380

EU ALA TYR ALA ARG SER SER VAL PHE ASN  
T G G C T T A T G C C A G A A G C A G T G T T T A A C C  
4390 4400 4410  
GLN GLU HIS GLY LYS THR ARG TYR GLY LEU G  
A A G A G C A T G G C A A A C T C G C T A T G G G T T A G  
4420 4430 4440

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# FIG.4Y

LY PHE GLU PHE LYS PRO ASP THR PRO TRP  
 G T T T G A G T T T A A G C C T G A C A C G C C A T G G T  
 4450 4460 4470  
 PHE LYS GLN ALA LYS LEU ASN LEU HIS GLN G  
 T T A A C A A G C A A A T T A A C C T A C A T C A A C  
 4480 4490 4500

IN ASN ILE GLN ILE ILE ASN HIS ASP ILE  
 A A A T A T C C A A A T C A T T A A C C A T G A C A T T A  
 4510 4520 4530  
 LYS LYS SER CYS SER GLN TYR PRO LYS VAL A  
 A A A A T C G T G C A G C C A A T A T C C C A A G G T G G  
 4540 4550 4560

SP LEU ASN CYS GLY ILE SER GLU ILE GLY  
 A T T A A A T G T G G C A T C A G T G A A A T T G G G C  
 4570 4580 4590  
 HIS TYR GLU TYR GLN ASN TYR ARG TYR L  
 A T T A T G A A T A T C A A A C A A T T A C C G T T A T A  
 4600 4610 4620

FIG.4Z

YS GLU GLY ARG THR THR LEU THR GLY LYS  
A A G A G G C C G T A C C A G T T T G A C A G G C A A A C  
4630 4640 4650  
LEU ASP PHE ASP PHE ASN PHE LEU LEU GLY GLN H  
T T G A T T T A A T T T G A C C T G C T G G G C C A G C  
4660 4670 4680

IS ASP LEU THR VAL LEU ALA GLY ALA ASP  
A C G A T T G A C G G T G T T G G C T G G T G C A G A T A  
4690 4700 4710  
LYS VAL LYS SER SER GLN PHE ARG ALA ASN ASN P  
A A G T T A A A G C C A A T T C G T G C C A A C A C C  
4720 4730 4740

RO ARG ARG THR ILE ILE ASP THR THR GLN  
C C A G A C G C A C A A T C A T T G A C A C C C A A G  
4750 4760 4770  
GLY ASP ALA ILE ILE ASP GLU SER THR LEU T  
G C G A T G C C A T C A T T G A T G A A A G C A C G C T G A  
4780 4790 4800

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FIG. 4A'

HR ALA GLN GLU GLN ALA LYS PHE LYS GLN  
CAGCACAGGAGCAGCCAAATTTAAGCAAT  
4810 4820 4830  
SER GLY ALA ALA TRP ILE VAL LYS ASN ARG L  
CAGGGGCGCAGCATGGATTGTCAAAATCGCT  
4840 4850 4860

EU GLY ARG LEU GLU GLU GLU LYS ASP ALA CYS  
TAGGACGCTTAGAGAGAAAGACCGCTGTG  
4870 4880 4890  
GLY ASN ALA ASN GLU CYS GLU ARG ALA PRO I  
GCAATGCCCAATGAATGTGAACGGCGCCCA  
4900 4910 4920

LE HIS GLY SER ASN GLN TYR VAL GLY ILE  
TTCA TGGCAGTAACCAATA TGTGGCA TTA  
4930 4940 4950  
ASN ASN LEU TYR THR PRO ASN ASP TYR VAL A  
ACAACCTTTATACCAACAATGATTGTGG  
4960 4970 4980

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## FIG. 4B'

SP LEU SER PHE GLY GLY ARG LEU ASP LYS  
 A T T A A G T T T G G T G G A C G C T T G G A T A A A C  
 4990 5000 5010  
 G L N A R G I L E H I S S E R T H R A S P S E R A S N I L E I  
 A A C G C A T T C A C A G C A C C G A T T C A A A C A T C A  
 5020 5030 5040

LE SER LYS THR TYR THR ASN LYS SER TYR  
 T C A G C A A A C T T A C A C C A A C A A A G C T A T A  
 5050 5060 5070  
 A S N P H E G L Y A L A A L A V A L H I S L E U T H R P R O A  
 A T T T G G A G C G G C G G T T C A T C T G A C A C C T G  
 5080 5090 5100

SP PHE SER LEU LEU TYR LYS THR ALA LYS  
 A T T T A G C C T G T T G T A T A A A C T G C C A A A G  
 5110 5120 5130  
 G L Y P H E A R G T H R P R O S E R P H E T Y R G L U L E U T  
 G C T T T C G T A C G C C A A G T T T T A T G A A C T G T  
 5140 5150 5160

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FIG.4C'

YR ASN TYR ASN SER THR ALA ALA GLN HIS  
A C A C T A T A C A G C A C C G C C C A G C A T A  
5170 5180 5190  
LYS ASN ASP PRO ASP VAL SER PHE PRO LYS A  
A A A T G A C C C T G A T G T G T C T T T C C C A A A C  
5200 5210 5220

RG ALA VAL ASP VAL LYS PRO GLU THR SER  
G A G C G T T G A T G T C A A C C T G A A C T T C C A  
5230 5240 5250  
ASN THR ASN GLU TYR GLY PHE ARG TYR GLN H  
A T A C C A A T G A A T A C G G C T T T C G C T A T C A G C  
5260 5270 5280

IS PRO TRP GLY ASP ILE GLU MET SER MET  
A C C C T T G G G G G A T A T T G A G A T G A G C A T G T  
5290 5300 5310  
PHE LYS SER ARG TYR LYS ASP MET LEU ASP L  
T C A A A A G C C G T T A C A A G G A C A T G T T A G A T A  
5320 5330 5340

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FIG.4D'

YS ALA ILE PRO ASN LEU THR LYS ALA GLN  
 A G C C A T A C C G A A C C T A A C C A A G C C C A G C  
 5350 5360 5370  
 GLN GLU TYR CYS LYS ALA HIS LEU ASP SER A  
 A A G A G T A T T G T A A G G C T C A T T G G A T T C C A  
 5380 5390 5400

SN GLU CYS VAL GLY ASN PRO PRO THR PRO  
 A T G A A T G T G T T G G T A A T C C A C C C A C G C C C A  
 5410 5420 5430  
 LYS THR SER ASP GLU VAL PHE ALA ASN LEU T  
 A A C C A G T G A T G A G G T A T T G C C A A C T T A T  
 5440 5450 5460

YR ASN ALA THR ILE LYS GLY VAL SER VAL  
 A T A A T G C C A C C A T C A A A G G G T G A G T G T C A  
 5470 5480 5490  
 LYS GLY LYS LEU ASP LEU HIS ALA MET THR S  
 A A G G C A A A C T G G A T T G C C A T G C C A T G A C A T  
 5500 5510 5520

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# FIG.4E'

```

ER  LYS  LEU  PRO  ASP  GLY  LEU  GLU  MET  THR
C A A A C T G C C A G A T G G T C T T G A A A T G A C C T
5530
LEU  GLY  TYR  GLY  HIS  THR  LYS  LEU  GLY  LYS  P
T G G G T T A T G G T C A T A C C A A A T T G G G G A A A T
5560
5570
5580

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```

HE  ASP  TYR  ILE  ALA  PRO  LYS  ASP  ALA  ASP
T T G A T T A C A T T G C A C C C A A A G A T G C C G A T G
5590
5600
GLY  TRP  TYR  GLN  ALA  ARG  PRO  ALA  PHE  TRP  A
G T T G G T A T C A G G C T C G C C C T G C T T T T G G G
5620
5630
5640

```

```

SP  ALA  ILE  THR  PRO  ALA  ARG  TYR  VAL  VAL
A T G C C A T C A C C C A G C G C C T A T G T G T C C G
5650
5660
5670
GLY  LEU  ASN  TYR  ASP  HIS  PRO  SER  GLN  VAL  T
G T C T A A A C T A T G A C C A C C C A G T C A A G T A T
5680
5690
5700

```

## FIG.4F'

RP GLY ILE GLY THR THR LEU THR HIS SER  
GGGCA TTGGCA CAACTTTAACGCACAGCA  
5710 5720 5730  
LYS GLN LYS ASP GLU ASN GLU LEU SER ALA L  
A C A A A A G A T G A A A T G A G C T A A G T G C C C  
5740 5750 5760

EU ARG ILE ARG ASN GLY LYS ARG GLU ILE  
TTAGAA TCCGA AATGGCA AAGAGAAATAC  
5770 5780 5790  
GLN THR LEU THR HIS THR ILE PRO LYS ALA T  
A A C C T T A C G C A C A C A T A C C C A A A G C C T  
5800 5810 5820

YR THR LEU LEU ASP MET THR GLY TYR TYR  
ATACCTTACTGGACATGACAGGCTATTATA  
5830 5840 5850  
SER PRO THR GLU SER ILE THR ALA ARG LEU G  
G C C A A C T G A G A G C A T C A C C G C T C G T C T G  
5860 5870 5880

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FIG. 4G'

LY ILE ASN ASN VAL LEU ASN THR ARG TYR  
 G T A T C A A C A A T G T A T T A A A C A C C G C T A  
 5890 5900  
 THR THR TRP GLU ALA ALA ARG GIN LEU PRO S  
 C A C C A C A T G G G A A G C G G C A C G C C A A C T G C C C A  
 5910 5920 5930 5940

ER GLU ALA ALA SER SER THR GIN SER THR  
 G C G A A G C T G C A A G C A G T A C C C A A T C A A C C C  
 5950 5960 5970  
 ARG TYR ILE ALA PRO GLY ARG SER TYR PHE A  
 G T T A C A T T G C A C C A G G T C G C A G T T A C T T T G  
 5980 5990 6000

LA SER LEU GLU MET LYS PHE \*\*\* MET THR  
 C C A G T C T T G A A A T G A A G T T T A A T A T G A C C  
 6010 6020 6030  
 CYS LEU PRO LYS THR ASN PRO ALA LEU LYS  
 T G T T T A C C A A A G A C C A C C C T G C T T T A A A A  
 6040 6050 6060

81 / 130

FIG.4H'

VAL LYS HIS ARG PHE LEU LYS GLN VAL LEU  
G T C A A G C A C A G A T T T T A A A G C A G G T G C T G  
6070 6080 6090  
LEU LEU LEU CYS VAL ASP THR LEU THR ALA  
T T A T T G C T T T G T G T T G A T A C A T T A A C A G C A  
6100 6110 6120

82/130  
GLN ALA TYR ALA HIS SER HIS HIS THR PRO  
C A G G C G T A C G C C C A C A G C C A T C A T A C G C C C  
6130 6140 6150  
ILE HIS THR PRO THR HIS GLU LEU SER SER  
A T T C A T A C A C C C A C G C A T G A G C T G T C A T C T  
6160 6170 6180

ALA ASP ALA LEU SER ASP GLU GLY LEU GLY  
G C T G A T G C T T T A T C A G A T G A A G G C T T G G G T  
6190 6200 6210  
LYS ASP LEU GLY SER LEU ASP SER PRO ASP  
A A G G A T T T G G G C A G T T T G G A C A G C C A G A T  
6220 6230 6240



FIG. 4I'

GLY LEU GLY ASP GLY LEU GLY LEU  
 GGT TGGGTGATGGTTTAGGGCGATGGTTTG  
 6250 6260 6270  
 GLY ASP GLY LEU LYS SER ASP LYS THR PRO  
 GGTGATGGCTTAAAGTGATAAACCCCT  
 6280 6290 6300

LEU PRO ILE ASN ALA LEU THR VAL ASN GLN  
 TTACCCATCAACGCCCTTGACCGTTATCAG  
 6310 6320 6330  
 SER ASN GLU SER GLN PRO ALA PRO PRO SER  
 AGCAATGAGAGCCAGCCCTGCCCAACCGAGC  
 6340 6350 6360

83/130

VAL ASP VAL ASN PHE LEU LEU ALA GLN PRO  
 GTAGATGTCATAATTTTACTTGCCCAAGCCA  
 6370 6380 6390  
 GLU ALA PHE TYR HIS VAL PHE HIS GLN ALA  
 GAGGCATTTTATCATGTCTTTTCATCAGCG  
 6400 6410 6420

FIG.4J'

ILE VAL GLN ASP ASP VAL ALA THR LEU ARG  
A T T G T G C A A G A T G A T G T G G C A A C A T T A C G C  
6430 6440 6450  
LEU LEU LEU PRO PHE TYR ASP ARG LEU PRO  
T T G T T A T T G C C A T T T T A T G A C C G C C T G C C T  
6460 6470 6480

84/130  
ASP ASP TYR GLN ASP ASP VAL LEU LEU LEU  
G A T G A T T A T C A A G A T G A T T T G T T G T T A  
6490 6500 6510  
PHE ALA GLN SER LYS LEU ALA LEU SER ASP  
T T T G C C C A A A G T A A A C T T G C C C T A A G T G A T  
6520 6530 6540

GLY ASN THR LYS LEU ALA LEU ASN LEU LEU  
G G C A A T A C C A A A T T G G C A T T G A A T C T G C T G  
6550 6560 6570  
THR ASP LEU SER SER ASN LYS GLU PRO THR LEU  
A C C G A T T T G A G T A A C A A A G A G C C A A C A C T T  
6580 6590 6600

FIG.4K'

THR ALA VAL LYS LEU GLN LEU ALA SER LEU  
 A C G C G G T A A A T T A C A C T T G C T T C C T T G  
 6610 6620 6630  
 LEU LEU THR ASN LYS HIS ASP LYS HIS ALA  
 T T G C T G A C C A C A A G C A C G A T A A C A C G C C  
 6640 6650 6660

GLN MET VAL LEU ASP GLU LEU LYS ASP ASP  
 C A A T G G T G C T A G A T G A C T C A A G A T G A T  
 6670 6680 6690  
 ALA HIS PHE LEU LYS LEU SER LYS LYS GLU  
 G C C A C T T T T A A A T T A A G C A A A A G A G  
 6700 6710 6720

GLN ARG TRP VAL LEU SER GLN SER ARG TYR  
 C A A G A T G G G T G C T A T C G C A A A G T C G C T A T  
 6730 6740 6750  
 LEU HIS LYS LYS TYR LYS MET GLY LEU ASP  
 T T A C A T A A A A T A T A A A T G G C T T G G A T  
 6760 6770 6780

85/130

FIG. 4L'

LEU GLY ILE ASN TYR LEU HIS LEU ASP ASN  
 TTGGGCATCAACTATCTGCATTGTGATAAT  
 6790 6800 6810  
 ILE ASN ALA ALA SER THR ILE THR GLN PRO  
 ATCAACGCCCTCCACCATCAACCAAGCCC  
 6820 6830 6840

ASN ILE LYS LYS ASP ALA PRO LYS PRO ALA  
 AACATTAAAGATGCCCAAAACCTGCT  
 6850 6860 6870  
 HIS GLY LEU ALA LEU SER LEU GLY VAL ASN  
 CATTGGCCTTGCCCTTATCGCTTGTTGTAAT  
 6880 6890 6900

LYS TYR THR PRO LEU SER HIS GLY MET SER  
 AATACACGCCCTTAGTCAATGGCAATGAGT  
 6910 6920 6930  
 ILE TYR THR ALA LEU ASP VAL ASP GLY LYS  
 ATTTATACAGCCCTAGATGTTGATGTTAA  
 6940 6950 6960

86 / 130

FIG. 4M'

PHE TYR ASP ASP LYS SER HIS ASN GLU LEU  
 TTTTATGATGACAAAGCCACAAATGAACTG  
 6970 6980 6990  
 ALA VAL PHE ALA HIS ALA GLY LEU ARG LYS  
 GCGGTTTTTGGCTCATGCTGGACCTAAGAA  
 7000 7010 7020

ASP HIS GLN LYS GLY TYR VAL ASP VAL VAL  
 GATCACCAAAAGGTTATGTTGATGTCGTA  
 7030 7040 7050  
 PRO PHE VAL GLY ARG ILE PHE ALA THR ASN  
 CCTTTGTTGGGCGGTATTTTGGCCACCAAT  
 7060 7070 7080

GLN GLN HIS GLY ARG LEU SER PRO ARG LYS  
 CAGCAGCATGGCAGATTATCCCCAGAA  
 7090 7100 7110  
 ASP SER GLN GLY VAL ALA PHE GLY SER HIS  
 GACAGTCAGGCGGTGGCGTTTGGCAGCAT  
 7120 7130 7140

87/130

# FIG.4N'

HIS ARG ILE ASN ASP LYS TRP GLN ASN ALA  
 C A T C G G A T C A A T G A T A A A T G G C A A A T G C G  
 7150 7160 7170  
 PHE PHE ALA ARG MET GLU LYS GLY ASN TYR  
 T T T T T G C A C G C A T G G A A A A G G C A A T T A T  
 7180 7190 7200

THR GLU HIS TYR GLN GLY TYR ASP GLY LYS  
 A C C G A G C A T T A T C A A G G T T A T G A T G G C A A G  
 7210 7220 7230  
 ARG TYR HIS VAL ASN ASP THR ILE LEU LEU  
 C G T T A T C A T G T G A A T G A C A C C A T T T G T T G  
 7240 7250 7260

GLN ASP GLY PRO ASN ARG ARG TYR SER LEU  
 C A A G A T G G C C C A A A T C G T C G T T A C T C T T T G  
 7270 7280 7290  
 GLY VAL GLY TYR GLN LEU SER HIS LEU GLN  
 G G C G T G G G G T A T C A G C T T A G C C A T C T G C A A  
 7300 7310 7320

88/130

FIG. 4O'

ASP ALA THR LYS SER SER HIS ALA THR LYS  
 G A T G C A C A A A A G C A G T C A T G C C A C A A A G  
 7330 7340 7350  
 ILE HIS PHE GLY VAL LEU GLN ARG LEU PRO  
 A T A C A T T T T G G G G T G T T G C A A A G A T T G C C A  
 7360 7370 7380

89/130

ASN GLY LEU THR VAL GLN GLY ARG VAL SER  
 A A T G G T C T G A C C G T G C A A G G T A G A G T G A G T  
 7390 7400 7410  
 ALA GLU ARG GLU ARG TYR HIS GLY LYS LEU  
 G C T G A G C G T G A G C G T T A T C A T G G T A A A T T A  
 7420 7430 7440

LEU ARG LEU VAL ASN PRO ASP ASP VAL TYR  
 T T G C G T C T G G T T A A T C C T G A T G T G T A T  
 7450 7460 7470  
 ARG THR ASP LYS THR LEU THR LEU GLN THR  
 C G C A C A G A T A A A C C C T A A C C C T A C A A C C  
 7480 7490 7500

FIG.4P'

SER ILE TRP HIS LYS ASP ILE HIS TRP LEU  
TCCATTGGCCACAAAGACATTCTGGCTT 7510  
7520  
GLY LEU THR PRO LYS LEU THR TYR ARG TYR  
GGATTACGCCCAAAGCTGACTTATCGTTAC 7540  
7550  
7560

SER LYS ASN ASN SER ASN LEU PRO ALA LEU  
AGTAAATAACAGTAACCTTACCGCACTT 7570  
7580  
TYR SER HIS ASN LYS LYS GLN ASN PHE TYR LEU  
TATAGCCATAACAACAATAATTATTG 7600  
7610  
7620

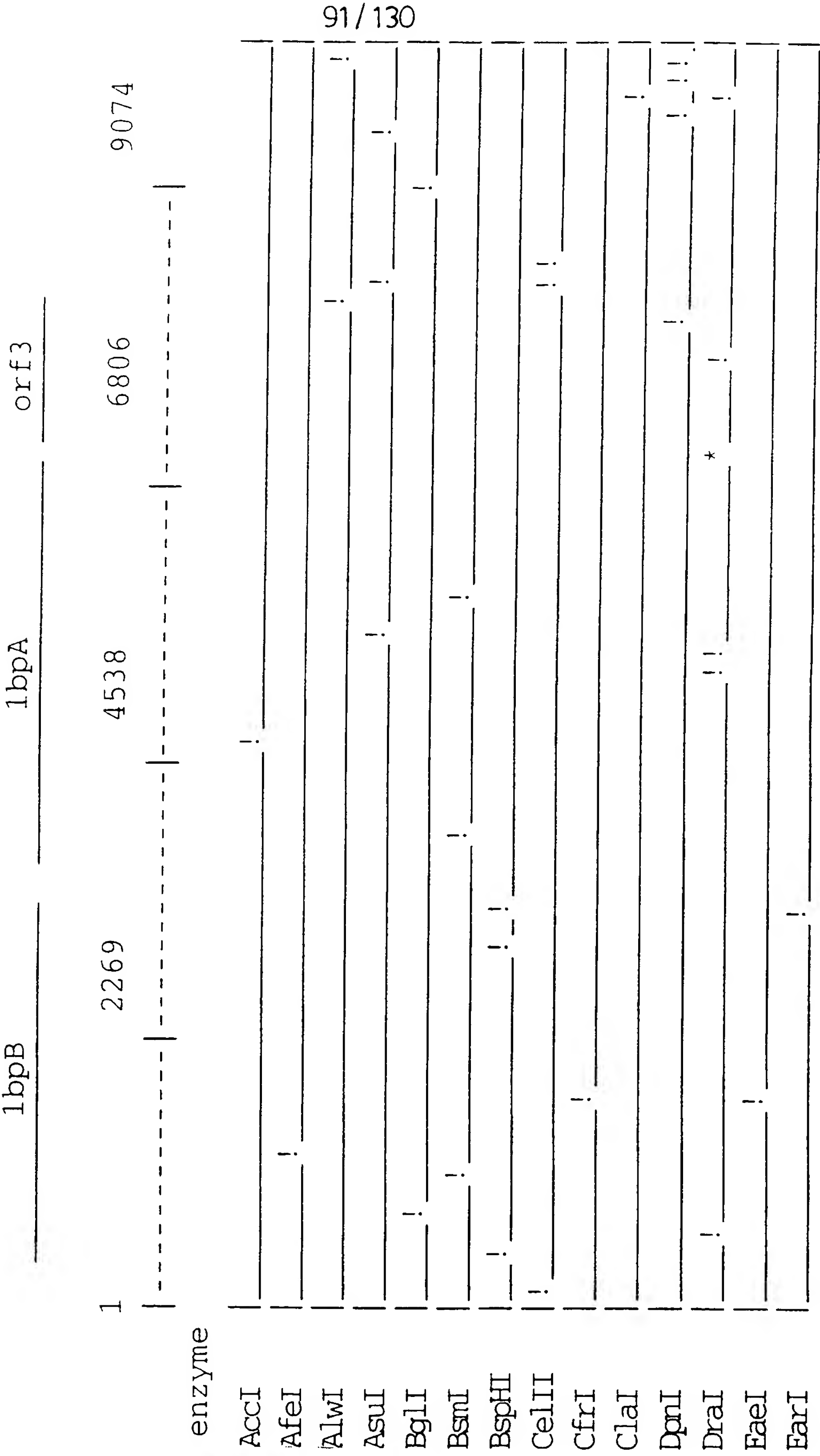
GLU LEU GLY ARG SER PHE \*\*\*  
GAGCTTGGTCGGTCGTTTATA 7630  
7640

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Restriction map of clone pLDW1, *M. catarrhalis* strain Q8 1fr



SUBSTITUTE SHEET (RULE 26)

FIG.5A



FIG.6A

Alignment of Lbp2 proteins

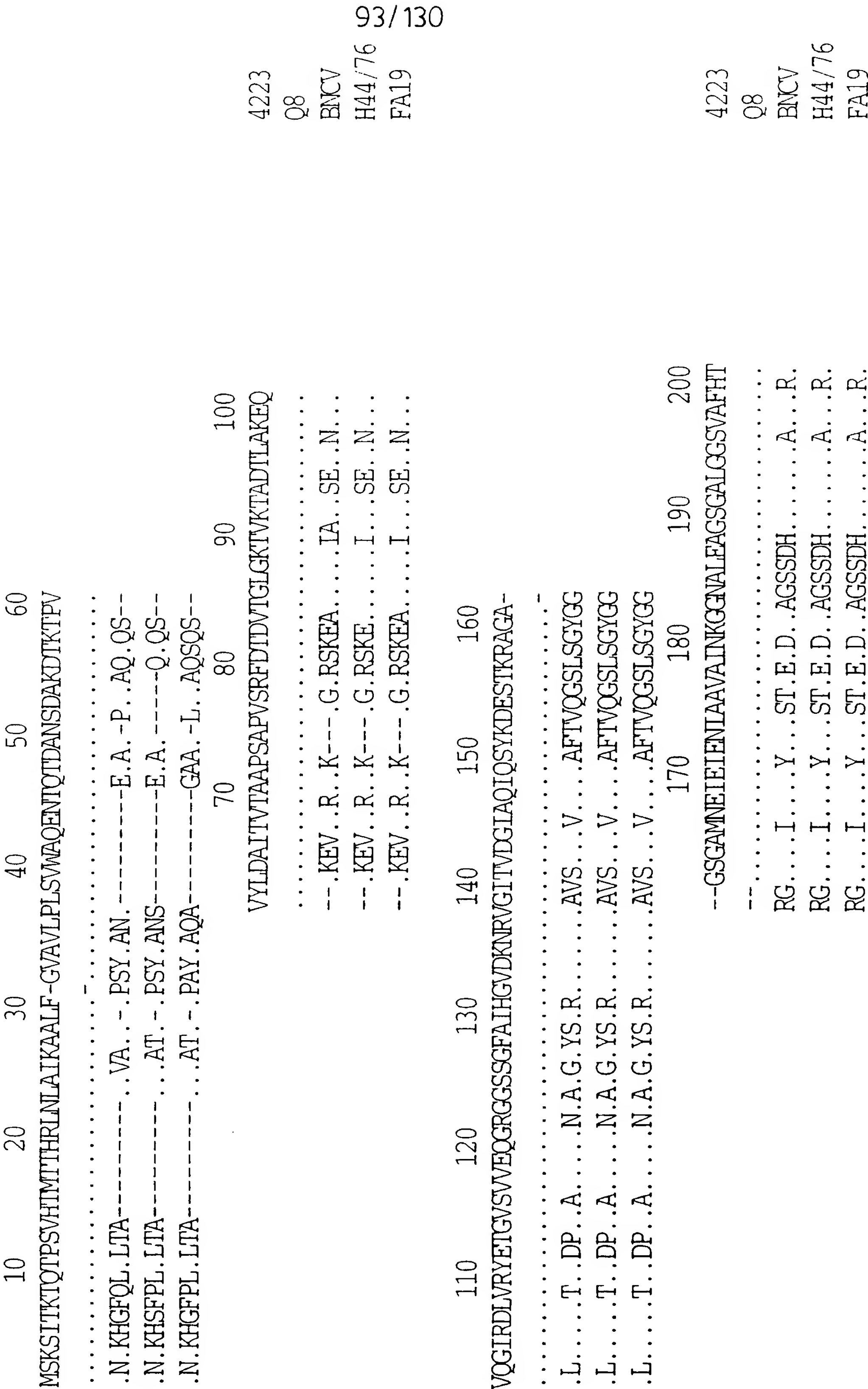


FIG.6B

210	220	230	240	250	260	
KDVSDVLKSGKNLGAQSKTTYNKNDHFSQTLAAAGKTERVEAMVOYTYRKGENKAHSDL						
.....N.....						
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.THP.G.I						
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.TRP.G.I						
.EAA.LISD..SW.I.A..A.G...RQ.MKS.G.GFSKDGW.GLLIR.E.Q.R.TRP.G.I						
		270	280	290	300	
		-NGINQSLYRLGAWQOKYDL-RKPNELFAGTSYITESCLAS				4223
		-.....T.....				Q8
		AD.VAYGIN..D.FR.T.GI-K..S.GGEYFLAEG..E.KP				BNCV
		AD.VAYGID..D.FR.T..I-Q.QNKKAIFYFLAEG..E.KP				H44/76
		AD.VAYGID..D.FR.T..IK..TT.P.--FLAEG.NT.KP				FA19
						94 / 130
310	320	330	340	350	360	
DDPKSCVQYPYVYTKARPDGIGNRNFSELSAEKAQYLASTHPHEVVSADYTGTYRLLPD						
.....T.....						
VAKVAGNGNYLNNQLN.WVKERIEQNQP..AE.E.MVREAQAR..NL..QA...GG.I...						
AAKLACNGNYLNNQLN.WVEERKKNQS..AE.E.MVREAQAR..NL..QA...GG.I...						
VAKLAGYGIYLNRLN.WVKERIEQNQP..AE.E..VREAQAR..NL..QA...GG.I...						
	370	380	390	400		
	PMDYRSDSYLARLNKIKITPNLVSKLLLEDTKQTYNIRDM					4223
	.....					Q8
	.....G.W..K.GYRFGGRHYVGVF.....R.D....					BNCV
	.....G.W..K.GYRFGGRHYVGVF.....R.D....					H44/76
	.....G.W..K.GYRFGGRHYVGVF.....R.D....					FA19



FIG.6C

410	420	430	440	450	460		
RHCSYHGARLGNCGKPANGSGTIVLCDDYQEYLNANDASQALFRPGANDAPIPKLAYARSSV							
.....S.....							
TEKQ.Y.TDEAKKFRDKS.—VYDG..FRDG.YFVFNIEE-WKGDQKLIRGIG.K.S.TK-							
TEKQ.Y.TDEATKFSDKS.—VYDG..FRDG.YFVFNIEE-WKGDK.LVKGIG.K.S.TK-							
TEKQ.Y.TDEAEKFRDKS.—VYDG..FRDG.YFVFNIEE-WKGDK.LVKGIG.K.S.TK-							
			470	480	490	500	
FNQEHGKTRY-GLSFEF---KPDTPWFKQAKNLHQQNIQIIN							4223
.....G.....							Q8
.ID..HRR.RM..LYRYENE.YSDN.ADK.V.SFDK.GVATD.							ENCV
.ID..HRR.RM..LYRYENEAYSDN.ADK.V.SFDK.GVATD.							H44/76
.ID..HRR.RM..LYRYENE.YSDN.ADK.V.SFDK.GVATD.							FA19
							95/130
510	520	530	540	550	560		
HDIKKSCSQYPKVDLNCGISEIGHYEYQ—NNYRYKEGRASLTGKLDNFIDL-LGQHDLTVLAG							
.....T.....							
NTL.LN.AV..A..KS.RA.ADKP.S.DSSDRFH.R.QHN.V.NASFEKSLKNKWTK.H..LGF.							
NTL.LN.AV..A..KA.RA.ADKP.S.DSSDRFH.R.QHN.V.NALFEKSLKNKWTK.H..LGF.							
NTL.LN.AV..A..KA.RA.ADKP.S.DSSDRFH.R.QHN.V.NASFEKSLKNKWTK.H..LGF.							
			570	580	590	600	
ADKVKSQFRANPRRTIIDTTQGDALIDESTLTAQEQAK							4223
.....							Q8
Y.ASNAIS.PEQLSHNAARISEYSDYT.KGD-----							ENCV
Y.AS.AIS.PEQLSHNAARISEFSDYA.DGKY-----							H44/76
Y.AS.AIS.PEQLSHNAARISE-STGF..KNQD-----							FA19

FIG.6D

610 620 630 640 650  
FKQSGAAWIVKNRGRLEEKDA--CGNANECEAP-----IHGSNQYVGINNLYTPNDYVD  
.....  
-----YL..KP.WEGSV..YIETLSRKCVPRK.N...IHISL.DRFSICK.F.  
-----YL..KP.WEGSV..YIETLSRKCVPRK.N...IHISL.DRFSICK.F.  
-----Y...KP.WEGSV..YIETLSRKCVPRK.N...IHISL.DRFSICK.F.

660 670 680 690 700  
LSFGGRLDKQRIHSTDNIISKTYTNKSYNFGAAVHLTPDFSLLYK  
.....  
F.L...Y.RKNFTTSEELVR.GR.VDR.W.S.IVFKPNRH...S.R.  
F.L...Y.RQNFITSEELVR.GR..DR.W.S.IVFKPSRHL...S.R.  
F.L...Y.RKNFTTSEELVR.GR.ADR.W.S.IVFKPNRH..VS.R.

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4223  
Q8  
BNCV  
H44/76  
FA19

710 720 730 740 750 760  
TAKGFRTPSFYELYNYNSTAAQHKNPDVSPFKRAVDVKPETSINNEYGFYQHFWGDVEM  
.....I..  
ASS.....Q..FGIDIYH-----YPKGWQRPAL.S.KAANR.I.LQWKGDF.FL.I  
ASS.....Q..FGIDIYH-----YPKGWQRPAL.S.KAANR.I.LQWKGDF.FL.I  
ASS.....Q..FGIDIYH-----YPKGWQRPAL.S.KAANR.I.LQWKGDF.FL.I

770 780 790 800  
SMFKSRYKMDMLDKAIPNLTKAQ-QEYCKAHLDSNECVGNP  
.....  
.S.RN..T..IAV.DHK-..LPN.AGQLTEI.IRDYV---  
.S.RN..T..IAV.DHK-..LPN.AGRLTEI.IRDYV---  
.S.RN..T..IAV.DOK-..LPDSAGRLTEI.IRDYV---

4223  
Q8  
BNCV  
H44/76  
FA19

FIG.6E

810	820	830	840	850	860		
PTPKTSDEVFANLYNATIKGVSVKGKLDLHAMTSKLPDGLEMTILGYGHTKLGKFDYIAPKD							
.....	.....	.....	.....	.....	.....	.....	
-----	AQ.MSLQ..NIL..I.WNGVYG...E..YT..A.NRI..-----	.S					
-----	AQ.MSLQ..NIL..I.WNGVYG...E..YT..A.NRI..-----	.S					
-----	AQ.MSLQ.INIL..I.WNGVYG...E..YT..A.NRI..-----	.S					
		870	880	890	900		
	ADGMYQA--RPAFWDAITPARYVVGILNYDHPQSQVWGIGITTL					4223	
	.....--.....					Q8	
	VSNRPGLSL.SYAL..VQ.S...L.FG..Q.EGK..ANIM.						BNCV 97/130
	VSNRPDLSL.SYAL..CQ.S...L.FG..Q.EGK..ANIM.					H44/76	
	VSNRPDLSL.SYAL..VQ.S...L.FG..Q.EGK..ANIM.					FA19	
910	920	930	940	950	960		
THSKQKDENELSAIRNGKREIQTLTHTIPKAYTLLDMTGYSPTESITARLGINNVLNT							
.....	.....I.....						
.Y..G.NPD..-.YLAGDQ..YS---.KRASSWSTA.VSA.LNLKKRL.L.AA.Y.IG.Y							
.Y..G.NPD..-.YLAGDQ..YS---.KRASSWSTA.VSA.LNLKKRL.L.AA.Y.IG.Y							
.Y..G.NPD..-.YLAGDQ..YS---AGRVTSWKTA.VSA.LNLKKRL.L.AA.Y.IG.Y							
	970	980	990	1000			
	RYTIWEAARQLPSEAASTQST-----RYIAPGRSYFASLEMKF*					4223	
	.....	.....	.....	.....	.....	Q8	
	..V...SL..T-A.STANRHGDSNYG..A...NFSLA.....*					BNCV	
	..V...SL..T-A.STANRHGDSNYG..A...NFSLA.....*					H44/76	
	..V...SL..T-A.STANRHGDSNYG..A...NFSLA.....*					FA19	

FIG.7A

Alignment of M. catarrhalis Lbp2 proteins

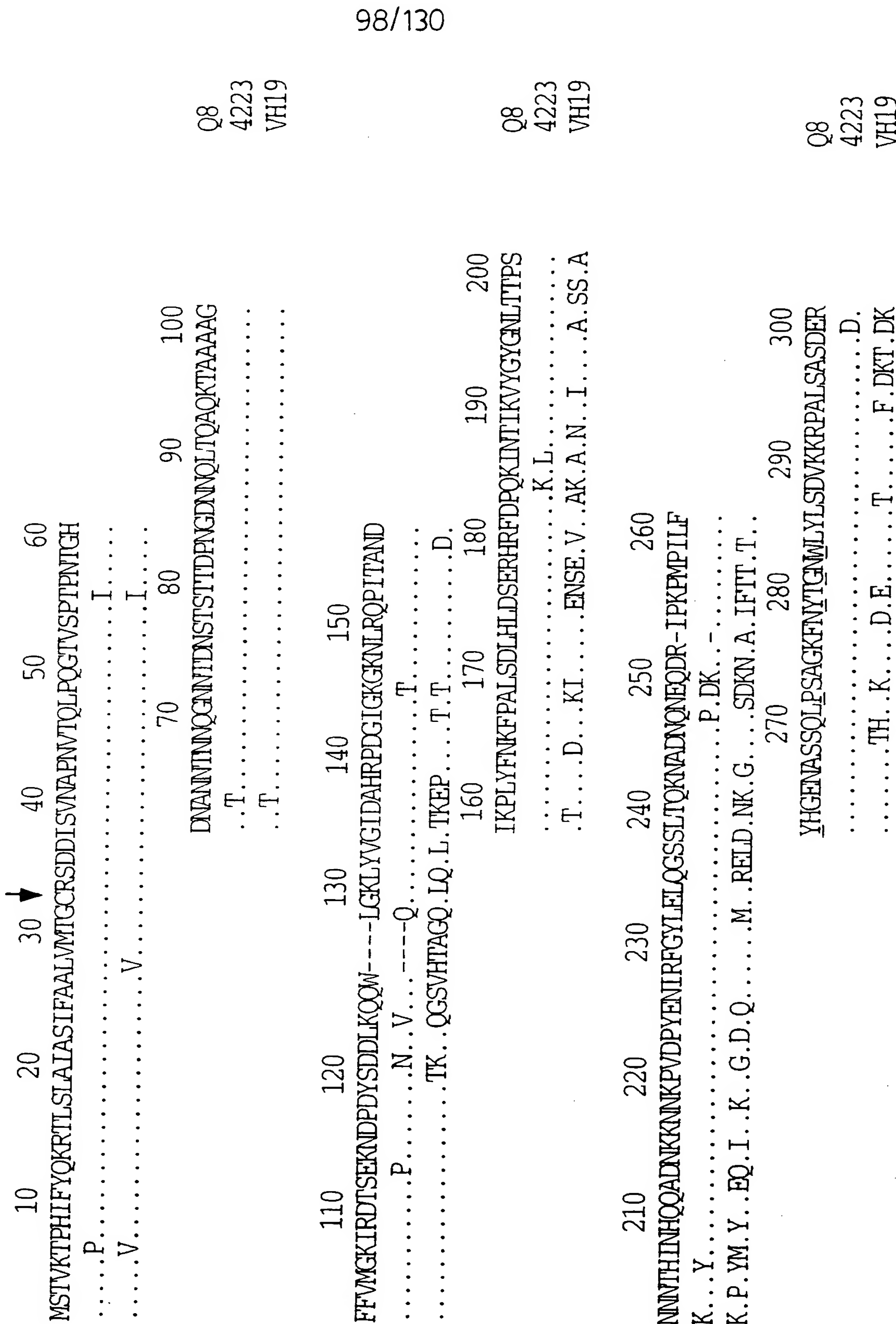




FIG.7B

310320330340350360  
VGVYLNASGKANEGDVWSAAHIYINGFYKHTPATYQVDFDINSLTGKLSYDNPNQNN  
.....S.....TA  
..T.F.STR.S...L.....S.K.....S...Q.T.K.....K.TA  
370380390400  
KGEYLKSFQDITTKKVNETDVYQIDAKINGNRFVGTAKSLV  
Q.K.I.....  
D.R.IR.....D....A...E.....T.....I  
Q8  
4223  
VH19

410420430440450460  
NEKTQTAPFIKELFSKKANPNPNPNSDTLEGGFYGESGDELAKFLSNDNASYVVFQGGK  
..N.E.....  
DDN.N....V.....D.....TF.....  
470480490500  
RDKTTPKPVATKTIVYFSAGFEKPSTSFVDNETIGGIIDRKG  
...D.....R..NS.K  
...E.....T.....G.E.S...G..  
Q8  
4223  
VH19

510520530540550  
LN----NHINEDEIIP-SDDSYGYTWGKPEKQFTKKVSSSTQVVPAYFGQHDKFYFNGN  
..DAVNEK.DNGD.PT-..ER.DEFP..EKKAE..  
..DEVN.Q.-...TV.V.NKE..E.NY.R.N.....INA.V.KN..  
560570580590600  
YYDLSASRVVDKLAPADAVKANQSIKEKYPNATLNKNQVTAIVLQ  
.....S.....  
.....KEAN..GVSQDTST.K..LA...D.KVST..K..K....  
Q8  
4223  
VH19

FIG.7C

610620630640650660

EAKDNKPYTAIRAKSYQHISFGETLYNDANQTPTRSYFVQGRADISTTLPOAGKFTYNG

.....K.....

Q...-...H...D...V...NKG...Q...V...Q...S.....

LK.I.TAEAD-I..T..AR.T.

670680690700

LWAGYLITQKKDKGYSDNAETIKEKCHPGYLLTENFTPEDD

.....I.....N.E....K...QD.....D.....

.....KDED...Q..LKD.I..KD.I.Q..

T.EARISKPIQWDMHADKKAA-----

710720730740750

DDD---LTASDDSQDDNTHGDDDLIASDDSQDDADGDDSDDLGCGADDDAAGKVYHA

-----DA.....A.....

...DDS.....T.....

-----

760770780790800

GNIRPEFENKYLPINEPTHEKITFALDCKNKAKFEVDENINSLTG

.....D...D.....

.....D.N.D.....

-----E.D...GEK.IS.

Q84223VH19H44/76

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Q84223VH19H44/76

FIG. 7D

810 820 830 840 850  
KLNDERGDIV-FDIKNGKIDGTGFTAKADVPNVREEV-GNNQGGFLYNIKDIDVKG  
.....  
.....

T.TEKN.VOPA.H.E..V.E.N..H.T.RTRDNGINLS..DSINPPSFKANNLL.T.  
RDNGINLS..GSTINPPSFKADNLL.T.

860 870 880

QFFGTNGEELACQLHDKGD--GINDTAEKA-----  
.....QY.....  
R.....T.....  
G.Y.PKA...G.IIFNND.KSL..TEGT.NKVE--ADVDDVDVDVD  
G.Y.PQA...G.IIFNND.KSL..TEDT.NEAE--AEVENEACVG--  
G.Y.PQAA..G.IIFNND.KSL..TEDI.NEVENEADV-----

101 / 130  
Q8  
4223  
VH19  
BNCV  
H44/76  
FA19

890  
-----GAVFGAVKD-----K\*  
-----\*  
-----\*  
ADADVEQLKPEVKPQF.V...K..NKEVE.\*  
-----EQLKPEAKPQF.V...K..NKEVE.\*  
-----EQLEPEVKPQF.V...K..NKEVE.\*

Q8  
4223  
VH19  
BNCV  
H44/76  
FA19

FIG.8A M. catarrhalis strain 4223 Tbp2/Lbp2 comparison



MKHIPLT--TLCVAISA---VLLTACGSGGS-NPPAPT-----PIPNASGSGNTG MSTVKTP..FYQKR..SL..ASIFAALVM.G.RSDDI.V.A.NV.QLPQGTVS...TGH-..N ** *	Tbp2 Lbp2
NTGNAGG-TDNTANAGNTGGTNSGTGSANTPEPKYQDVPTKNEKDKVSSIQEPAMG-----YGM ..N.Q.NN...STSTDPN.D.NQLTQ.QKTAAGFFVMG.I-R.TSPKNDPDYSNDLVQQWQG *	Tbp2 Lbp2
ALSKINLHNRQDTPLEKNIITLDGKKQVAEGKKSPLPFLDVENKLLDGYIAKMNVADKNAIGD K.YVGIDAH.P.GIGTG..LRQPITANDIKPLYFNKF.ALS.L-----HL.SERHRF *	Tbp2 Lbp2
RIKKGNKEISDEELAKQIKEAVRKSHEFQQVLSSLENKIFHSNDGTTKATTRDLKYVDYGYLAN DP..L.TIKVGYGNLTTPSKNNTYINH..ADNKN..PVDPYENIRFGYLELQGSLLTQKNADT *	102/130 Tbp2 Lbp2
DGNYLTVKTDKLWNLGPVGGVFYNGTTAKELPTQDAVKYKGHWDFMTDVANRRNRFSEVKENSQ PNDKDRIPK-----MPIL..H.ENASSQ..SAGKFN.T.N.LYLS..KK.PALSASDDRV-- *	Tbp2 Lbp2
AGWYYGASSKDEYNRLLTKEDSAPDGHSGEYCHSSEFTVNFKEKLTGKL-----F -V.LN..G.SNEGDVSAAHYLN.FQYKHTPAT-YQ.D.DTNS...SYYDNPNQQTAAQGY *	Tbp2 Lbp2
SNLQDRHKGNVTKTERYDIDANIHGNNRFRGSATA-----SNKNDTSKHPFTSDAN-----NR IKS.FDTTKK.NE.DV.Q...K.N...V.T.KSLVNENTETAPFI.EL.SKK..PNNPNPNSDT *	Tbp2 Lbp2

FIG.8B

LEGGFYGPKEELAGKFLTNDNKLFGVFGAKRESKAEEKTEAILDAYALGTFNTSNATFTTPTFTE Tbp2  
.....ES.D.....S...ASYV...G..DKTDKPVATKTVYFS.AFEKP----- Lbp2  
\*\*\*\*\* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \*

KQLDNFGNAKKLVLGSTVIDLVPTDATKNEFTKDKPESATNEAGETLMVNDEVSVKTYGKNFEYL Tbp2  
----- Lbp2

KFGELSIGGSHSVFLQGERTATTGEKAVPTTGTAHYLGWVGYITGKDTGTGTGKSFTDAQDVAD Tbp2  
----- Lbp2  
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FDIDFGNKSVSGKLITKGRQDPVFSITGQIAGNGWTGTASTTKADAGGYKIDSSSTGKSIKDA Tbp2  
----- Lbp2  
\* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \*

-----NVTGGFYGPAN-EMGGSFTHN-----ADDS-----KASVVFGTKRQQEVK \* Tbp2  
REEVGN.QG...LYNIKDIDVK.Q.FGTNGEEL.GQLQYDKGDGINDTAE..GA...AVKD---. \* Lbp2  
\* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \* \*

FIG.9A Alignment of *M. catarrhalis* Lbp3 sequences

10	20	30	40	50	4223
MTCLPKTNPALKVKHRFLKQVLLLCVDTLTAQAYAHSHHTPIHTPTHEL					Q8
.....					
60	70	80	90	100	4223
PSADALSDEGLGKDLGSLDSDSPDGLGDGLGDKSDKAPLPINA					Q8
S.....T.....					
110	120	130	140	150	4223
LTAHQTNESQPAPPSVDVNFLLAQPEAFYHFHQAIVQDDVATLRLLLPF					Q8
..VN.S.....					
160	170	180	190	200	4223
YDRLPDDYQDDVLLLFAQSKLALSDGNTKLALNLLTDLSNKEPTLTAVKL					Q8
.....					
210	220	230	240	250	4223
QLASLLLTNKHDKHAQMVLDELKDDAHFLKLSKKEQRWVLSQSRYLHKKY					Q8
.....					
260	270	280	290	300	4223
KMGDLGINYLHLDNINAASTITQPNIKKDAPKPAHGLALSLGVNKYTPL					Q8
.....					
310	320	330	340	350	4223
SHGMSIYTALDVGKFYDDKSHNELAVFAHAGLRKDHQKGYVDVVPFVGR					Q8
.....					



FIG.9B

360	370	380	390	400	
IFATNQQHGR	LSPRKDSQGV	AFGSHHR	INDKWQNA	FFARMEKGN	YTERYQ
.....	.....	.....	.....	.....	H..
410	420	430	440	450	
GYDGKRYHV	NDTILLQDGP	NRRLYSLG	VGYQLSHLQ	DATKSSHAT	KIHFGV
.....	.....	.....	.....	.....	.....
460	470	480	490	500	
LQRLPNGLT	VQGRVSAER	ERYHGKLL	RVLNPD	DDVYRTDK	TLTLQTSI
.....	.....	.....	.....	.....	.....
510	520	530	540		
DIHWLGLT	PKLT	RYSKNNS	NLPALY	SHNKQNF	YLELGRSF*
.....	.....	.....	.....	.....	.....*

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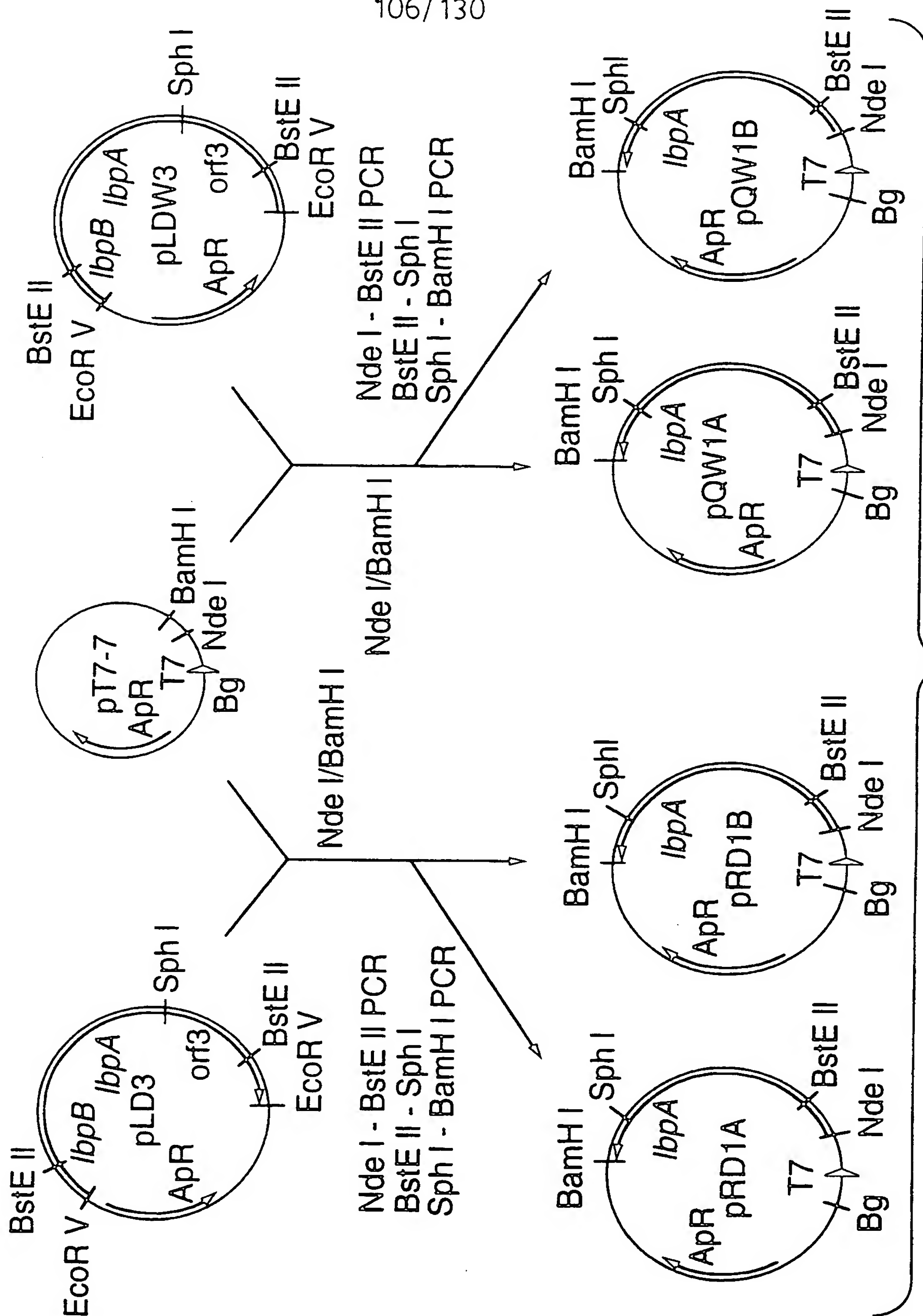


FIG.10



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## EXPRESSION OF Q8 AND 4223 rLbp 1 PROTEINS.

FIG. 11A.

Q8 rLbp1

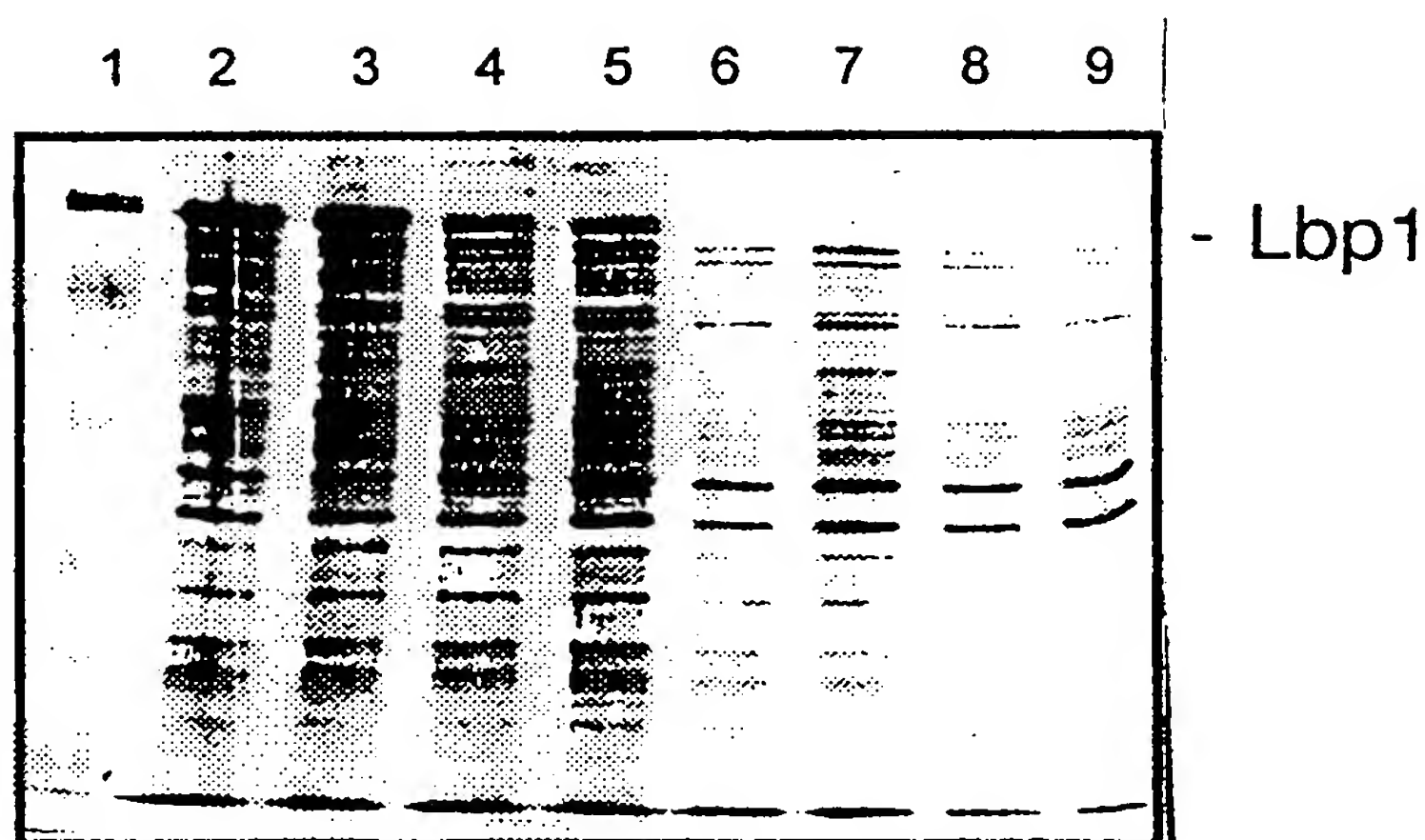
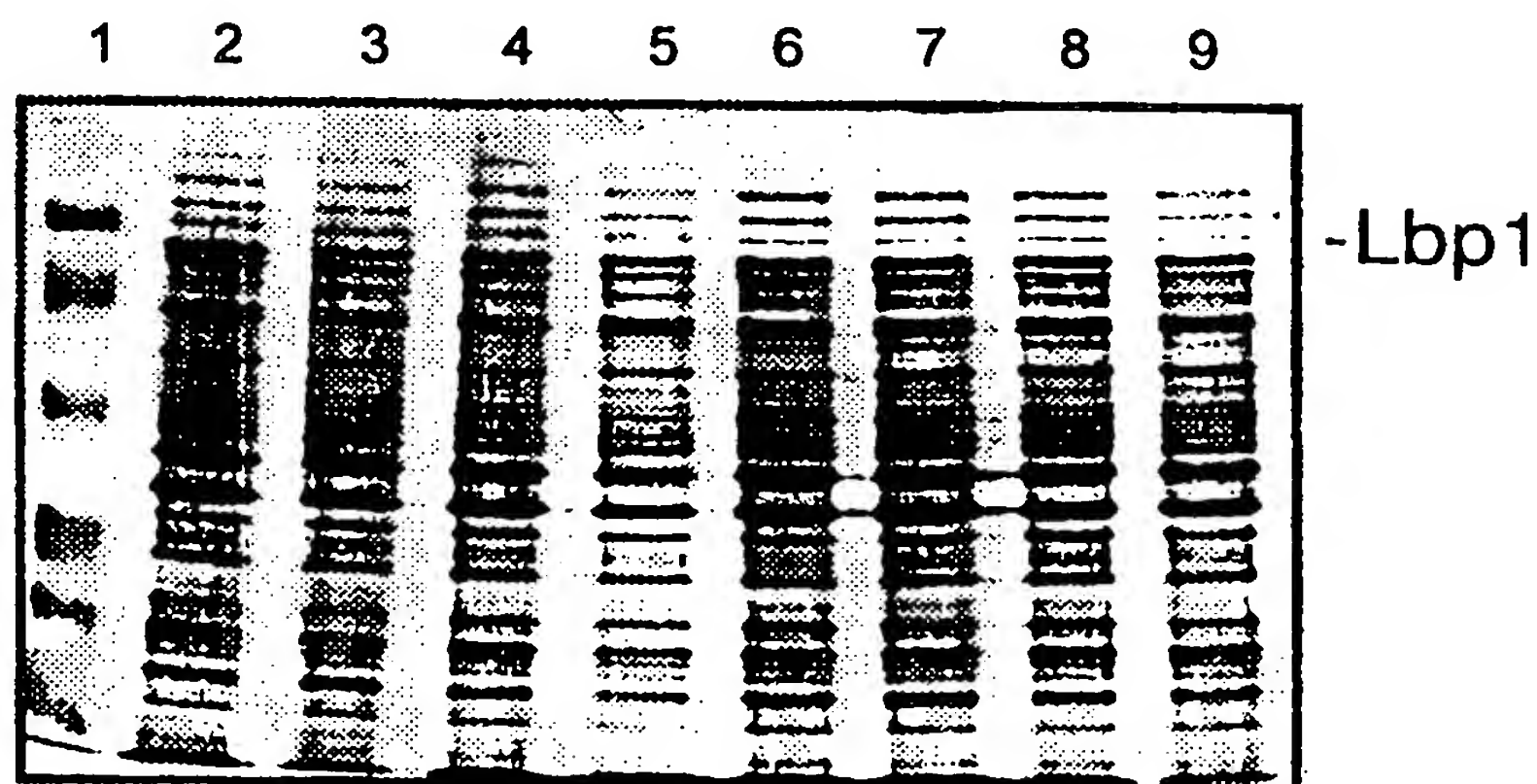


FIG.11B.

rLbp1



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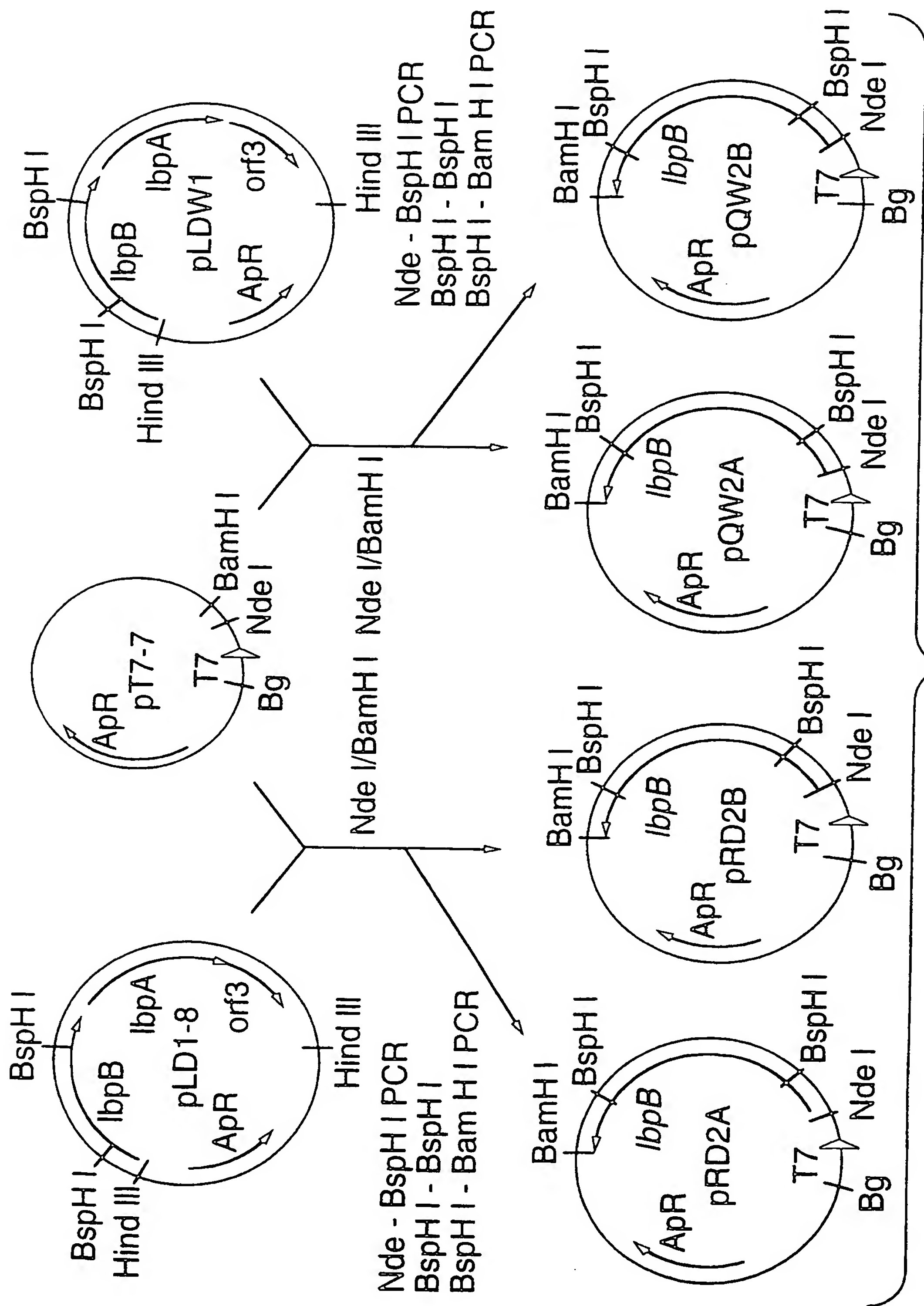


FIG.12

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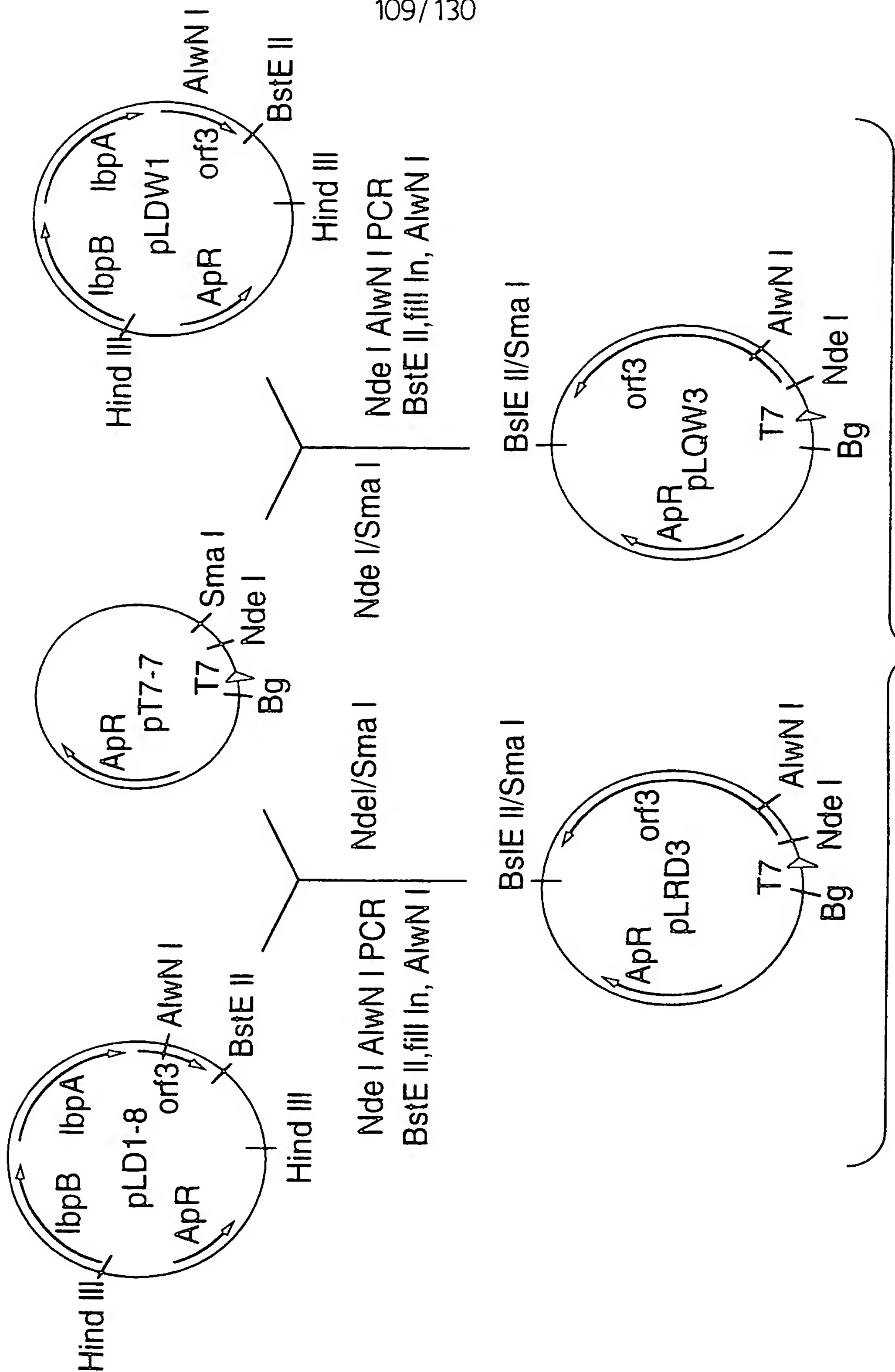
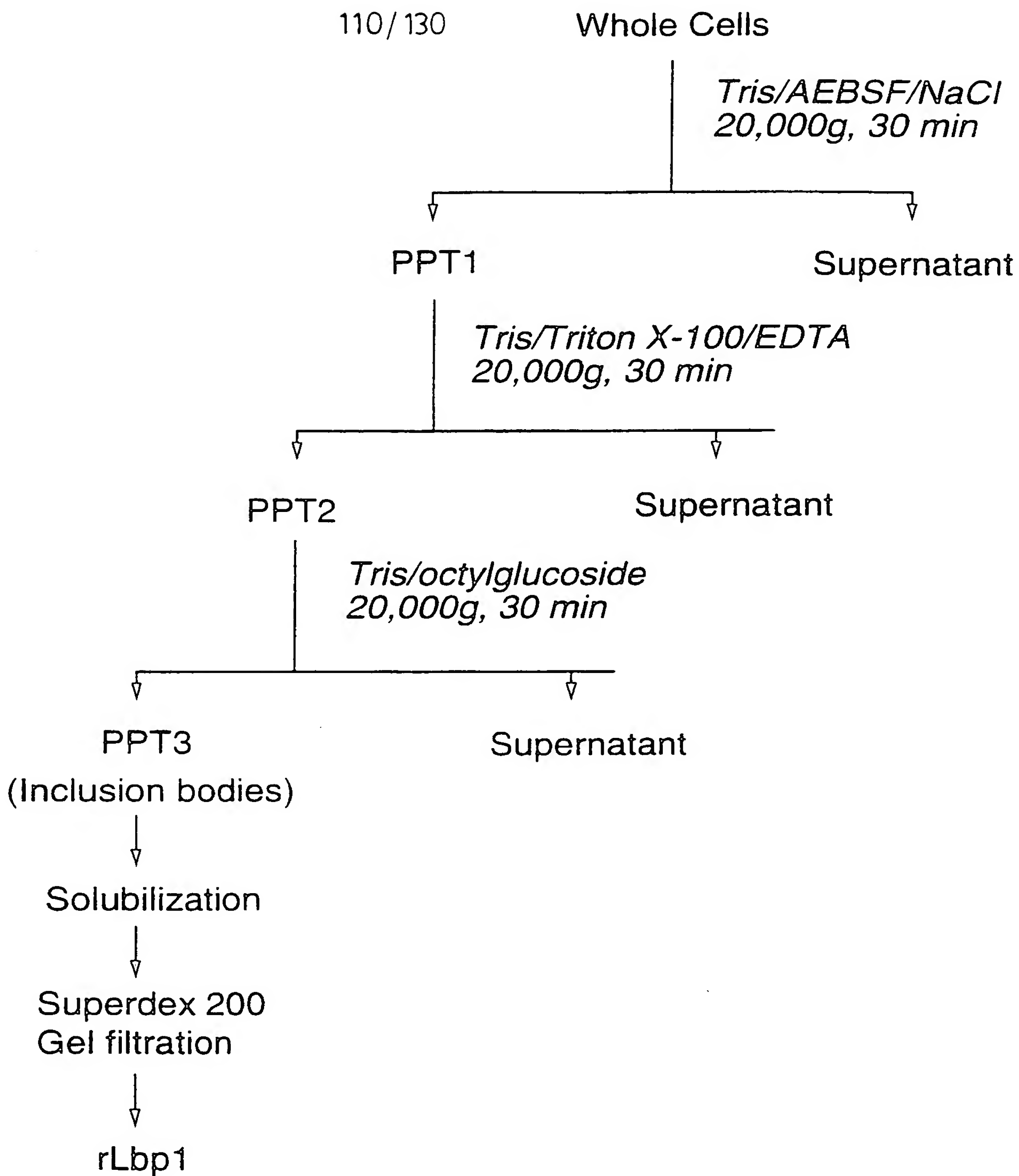


FIG.13

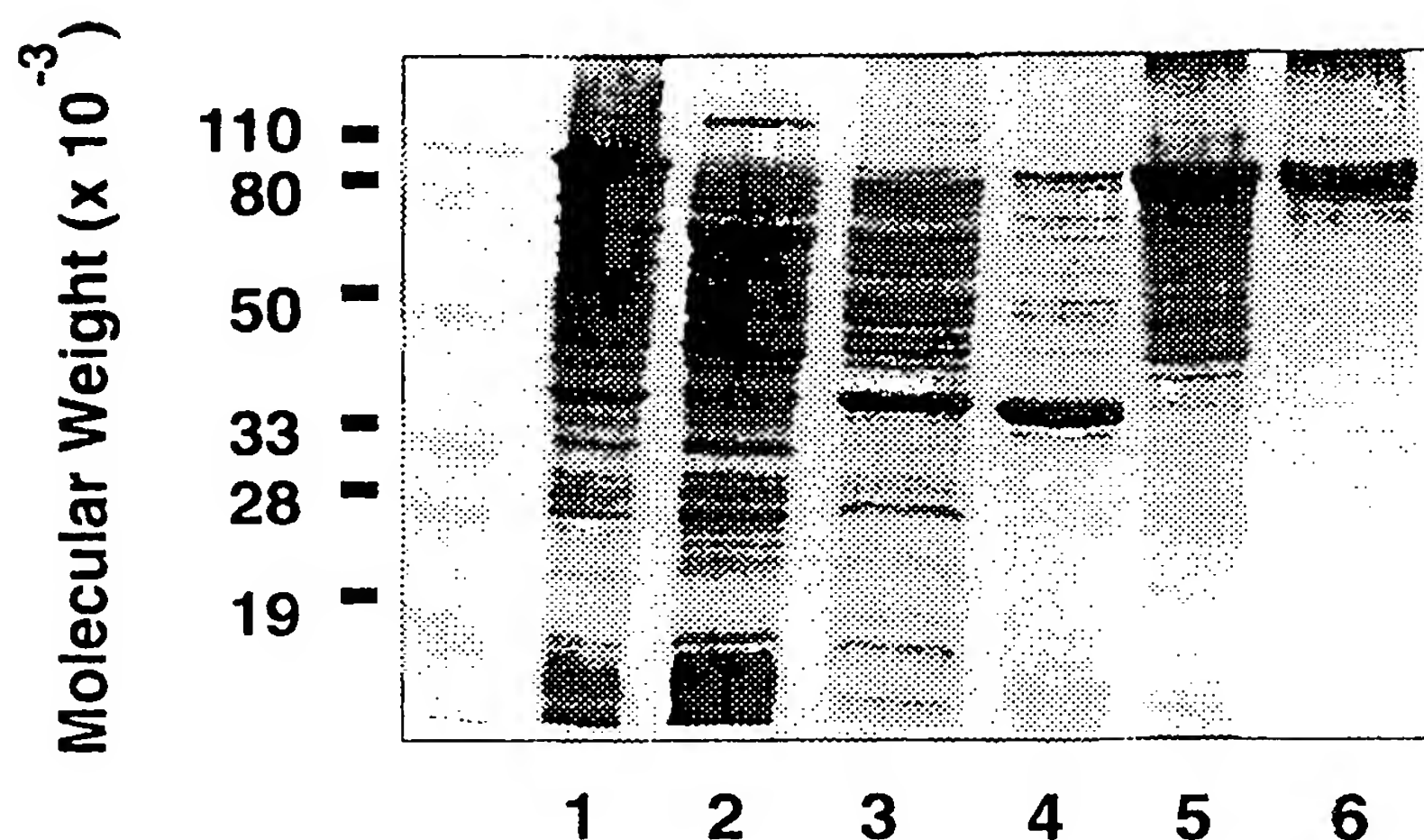


Purification scheme for rLbp1 expressed from E.coli

FIG.14

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## Purification of Q8 rLbp1 from *E. coli*



1. *E. coli* whole cells
2. Soluble proteins in 50 mM Tris/ NaCl extraction
3. Soluble proteins in Tris/ Triton X-100 extraction
4. Soluble proteins in Tris/ octylglucoside extraction
5. rLbp1 inclusion bodies
6. rLbp1

FIG.15.

Nucleotide and deduced amino acid sequence of the VH19 lbpB gene.

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```

MET  SER  THR  VAL  LYS  VAL  PRL  HIS  ILE  PHE
A T G A G T A C T G T C A A A G T C C C C A C A T T T C
10      20      30
      TYR  GLN  LYS  ARG  THR  LEU  SER  LEU  ALA  ILE
      T A C C A A A A C G C A C C C T T A G C C T T G C C A T C
      40      50      60

ALA  SER  ILE  PHE  ALA  ALA  VAL  VAL  MET  THR
G C C A G T A T T T T G C C G T G G T G A T G A C A
70      80      90
      GLY  CYS  ARG  SER  ASP  ILE  SER  VAL  ASN
      G G C T G C C G C T C T G A T G A C A T C A G C G T C A A T
      100     110     120

ALA  PRO  ASN  VAL  THR  GLN  LEU  PRO  GLN  GLY
G C A C C C A A T G T T A C C C A A C T G C C C C A A G G C
130     140     150
      THR  VAL  SER  PRO  ILE  PRO  ASN  THR  GLY  HIS
      A C G G T T T C A C C A A T A C C G A A C A C A G G T C A T
      160     170     180
```

FIG.16B

ASP ASN THR ASN THR ASN THR ASN GLN GLY  
G A C A C C A A T A A C A C C A A C A T C A G G G C  
190 200 210  
ASN ASN THR ASP ASP ASN SER THR THR THR  
A A C A A C A C G G A T A A C A G C A C C A C A A C T  
220 230 240

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ASP PRO ASN GLY ASP ASN ASN GLN LEU THR  
G A C C A A A T G G C G A T A A C A A C C A A C T G A C A  
250 260 270  
GLN ALA GLN LYS THR ALA ALA ALA GLY  
C A A G C A C A A A A A C T G C C G C C G C A G G G  
280 290 300

PHE PHE VAL MET GLY LYS ILE ARG ASP THR  
T T T T T G T G A T G G G T A A A T T C G T G A T A C C  
310 320 330  
SER GLU LYS ASN ASP PRO ASP TYR THR LYS  
A G C G A A A A A A T G A C C A G A T T A T A C C A A A  
340 350 360

FIG. 16C

ASP LEU GLN GLY SER VAL HIS THR ALA GLY  
 G A T T A C A A G G C A G C G T A C A T A C A G C A G G G  
 370 380 390  
 GLN GLY LEU GLN TYR LEU GLY THR LYS GLU  
 C A A G G C T T A C A G T A C T T A G G C A C C A A A G A G  
 400 410 420

PRO ARG PRO ASP GLY THR GLY LYS  
 C C T C G G C C A G A T G G C A C A G G T A C A G G T A A A  
 430 440 450  
 ASN LEU ARG GLN PRO ILE THR ALA ASP ASP  
 A A C T T A C G C C A G C C C A T C A C A G C T G A T G A C  
 460 470 480

ILE THR PRO LEU TYR PHE ASP LYS PHE PRO  
 A T T A C A C C A C T T T A T T T G A T A A T T C C C C  
 490 500 510  
 LYS ILE SER ASP LEU HIS LEU GLU ALS SER  
 A A A T A T C C G A T C T G C A C C T A G A A A C A G C  
 520 530 540

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SER SER PRO ALA LYS ASN PRO THR TYR MET  
T C A T C A C C T G C C A A A C C C A C C T A C A T G

610                  620                  630

ASN TYR GLN GLU GLN ASN ILE LYS ASN  
A A T A T C A C A G A C A A A C A T C A A A A C

640                  650                  660

[illegible]

FIG.16E

ASP LEU ASN LYS LYS GLY ALA ASP THR GLN  
 G A C C T A A A T A A A A A G G T G C A G A C A C C C A G  
 730 740 750  
 SER ASP LYS ASN ARG ALA ILE PHE THR  
 A G C G A C A A G A A C C G T G C C A T C A T T T C A C C  
 760 770 780

THR PRO THR LEU PHE TYR HIS GLY GLN ASN  
 A C A C C T A C T T A T T T A T C A T G G T G A G A A T  
 790 800 810  
 ALA SER THR HIS LEU PRO LYS ALA GLY LYS  
 G C C A G C A C C C A T C T G C C A A A G G C G G T A A A  
 820 830 840

PHE THR ASP ASP LYS VAL GLY THR TYR PHE  
 T T T G A C T A T G A G G C A A T T G G T T G T A T C T G  
 850 860 870  
 ASN SER THR ARG LYS SER ASN GLU GLY ASP  
 A C C G A T G T C A A A A A C G C C C A T T T T A G A T  
 880 890 900

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## FIG.16F

LYS THR ASP ASP LYS VAL GLY THR THR PHE  
 A A A C A G A C G A T A A A G T A G G C A C T T A T T T  
 910 920 930  
 ASN SER THR ARG LYS SER ASN GLU GLY ASP  
 A A C T C A A C C A G A A A A T C A A A T G A A G G C G A T  
 940 950 960

LEU VAL SER ALA ALA HIS ILE TYR LEU ASN  
 T T G G T G A G T G C A G C A C A T T T A T C T A A A C  
 970 980 990  
 SER PHE LYS TYR LYS HIS THR PRO ALA THR  
 A G C T T T A A A T A T A A C A C C C G C C A C T  
 1000 1010 1020

TYR SER VAL ASP PHE ASP GLN ASN THR LEU  
 T A T A G C G T G G A C T T T G A T C A A A T A C C C T A  
 1030 1040 1050  
 LYS GLY LYS LYS LEU SER TYR TYR ASP ASN PRO  
 A A A G G C A A A T G T C T T A T T A T G A C A C C C A  
 1060 1070 1080

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Fig. 166

ASN	LYS	GLN	THR	ALA	ASP	GLY	ARG	TYR	ILE
AACAGCAACAAGCCGATGGGCCTATTATC									
1090						1100			1110
	ARG	SER	GLN	PHE	ASP	THR	ASP	LYS	VAL
	AAGAATCAGTTTGATACCGAACAAAGGTC								
	1120					1130			1140
ASN	GLU	ALA	ASP	VAL	TYR	GLU	ILE	ASP	ALA
AATGAAGCCGATGTCTATGAGATTGACGCC									
1150						1160			1170
	LYS	ILE	ASN	GLY	ASN	ARG	PHE	THR	GLY
	AAGATTATA TG G C A A C C G C T T A C T G G C A C A								
	1180					1190			1200
ALA	LYS	SER	LEU	ILE	ASP	ASP	ASN	THR	ASN
GCCAAATCTTGTGATTGTGATTAACACCATT									
1210						1220			1230
	THR	ALA	PRO	PHE	VAL	LYS	GLU	LEU	PHE
	ACCGCACCTTTTGTTAAGAGCTGTTCTCC								
	1240					1250			1260

FIG.16H

LYS LYS ALA ASN PRO ASN PRO ASP PRO  
A A A A G C C A A T C C C A A C C C A G A C C C  
1270 1280 1290  
ASN SER ASP THR LEU GLU GLY GLY PHE TYR  
A A C T C A G A T A C G C T A G A A G G C G G G T T T A T  
1300 1310 1320  
119/130  
GLY GLU SER GLY ASP GLU LEU ALA GLY LYS  
G G T G A G T C G G G C G A T G A G C T G G C G G T A A A  
1330 1340 1350  
PHE LEU SER ASN ASP ASN ALA THR PHE VAL  
T T T T A T C C A A T G A C A A C G C A A C T T T G T G  
1360 1370 1380  
VAL PHE GLY GLY LYS ARG ASP LYS THR THR  
G T C T T G G T G G C A A A C G A G A C A A A C G A C C  
1390 1400 1410  
GLU PRO VAL ALA THR LYS THR VAL THR PHE  
G A A C C T G T C G C C A C A A A A C G G T G T A T T T  
1420 1430 1440

FIG.16I

SER THR GLY PHE GLU LYS POE SER THR SER  
 AGTACAGGATTGTGA A A A C C C A G C A C C A G C  
 1450 1460 1470  
 PHE VAL GLY ASN GLU GLU ILE GLY SER ILE  
 TTTGTTGGCAATGAGAGATTGGTAGCATTT  
 1480 1490 1500  
 ILE ASP GLY LYS LYS LEU ASN ASP GLU VAL  
 ATTGACGGTAA A A A G T T A A T G A T G A A G T C  
 1510 1520 1530  
 ASN ASN GLN ILE GLU ASP GLU THR VAL PRO  
 AATAATCA A A T T G A A G A T G A A C T G T C C C T  
 1540 1550 1560  
 VAL SER ASN LYS LYS GLU TYR TYR GLU TYR ASN  
 GTCAGTAAATAAGAA T A T T A T G A A T A T A A T  
 1570 1580 1590  
 TYR GLY ARG PRO ASN LYS GLN PHE THR LYS  
 TATGGACGACCC A A C A A A C A A T T C A C C A A A  
 1600 1610 1620

FIG. 16J

LYS ILE ASN ALA SER VAL GLN LYS ASN PRO  
 A A A T A A C G C C A G C G T C C A A A A A C C C T  
 1630 1640 1650  
 ALA TYR PHE GLY GLN HIS ASP LYS PHE TYR  
 G C T T A T T T G G T C A G C A T G A T A G T T T A T  
 1660 1670 1680

PHE ASN GLY ASN TYR TYR ASP LEU SER ALA  
 T T T A A T G G T A A C T A T T A T G A C T T A T C A G C C  
 1690 1700 1710  
 LYS GLU ALA ASN LYS LEU GLY VAL THR ASP  
 A A A G A G C C A A A C A A G C T T G G T G T C T C C C A A  
 1720 1730 1740

ASP THR SER SER THR ASN LYS SER ILE LEU ALA  
 G A T A C C A G C A C C A A T A A G A G T A T T T G G C T  
 1750 1760 1770  
 LYS TYR PRO ASP ALA LYS VAL SER THR ASP  
 A A A T A C C C A G A T G C C A A A G T A A G C A C A G A C  
 1780 1790 1800

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[illegible]



FIG.16L

ALA GLY LYS PHE THR TYR ASN GLY LEU TRP  
 G C A G G T A A A T T C A C C T A T A A T G G T C T T T G G  
 1990 2000 2010  
 ALA GLY TYR LEU THR GLN LYS ASP LYS  
 G C A G G C T A C C T G A C C C A G A A A A G A C A A A  
 2020 2030 2040

GLY TYR SER LYS ASP GLU ASP THR ILE LYS  
 G G T A T A G C A A A G A T G A G A T A C C A T C A A G  
 2050 2060 2070  
 GLN LYS GLY LEU LYS ASP TYR ILE LEU THR  
 C A A A A G G T C T T A A A G A T T A T A T T G A C C  
 2080 2090 2100

LYS ASP PHE ILE PRO GLN ASP ASP ASP  
 A A G A C T T T A T C C C A C A G A T G A C G A T G A C  
 2110 2120 2130  
 ASP ASP ASP SER LEU THR ALA SER ASP ASP  
 G A T G A C G A T A G T T T G A C C G C A T C T G A T G A T  
 2140 2150 2160

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## FIG.16M

SER GLN ASP ASP ASN THR HIS GLY ASP ASP  
T C A C A G A T G A T A A T A C A C A T G G C G A T G A T  
2170 2180 2190  
ASP LEU ILE ALA SER ASP ASP SER GLN ASP  
G A T T G A T T G C A T C T G A T G A T T C A C A G A T  
2200 2210 2220

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ASP ASP THR ASP ASP GLY ASP ASP SER ASP  
G A T G A C A C A G A T G G C G A T G A C G A T T C A G A T  
2230 2240 2250  
ASP LEU GLY ASP GLY ALA ASP ASP ASP ALS  
G A T T G G G T G A T G G T G C A G A T G A T G A C G C C  
2260 2270 2280

ALA GLY LYS VAL TYR HIS ALA GLY ASN ILE  
G C A G G C A A A G T G T A T C A T G C A G G T A A T A T T  
2290 2300 2310  
ARG PRO GLU PHE GLU ASN LYS TYR LEU PRO  
C G C C C T G A A T T T G A A A C A A A T A C T T G C C C  
2320 2330 2340

FIG.16N

ILE ASN GLU PRO THR HIS GLU LYS THR PHE  
A T T A A T G A G C C T A C T C A T G A A A A A C C T T T  
2350 2360 2370  
ALA LEU ASP GLY LYS ASN LYS ALA LYS PHE  
G C C C T A G A T G G T A A A A T A A G G C T A A G T T T  
2380 2390 2400

ASP VAL ASN PHE ASP THR ASN SER LEU THR  
G A T G T A A A C T T T G A C A C C A A C A G C C T A A C T  
2410 2420 2430  
GLY LYS LEU ASN ASP GLU ARG GLY ASP ILE  
G G T A A A T T A A A C G A T G A G A G G T G A T A T C  
2440 2450 2460

VAL PHE ASP ILE LYS ASN GLY LYS ILE ASP  
G T C T T G A T A T C A A A A T G G C A A A T T G A T  
2470 2480 2490  
GLY THR GLY PHE THR ALA LYS ALA ASP VAL  
G G C A C A G G A T T A C C G C C A A A G C C G A T G T G  
2500 2510 2520

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FIG. 16.O

PRO ASN TYR ARG GLU GLU VAL GLY ASN ASN  
 C C A A C T A T C G T G A A G A G T G G G T A A C A A C  
 2530 2540 2550  
 GLN GLY GLY GLY PHE LEU TYR ASN ILE LYS  
 C A A G G T G G C G G T T T C T T A T A C A A C A T C A A A  
 2560 2570 2580

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ASP ILE ASP VAL LYS GLY GLN PHE PHE GLY  
 G A T A T G A T G T T A A G G G C C A A T T T T T G G C  
 2590 2600 2610  
 THR ASN GLY GLU LEU ALA GLY ARG LEU  
 A C A A A T G G C G A A G A G T T G G C A G G A C G G T T A  
 2620 2630 2640

HIS HIS ASP LYS GLY ASP GLY ILE THR ASP  
 C A T C A T G A C A A A G G C G A T G G C A T C A C T G A C  
 2650 2660 2670  
 THR ALA GLU LYS ALA GLY ALA VAL PHE GLY  
 A C C G C C G A A A A G C A G G G C T G T C T T T G G G  
 2680 2690 2700

ALA VAL LYS ASP LYS \*\*\*  
 G C T G T T A A A G A T A A T A A  
 2710

Partial restriction map of *M. catarrhalis* strain VH19 *lbpB*

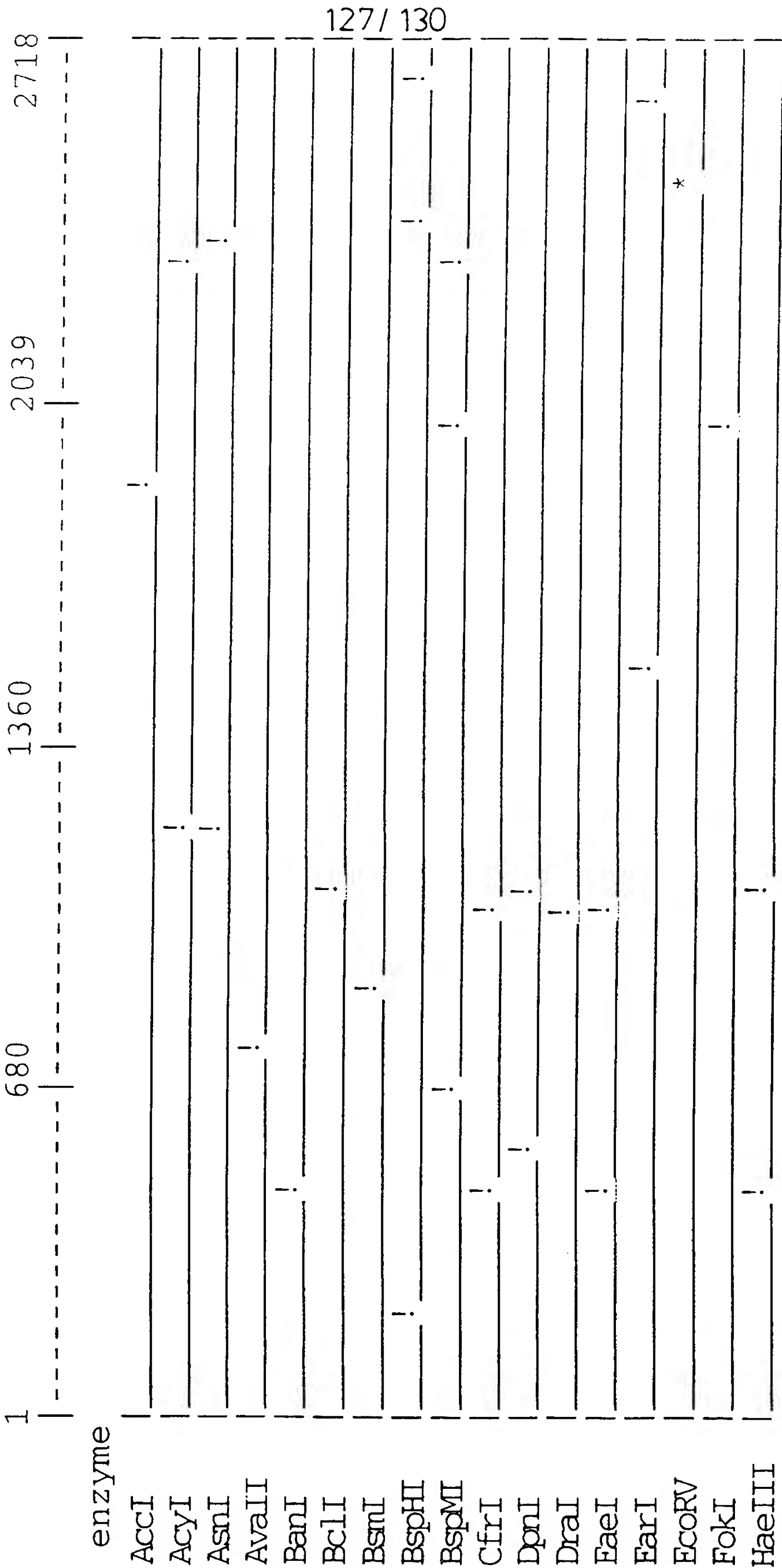


FIG.17A

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HindIII	!
HpaII	!
MaeI	!
MboI	!
MnlI	!
NciI	!
NspBII	!
NspI	!
PaiI	!
PvuII	!
ScaI	!
SspI	!
StyI	!
XbaI	!

FIG.17B

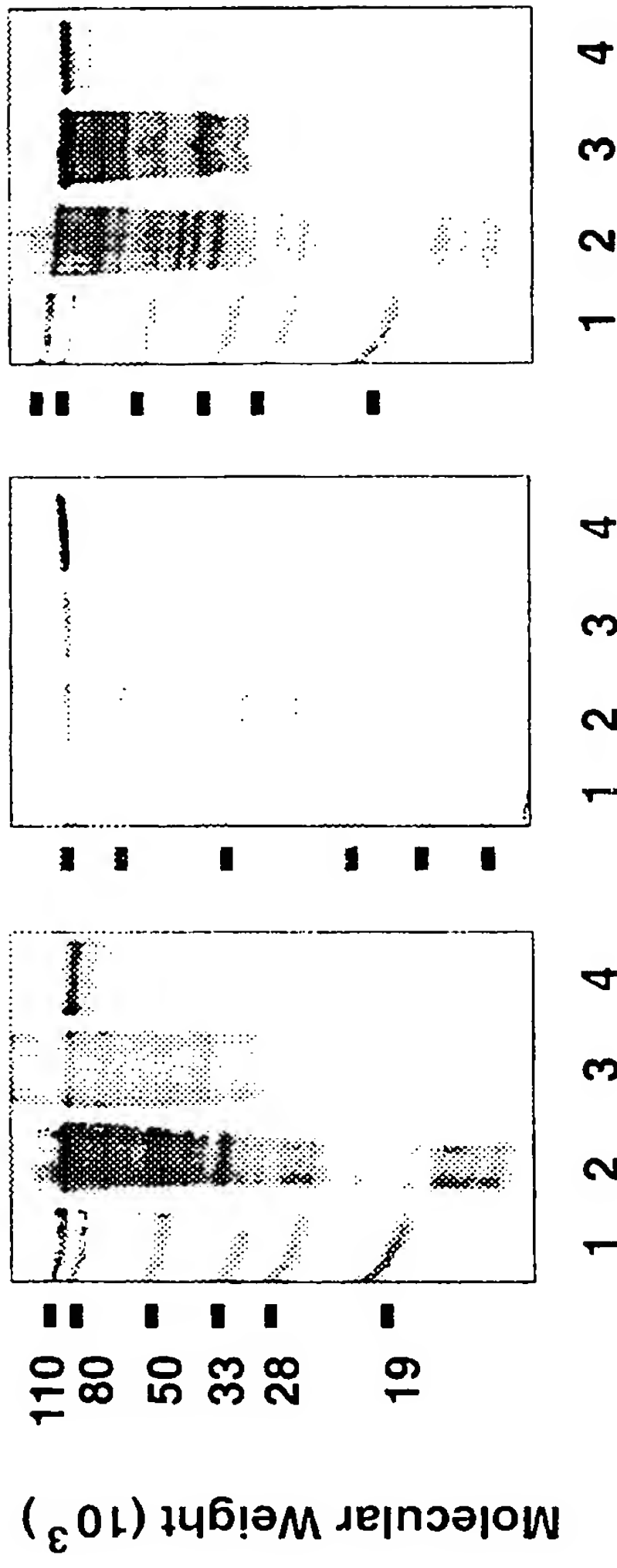


FIG. 18A.

FIG. 18B.

FIG. 18C.

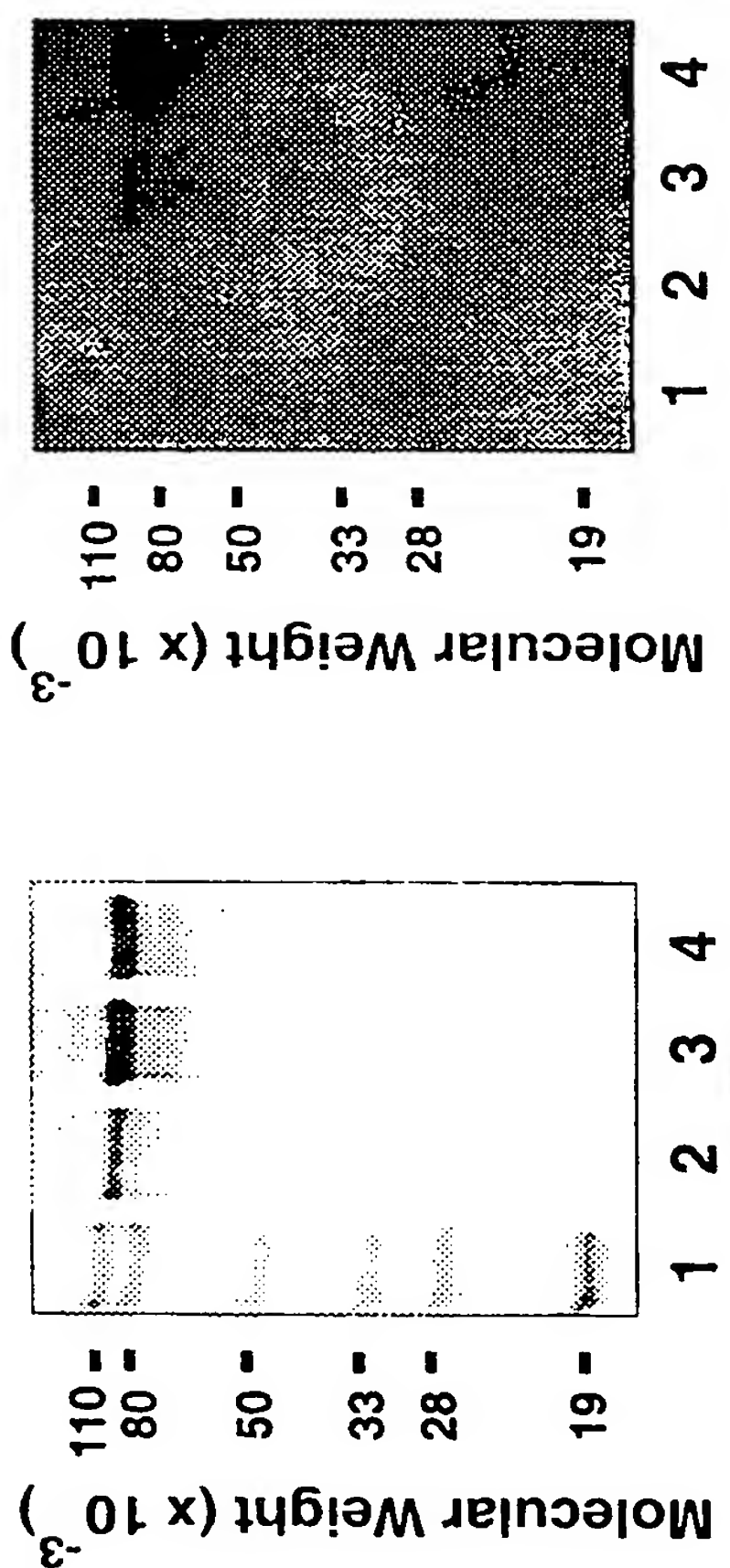


FIG. 19A.

FIG. 19B.

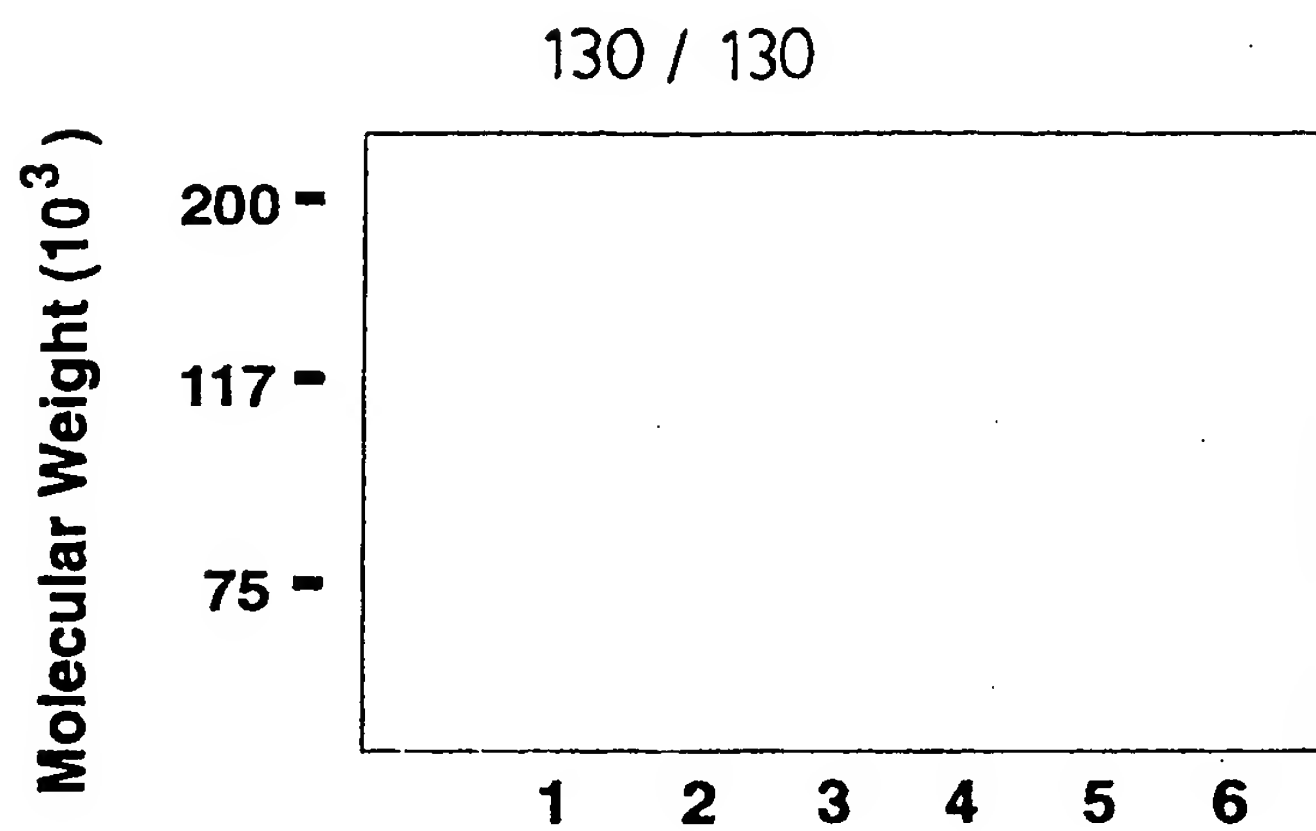


FIG.20A.

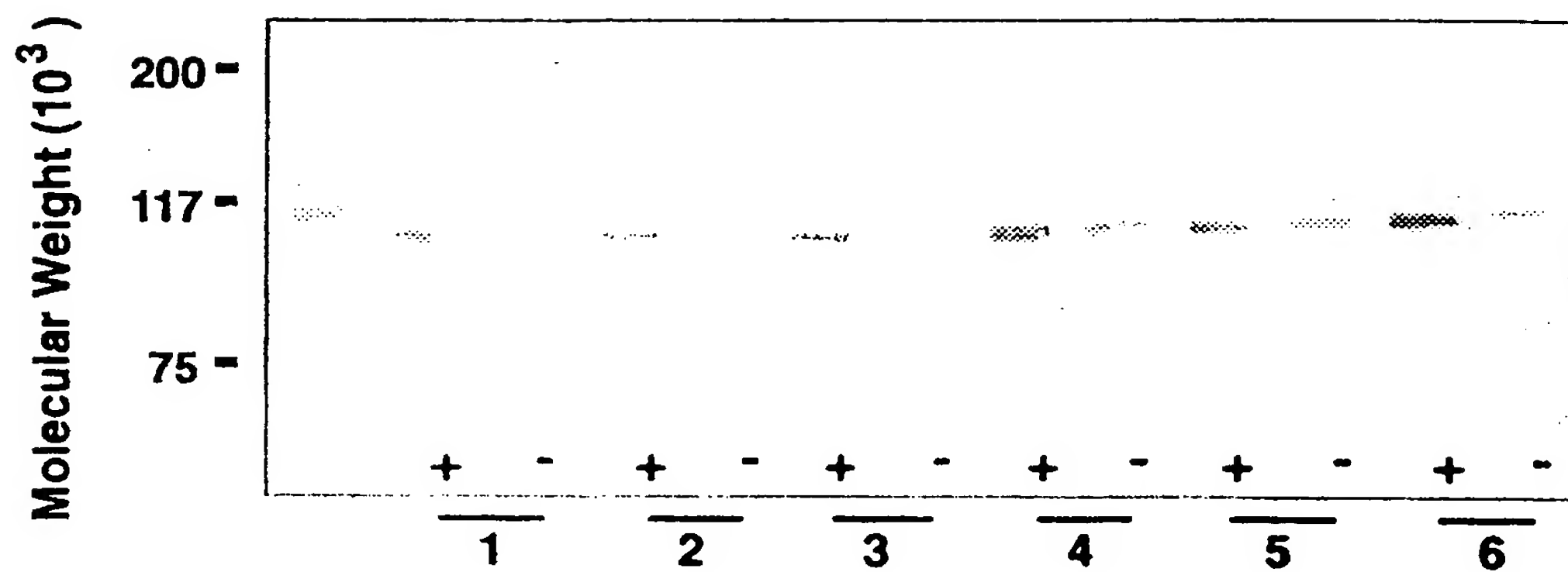


FIG.20B.

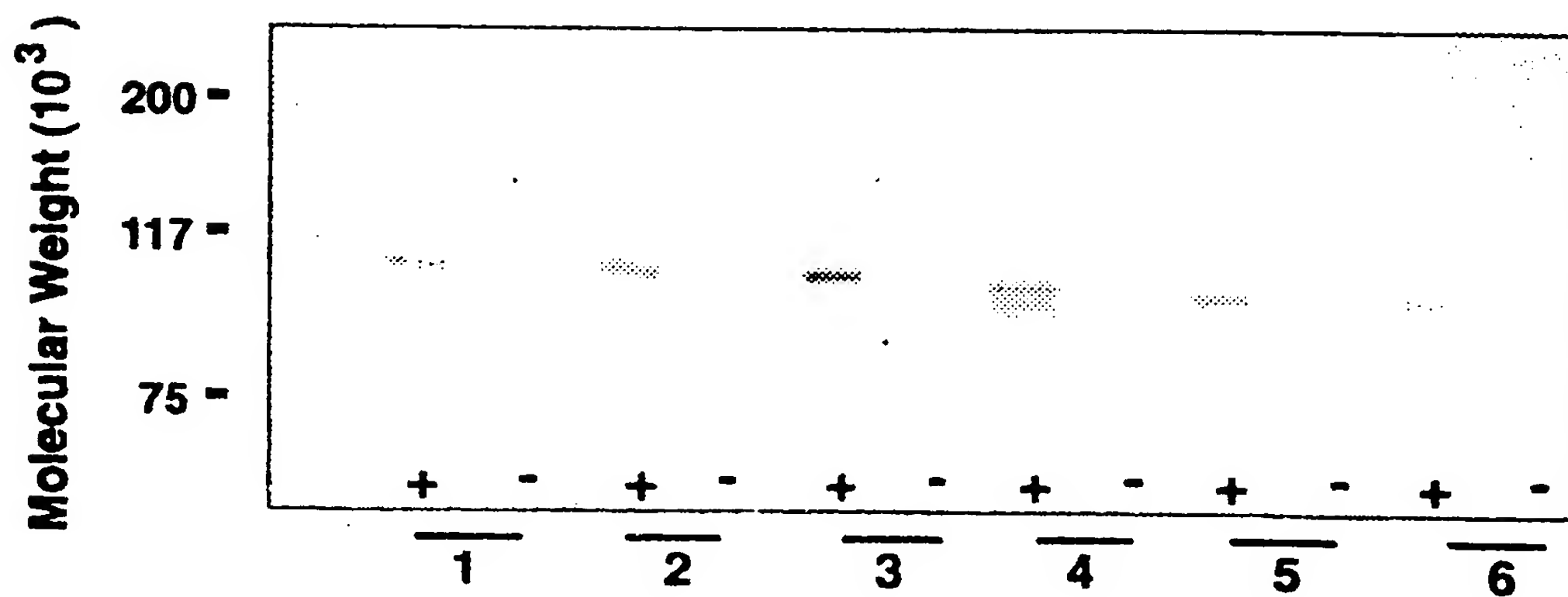


FIG.20C.





## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>6</sup> : <b>C12N 15/12, C07K 14/79, 14/22, A61K 38/17, G01N 33/68, C12Q 1/68</b>		<b>A3</b>	(11) International Publication Number: <b>WO 98/55606</b> (43) International Publication Date: 10 December 1998 (10.12.98)
(21) International Application Number: PCT/CA98/00544 (22) International Filing Date: 2 June 1998 (02.06.98) (30) Priority Data: 08/867,941 3 June 1997 (03.06.97) US 09/074,658 8 May 1998 (08.05.98) US (71) Applicant (for all designated States except US): CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Av- enue West, North York, Ontario M2R 3T4 (CA). (72) Inventors; and (75) Inventors/Applicants (for US only): LOOSMORE, Sheena, M. [CA/CA]; 70 Crawford Rose Drive, Aurora, Ontario L4G 4R4 (CA). DU, Run-Pan [CA/CA]; (CA). WANG, Quijun [CA/CA]; 299 Chelwood Drive, Thornhill, Ontario L4J 7Y8 (CA). YANG, Yan-Ping [CA/CA]; Apartment 709, 120 Torresdale Avenue, Willowdale, Ontario M2R 3N7 (CA). KLEIN, Michel, H. [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA). (74) Agent: STEWART, Michael, I.; Sim & McBurney, 6th floor, 330 University Avenue, Toronto, Ontario M5G 1R7 (CA).			(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> (88) Date of publication of the international search report: 4 March 1999 (04.03.99)
(54) Title: LACTOFERRIN RECEPTOR GENES OF MORAXELLA			
(57) Abstract  Purified and isolated nucleic acid molecules are provided which encode lactoferrin receptor proteins of <i>Moraxella</i> , such as <i>M. catarrhalis</i> , or a fragment or an analog of the lactoferrin receptor protein. The nucleic acid sequence may be used to produce recombinant lactoferrin receptor proteins Lbp1, Lbp2 or ORF3 of the strain of <i>Moraxella</i> free of other proteins of the <i>Moraxella</i> strain for purposes of diagnostics and medical treatment. Furthermore, the nucleic acid molecule may be used in the diagnosis of infection.			

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# INTERNATIONAL SEARCH REPORT

Interi      nar Application No

PCT/CA 98/00544

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6    C12N15/12    C07K14/79    C07K14/22    A61K38/17    G01N33/68  
C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6    C07K    C12N    A61K    G01N    C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	CA 2 162 193 A (CONNAUGHT LAB) 3 May 1997	20-22, 24,29,30
Y	see the claims see abstract; figures 1-3; examples 1,4  see page 4, line 21 - page 7, line 15 ---	3,4, 6-19,23, 25-28
X	OGUNNARIWO J A AND SCHRYVERS A B: "Rapid identification of bacterial transferrin and lactoferrin receptor protein genes" JOURNAL OF BACTERIOLOGY, vol. 178, no. 24, December 1996, pages 7326-7328, XP002083300	1,2,5, 31,32
Y	see abstract; figure 1; table 1  see page 7327 - page 7328 ---	3,4, 6-19,23, 25-28

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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents :

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- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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Date of the actual completion of the international search

5 November 1998

Date of mailing of the international search report

23/12/1998

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## INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 98/00544

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 90 12591 A (UNIV TECHNOLOGIES INT ;SCHRYVERS ANTHONY BERNARD (CA)) 1 November 1990 see abstract; claims 5-8,16-26; figure 1; examples 1,2 see page 5, line 4 - page 6, line 26 see page 7, line 30 - page 8, line 1 see page 10, line 8 - page 11, line 8 ---	20,24,29
X	SCHRYVERS A B ET AL: "COMPARATIVE ANALYSIS OF THE TRANSFERRIN AND LACTOFERRIN BINDING PROTEINS IN THE FAMILY NEISSERIACEAE" CANADIAN JOURNAL OF MICROBIOLOGY, vol. 35, no. 5, May 1989, pages 409-415, XP002020995 cited in the application see abstract; figures 1,4-6 see page 410, paragraph 9 see page 414, paragraph 2 ---	20-22,24
T	DU R-P ET AL: "Cloning and expression of the Moraxella catarrhalis lactoferrin receptor genes" INFECTION AND IMMUNITY, vol. 66, no. 8, August 1998, pages 3656-3665, XP002083301 see the whole document -----	1-32

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/CA 98/ 00544

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:  
Remark: Although claim(s) 30  
is(are) directed to a method of treatment of the human/animal  
body, the search has been carried out and based on the alleged  
effects of the compound/composition.
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such  
an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please see additional sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all  
searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment  
of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report  
covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is  
restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1, 5-20, 24, 29-32 all partially and 2, 21.

Lbp1 polypeptide, corresponding nucleic acids, vectors containing said nucleic acid, host cells transformed with said vector, immunogenic composition containing said nucleic acid or protein, methods for detection and diagnostic kit involving said nucleic acid.

2. Claims: 1, 5-20, 24, 29-32 all partially and 3, 22.

Same as in invention 1 but comprising Lbp2.

3. Claims: 1, 5-20, 24, 29-32 all partially and 4, 23, 25-28.

Same as in invention 1 but comprising ORF3.

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/CA 98/00544

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
CA 2162193 A	03-05-1997	NONE	
WO 9012591 A	01-11-1990	US 5292869 A	08-03-1994
		AU 649950 B	09-06-1994
		AU 5526190 A	16-11-1990
		EP 0528787 A	03-03-1993
		JP 4506794 T	26-11-1992
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		US 5141743 A	25-08-1992

